



THE IMPLEMENTATION OF A SCREENING WORKFLOW FOR ION MOBILITY QUADRUPOLE TIME-OF-FLIGHT MASS SPECTROMETRIC ANALYSIS OF PFOS ISOMERS

Ken Rosnack

ken_rosnack@waters.com

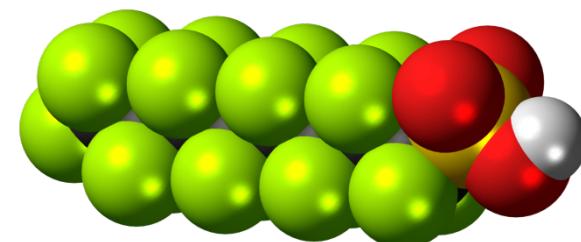
**Ingrid Ericson Jogsten and Bert van Bavel
(MTM Research Centre)**

**Mike McCullagh, Leonard Dillon, Mike Hodgkinson,
Lauren Mullin, & Jennifer Burgess
(Waters Corporation)**



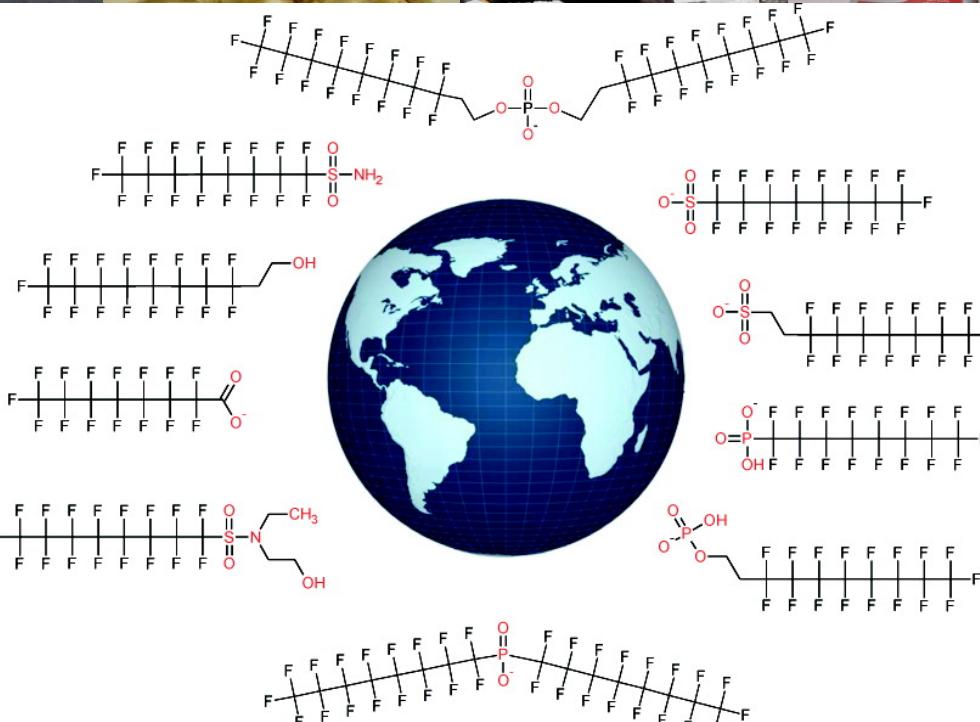
Overview

- PFOS analysis collaboration between Waters Corporation and Örebro University
 - Challenges of PFOS analysis
- Instrumentation: Synapt G2-S and HDMS^E
 - Ion mobility
- Analysis of environmental samples for PFOS using HDMS^E
 - Experimental
 - Results
 - Screening approach vs Software enabled identification (Development of prototype software)
- Conclusions



Introduction: Poly and perfluoroalkyl substances (PFASs)

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PFCs research

Perfluorochemicals in women ages 16 to 49 years: in blood serum, 1999-2008

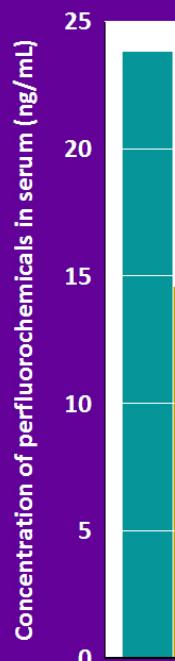


Figure 3: PFAAs composition profiles in different groups of sample matrices.

From

Elevated levels of perfluoroalkyl acids in family members of occupationally exposed workers: the importance of dust transfer

Jianjie Fu, Yan Gao, Thanh Wang, Yong Liang, Aiqian Zhang, Yawei Wang & Guibin Jiang
Scientific Reports | 5, Article number: 9313 | doi:10.1038/srep09313

Houde M, Martin JW, Letcher RJ, Solomon KR, Muir DC (June 2006). "Biological monitoring of polyfluoroalkyl substances: A review". *Environ. Sci. Technol.* **40** (11): 3463–73. doi: [10.1021/es052580b](https://doi.org/10.1021/es052580b). PMID [16786681](https://pubmed.ncbi.nlm.nih.gov/16786681/). Supporting Information(PDF).

Species	Geography	Year	Sample	PFOS (ppb)
Bald eagle	Midwestern USA	1990–93	plasma	2,200
Brandt's cormorant	California, USA	1997	liver	970
Guillemot	Baltic Sea	1997	egg	614
Carriion crow	Tokyo Bay, Japan	2000	liver	464
Red-throated loon	North Carolina, USA	1998	liver	861
Polar bear	Sanikiluaq, Nunavut	2002	liver	3,100
Harbor seal	Dutch Wadden Sea, Denmark	2002	muscle	2,725
Bottlenose dolphin	Charleston, South Carolina, USA	2003	plasma	1,315
Common dolphin	Mediterranean Sea, Italy	1998	liver	940
Mink	Michigan, USA	2000–01	liver	59,500

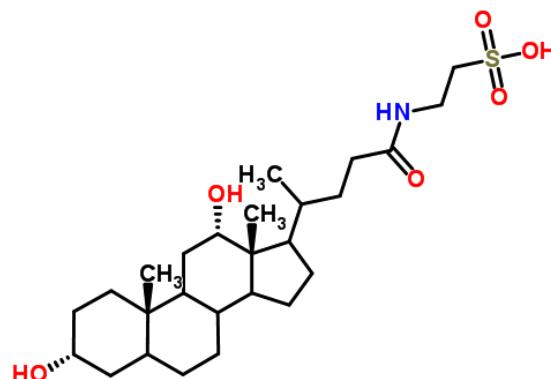
PFOS Analysis Challenges

- Matrix effects, retention time shifts.
- Correct PFOS isomer identification:
 - The physical, chemical and biological properties may be affected by perfluoromethyl branching.
 - Source elucidation.
 - Response factors of individual isomers.
- Increased scientific interest in toxicity, environmental transport, degradation and bioaccumulation of isomers.
- PFOS and TDCA as well as other cholic acids have similar isomeric profiles, retention times and MRM transitions (499 m/z → 80m/z).
- Interferences can be mistaken for PFOS and lead to a positive bias.

Chemical Structures PFOS and Cholic Acid Interference's

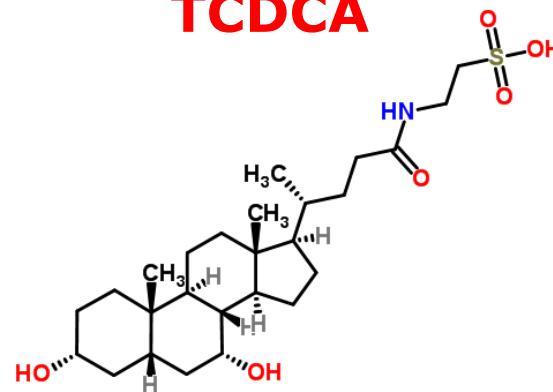
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TDCA



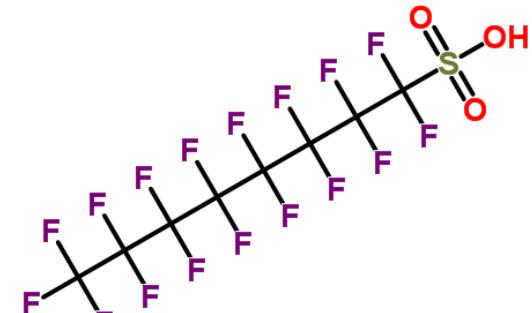
Taurodeoxycholate

TCDCA



Taurochenodeoxycholic acid

PFOS

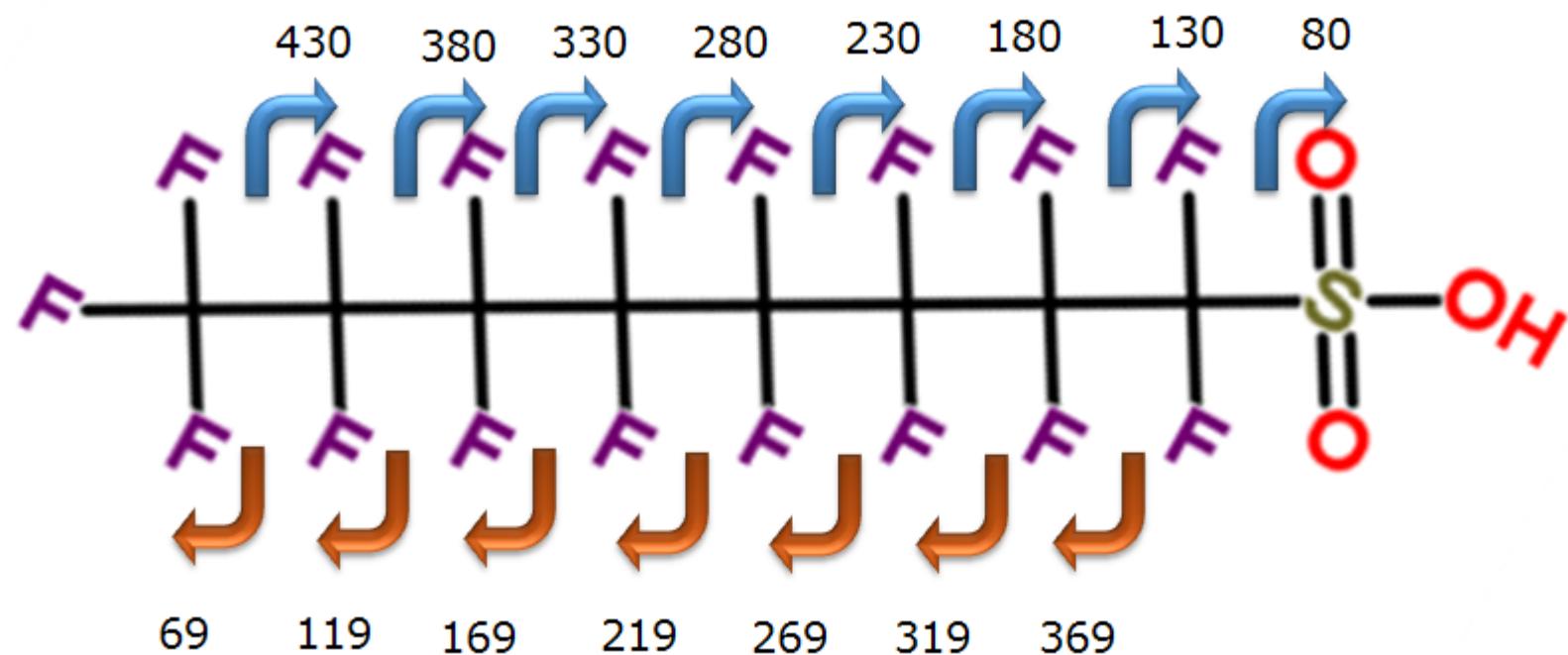


Perfluorooctane sulfonate

$$\text{C}_{26}\text{H}_{45}\text{NO}_6\text{S} = [\text{M}-\text{H}]^- = 498.2889$$

$$\text{C}_8\text{HF}_{17}\text{O}_3\text{S} = [\text{M}-\text{H}]^- = 498.9297$$

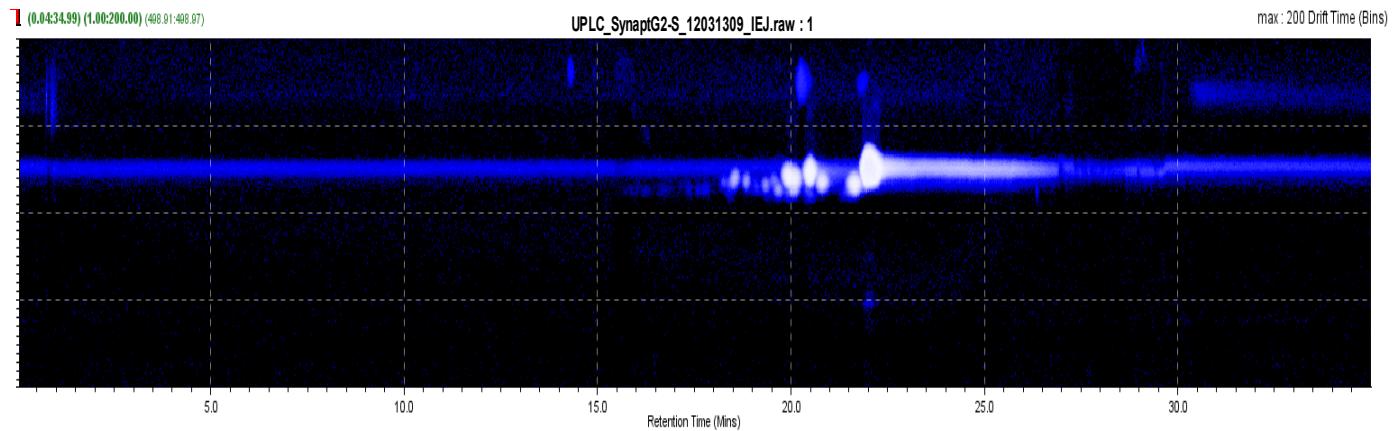
Fragmentation Series for PFOS



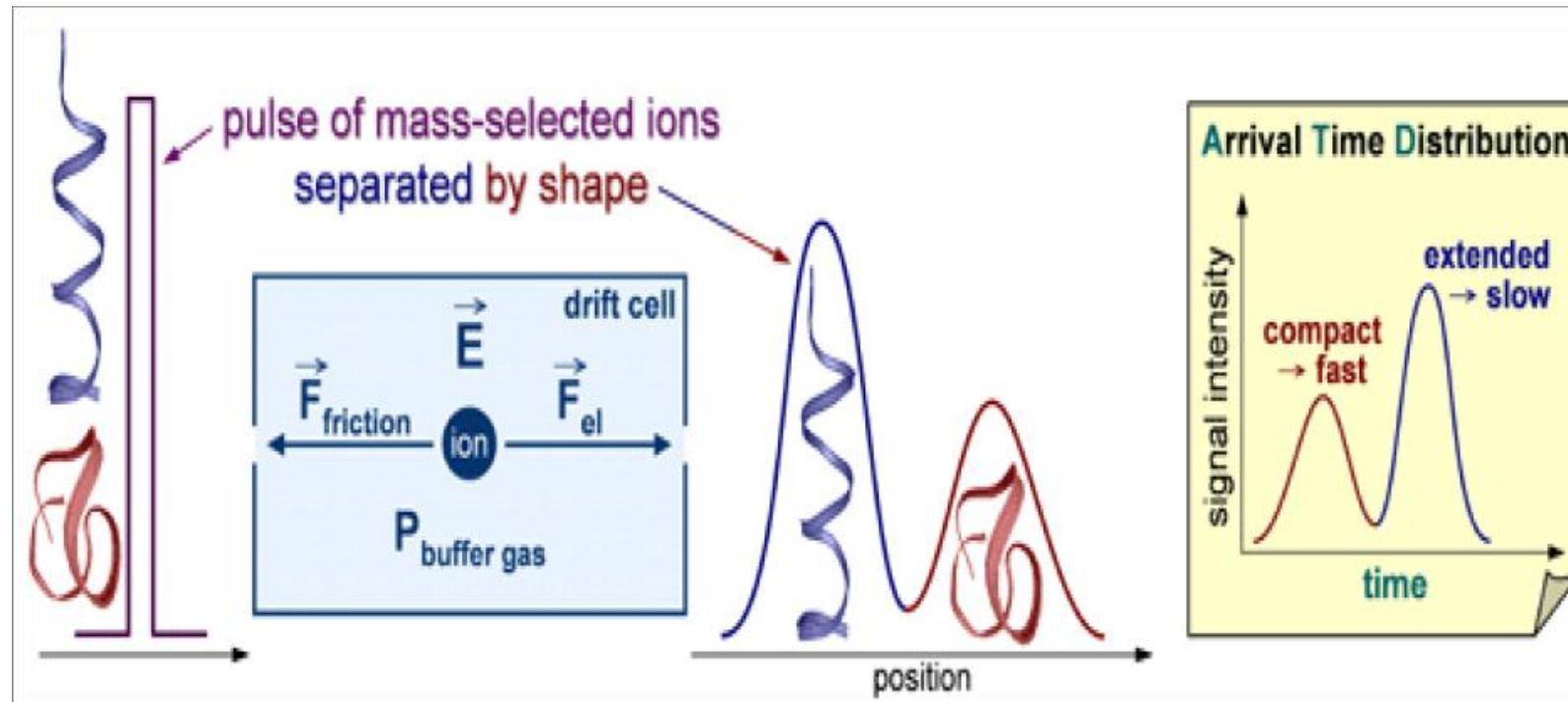
HDMS^E

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- Uses high resolution MS and high efficiency ion mobility based measurements and separations.
- Both precursor ion and fragment ion information can be acquired in a single HDMS^E experiment.
- This technique offers some unique advantages to profiling complex matrices.
- HDMS^E can provide a route to specific and unambiguous identification, enabling the distinction of PFOS isomers.



Ion Mobility Spectrometry



- Ion mobility spectrometry (IMS) is a rapid, orthogonal, gas phase separation technique which allows another dimension of separation.
- Separation is driven by electric fields not under vacuum.
- Compounds can be differentiated based on size, shape and charge.

Ion Mobility Spectrometry (IMS)

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**Small
Compact**



**Large
Extended**

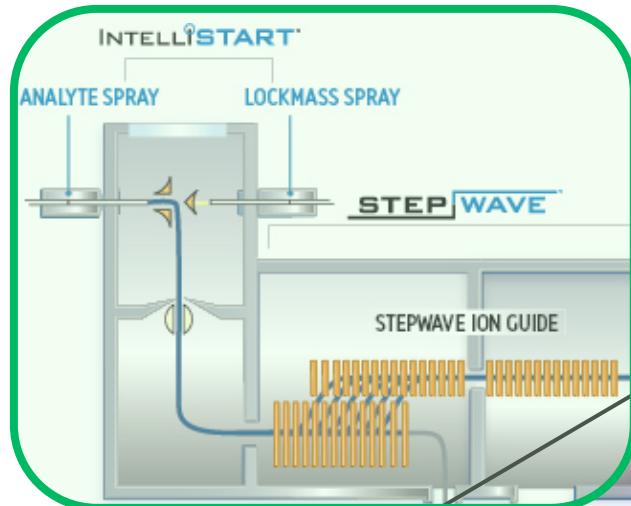
Slide courtesy of Severine Goscinny, ISP-WIV, Belgium
©2015 Waters Corporation

SYNAPT G2-S High Definition MS

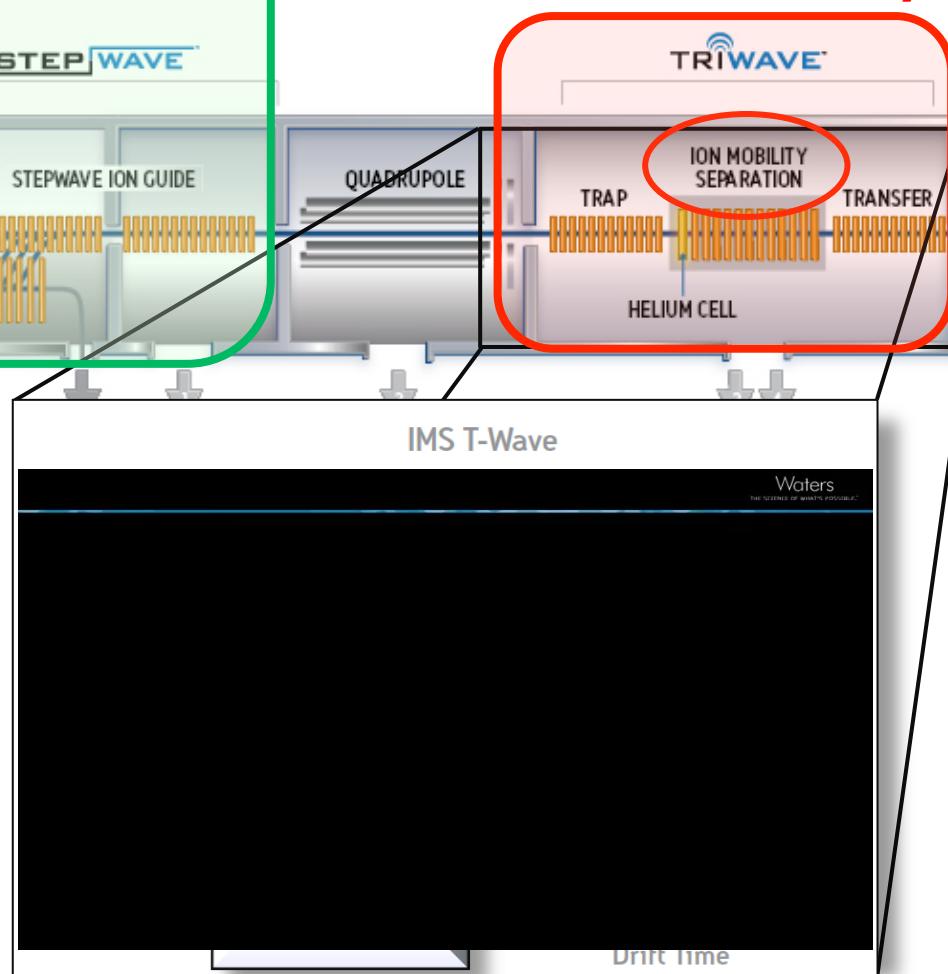
- Ion Mobility Explained

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1. Increased sensitivity



2. Ion mobility

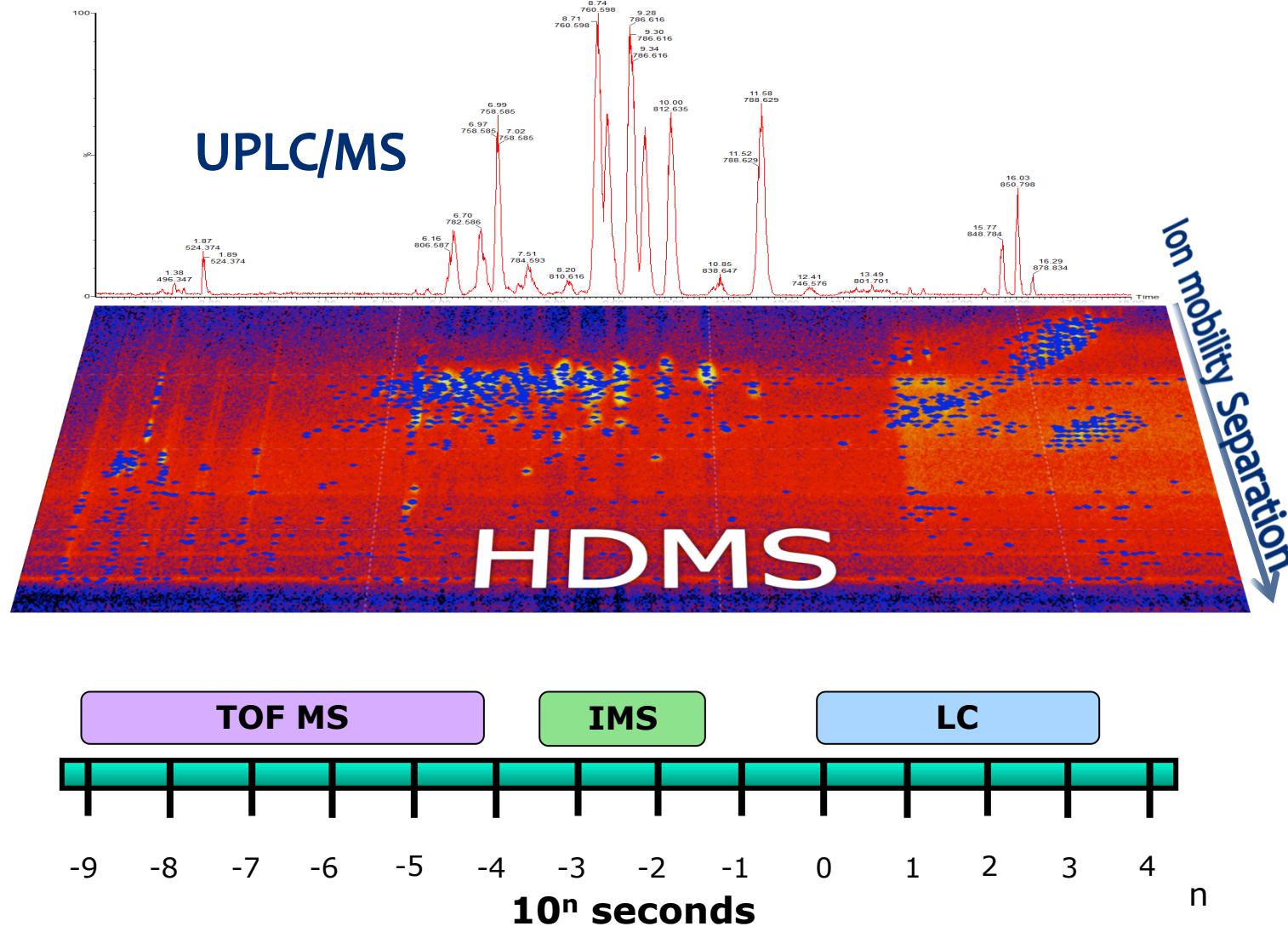


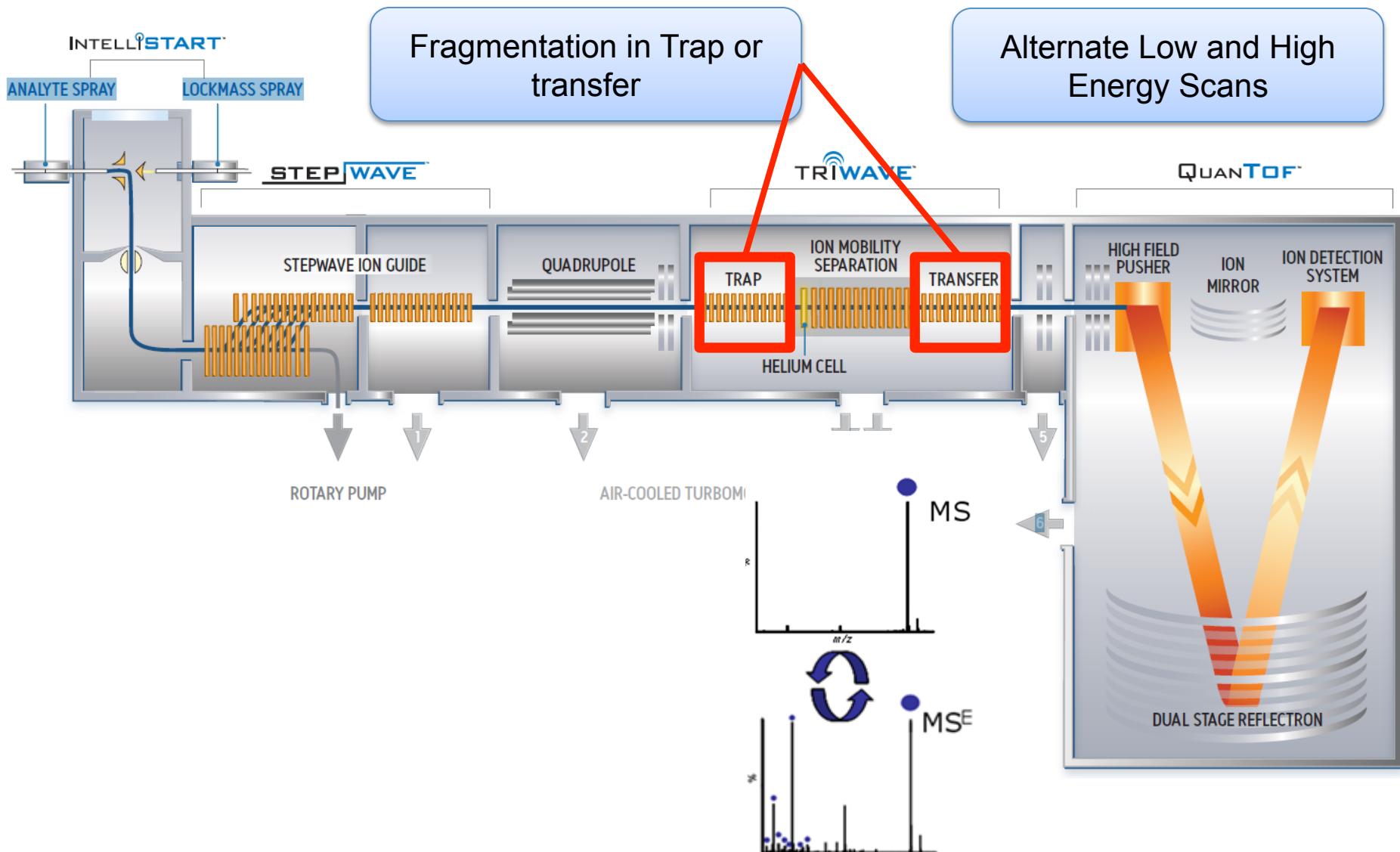
3. Accurate mass



Ion Mobility: an Orthogonal Dimension of Separation

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Environmental samples

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- SLU project
 - Anna Rotander, Sara Persson

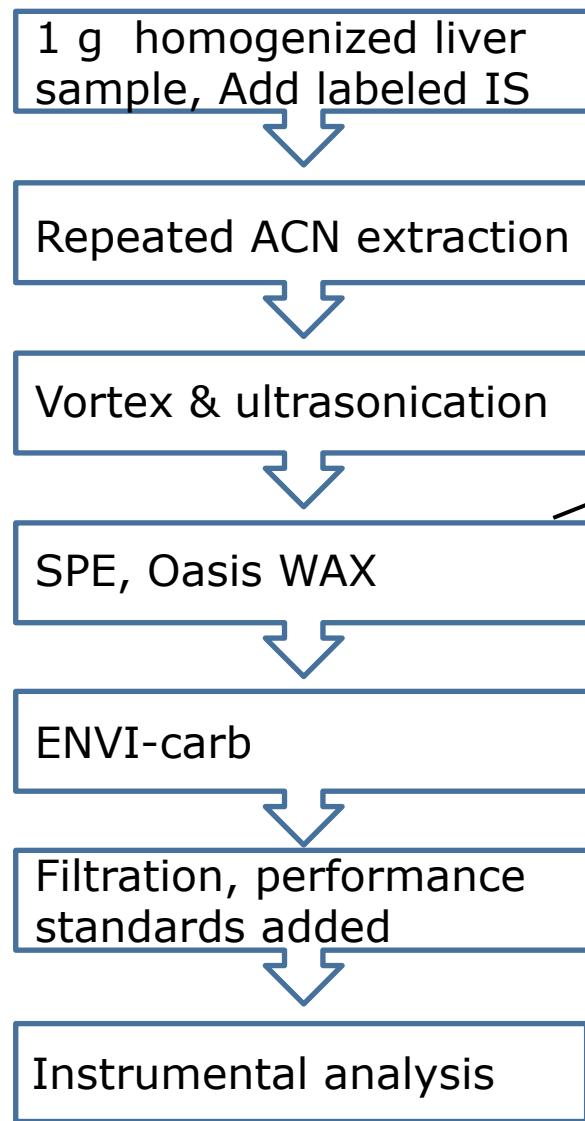
- Mink



Extraction of environmental samples

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Condition: MeOH and water

Load samples

Wash: 4 ml NaAc (pH 4)
4 ml 20% MeOH

Dry cartridges

Elute: 4 ml MeOH (discarded)
4 ml NH₄OH

Experimental

- **UPLC: Waters ACQUITY UPLC I-Class (equipped with PFC kit)**
- Column: Waters ACQUITY UPLC BEH C18 (100 mm x 2.1 mm, 1.7 µm)
- Column temperature: 50°C Flow: 0.30 mL/min
- Mobile phase A: H₂O:MeOH/ACN 70:30 (80/20, 2 mM Ammonium Acetate)
B: MeOH:ACN 80:20 (2 mM Ammonium Acetate)

■ **Gradient**

Time(min)	Flow Rate	%A	%B
Initial	0.300	100.0	0.0
0.50	0.300	100.0	0.0
16.00	0.300	65.0	35.0
22.00	0.300	65.0	35.0
27.00	0.300	10.0	90.0
27.10	0.300	0.0	100.0
28.00	0.300	0.0	100.0
28.10	0.450	100.0	0.0
34.00	0.450	100.0	0.0

Experimental

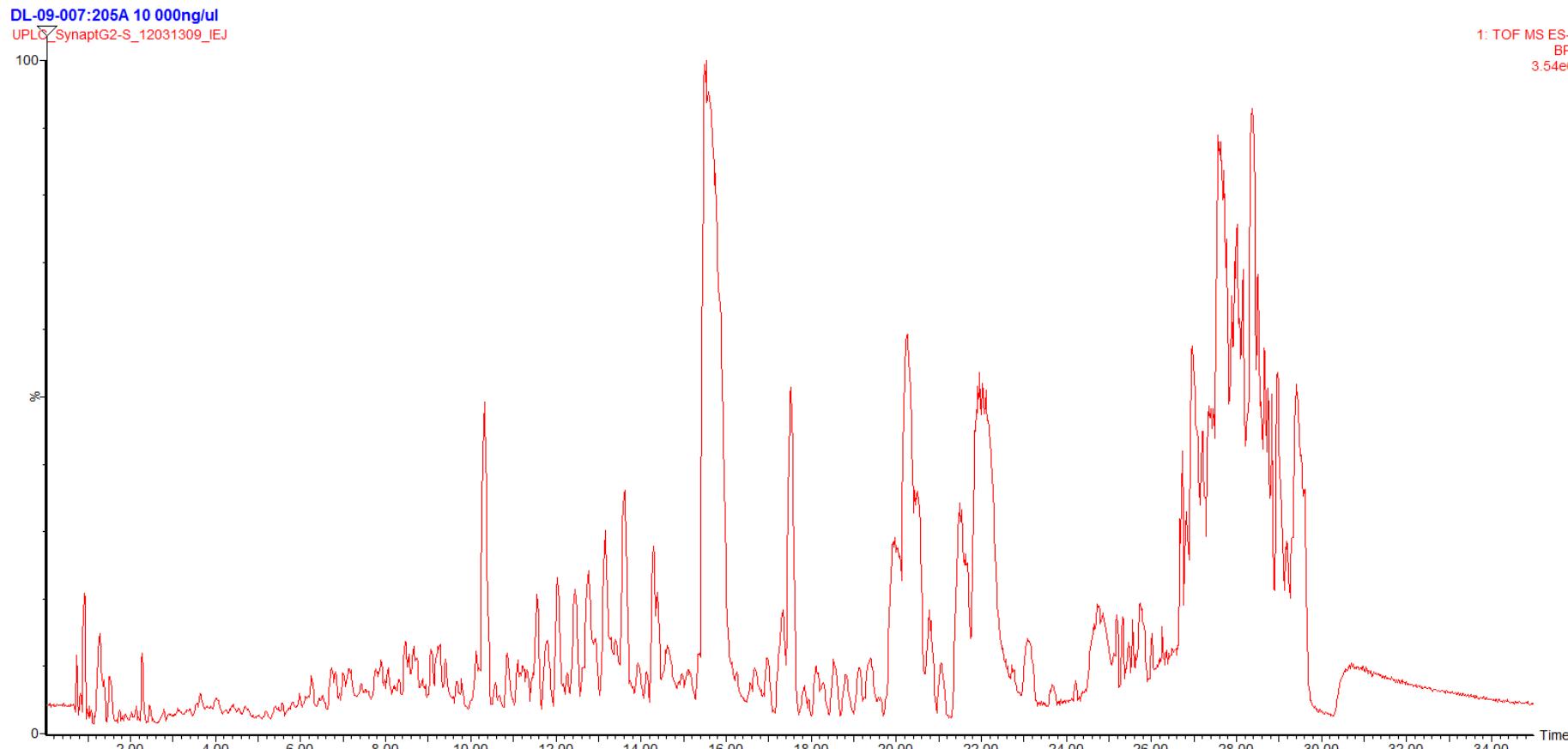
- **MS: Waters SYNAPT G2-S**
- Ionisation Mode: ES-
- Desolvation Temperature: 550 °C
- Acquisition Modes: IMS MS^E
- M/Z Range: 50-600
- Acquisition rate: 10 spectra/second
- Capillary Voltage: 2.3 kV
- Cone Voltage: 15 V
- Drift Gas: CO₂ and N₂
- Collision Energy Ramp: 35-75 eV
- IMS Wave Velocity Range: 400 m/s to 550 m/s
- IMS Wave Height: 40 V

Mobility Drift Gas	Mass	Polarisability (10e ⁻²⁴ cm ³)
Nitrogen N ₂	28.0123	1.7403
Carbon Dioxide	44.0098	2.9110

BPI Chromatogram for HDMS Analysis of Extract of Mink Liver for PFOS.

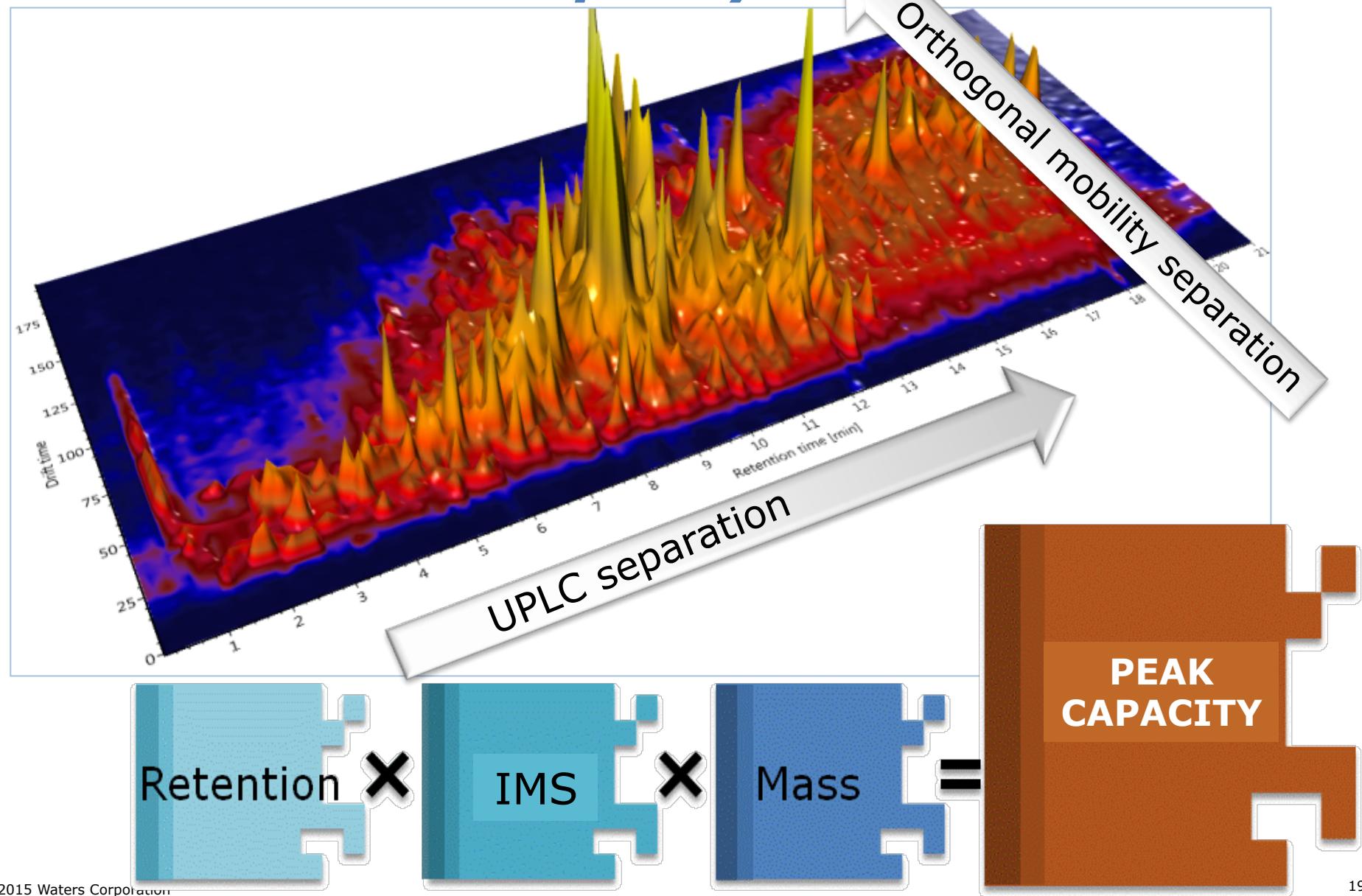
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50559 Components Detected



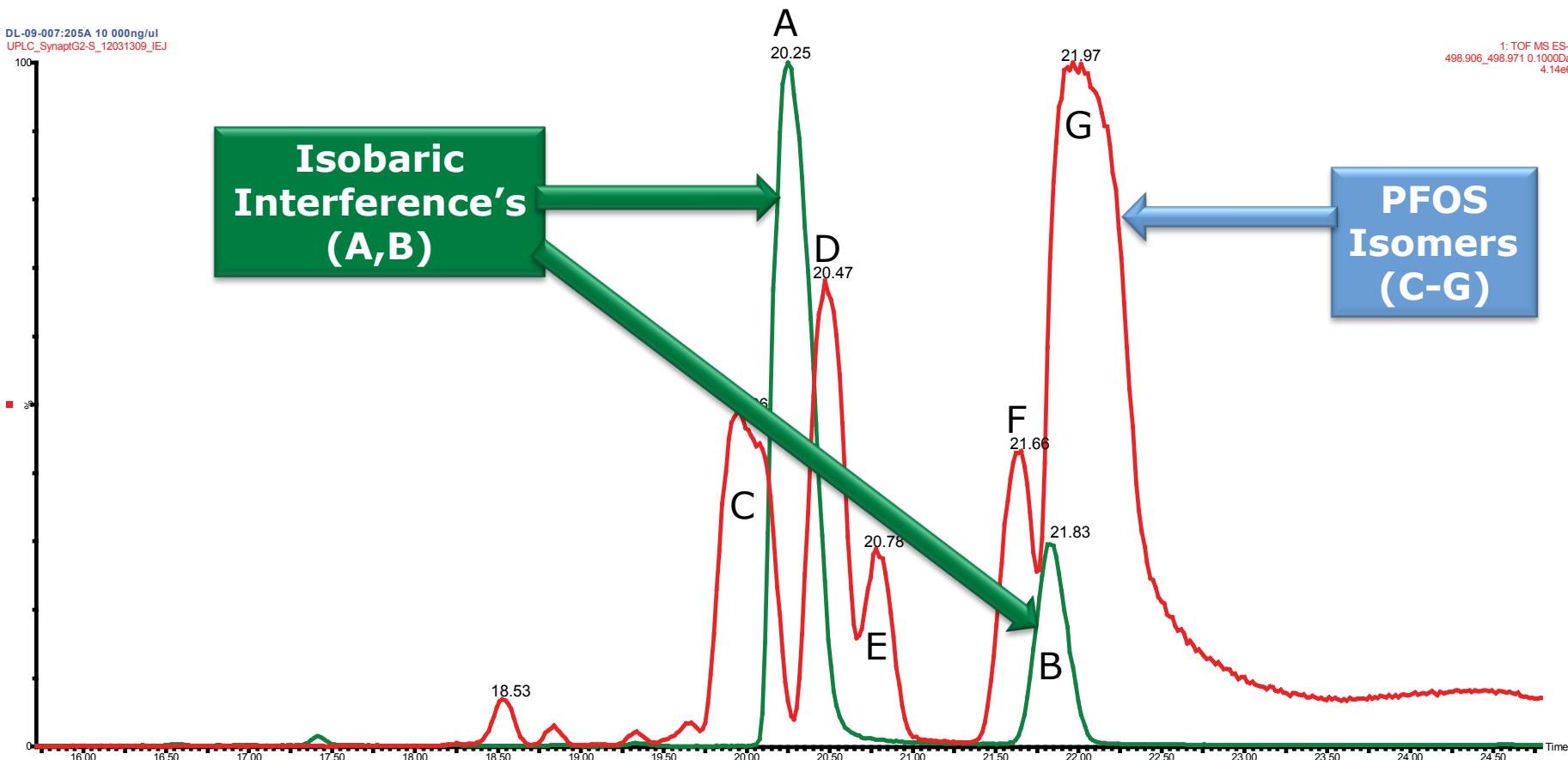
LC-HDMS^E means..... ***Increased Peak Capacity***

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Extracted ion chromatograms for matrix interferences and the PFOS isomers

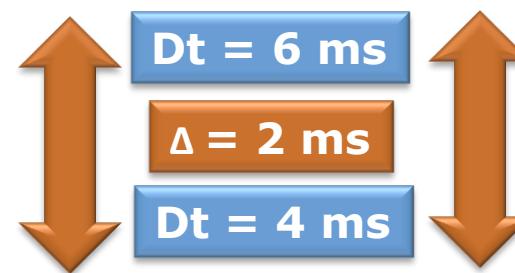
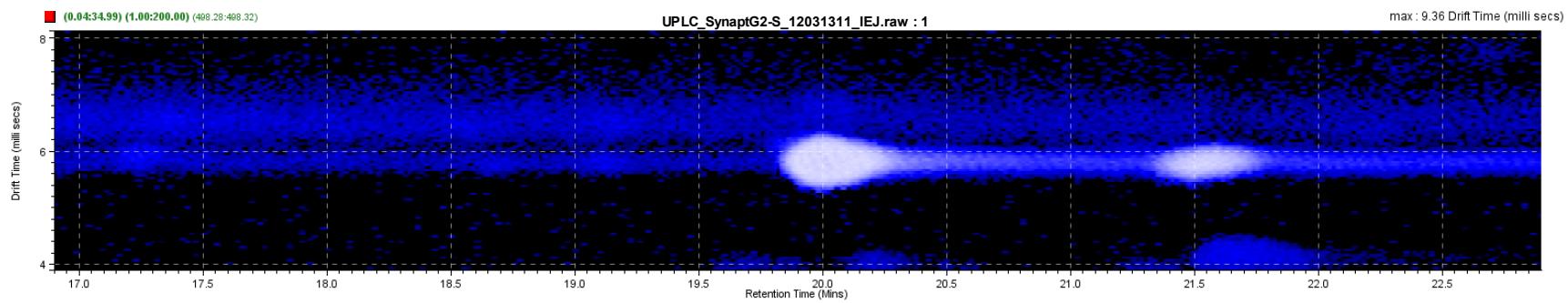
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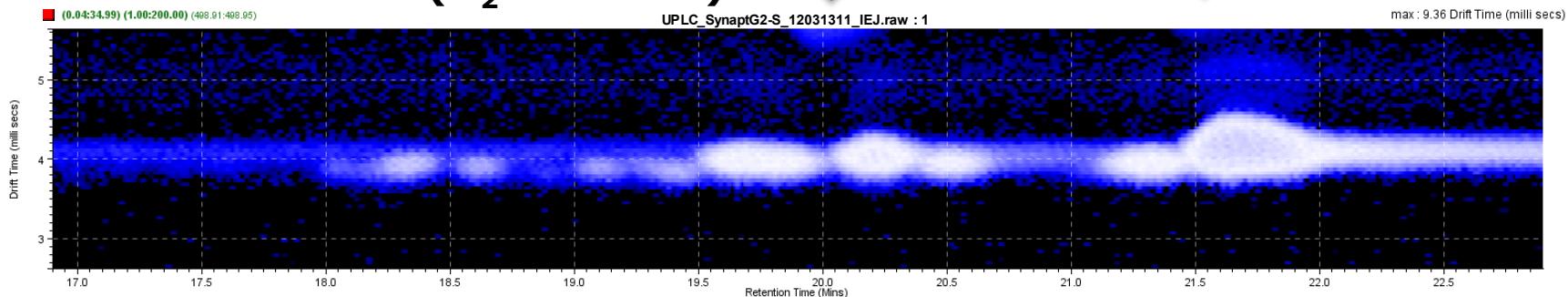
Mobility plots for the isobaric interferences and PFOS isomers using N₂ drift gas.

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Ion Mobility Separation TDCA Interference's (N₂ Drift Gas)

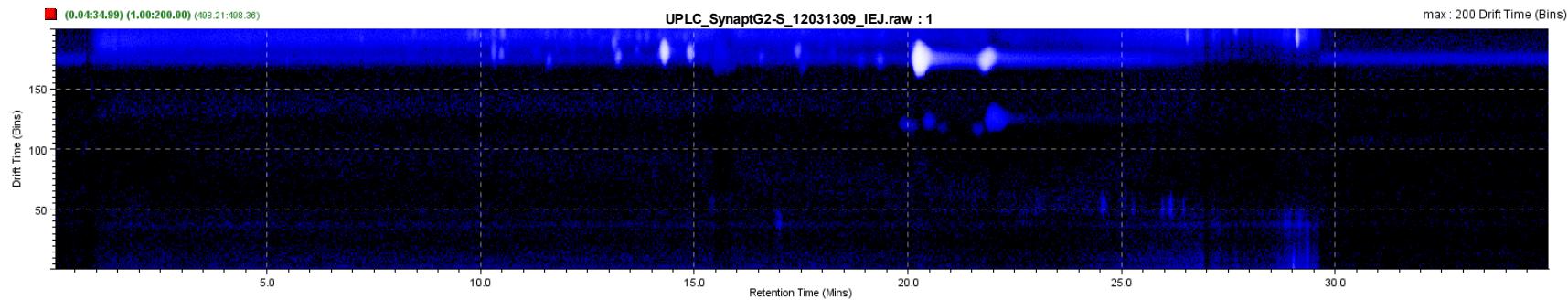


Ion Mobility Separation PFOS Isomers (N₂ Drift Gas)



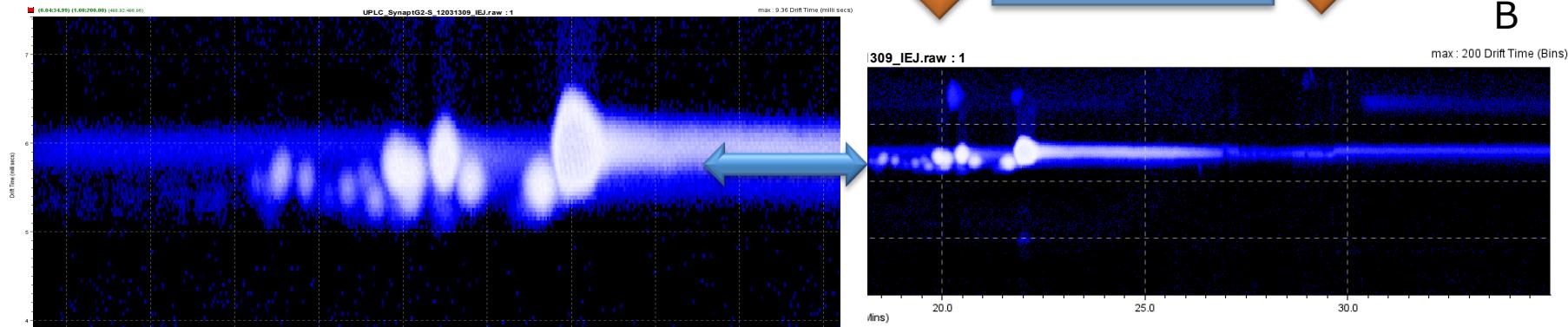
Mobility plots for the isobaric interferences and PFOS isomers using CO₂ drift gas.

TDCA/TCDCA Interference's (CO₂ Drift Gas)



A

PFOS Isomer Ion Mobility Separations (CO₂ Drift Gas)

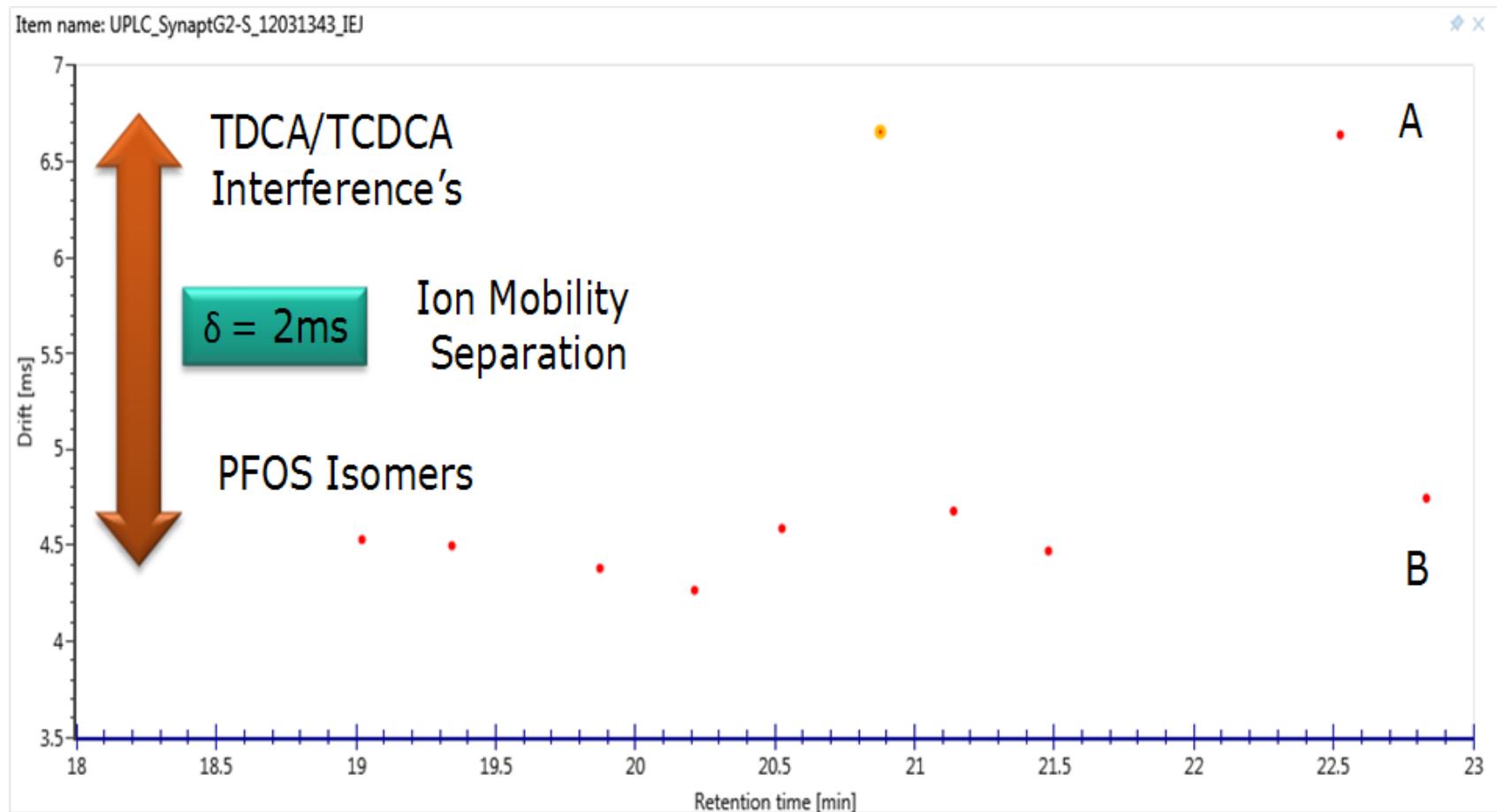


B

$$\begin{array}{c} \text{Dt} = 9.2 \text{ ms} \\ \Delta = 2.7 \text{ ms} \\ \text{Dt} = 6.5 \text{ ms} \end{array}$$

Ion Mobility Separation

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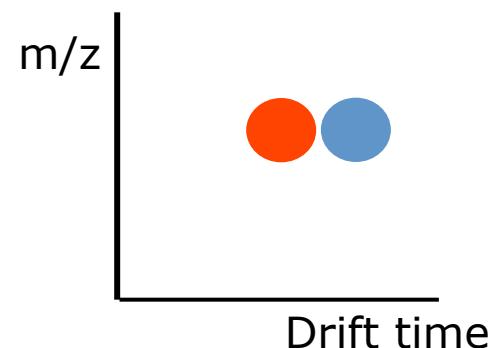
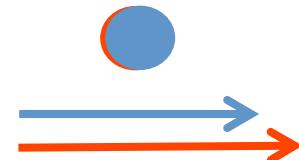
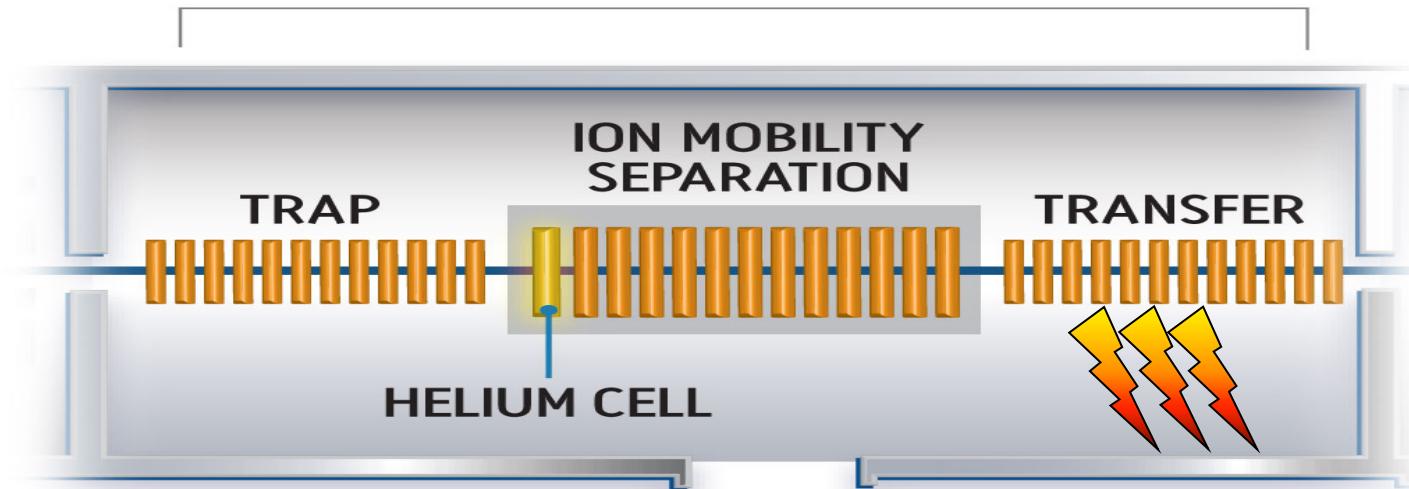
Component drift plot showing drift times vs retention time for nominally isobaric interferences (A) and PFOS isomers (B)

Time and Mobility Aligned Fragmentation

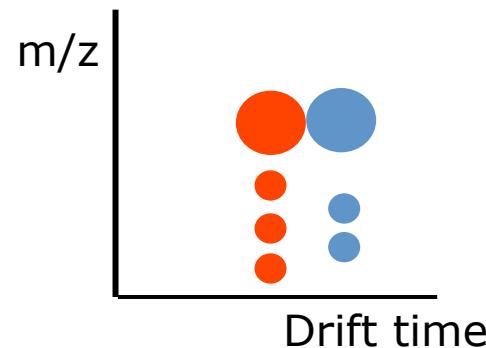
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TRI WAVE™



Precursor ions



Precursor ions

Precursor and products
are time aligned

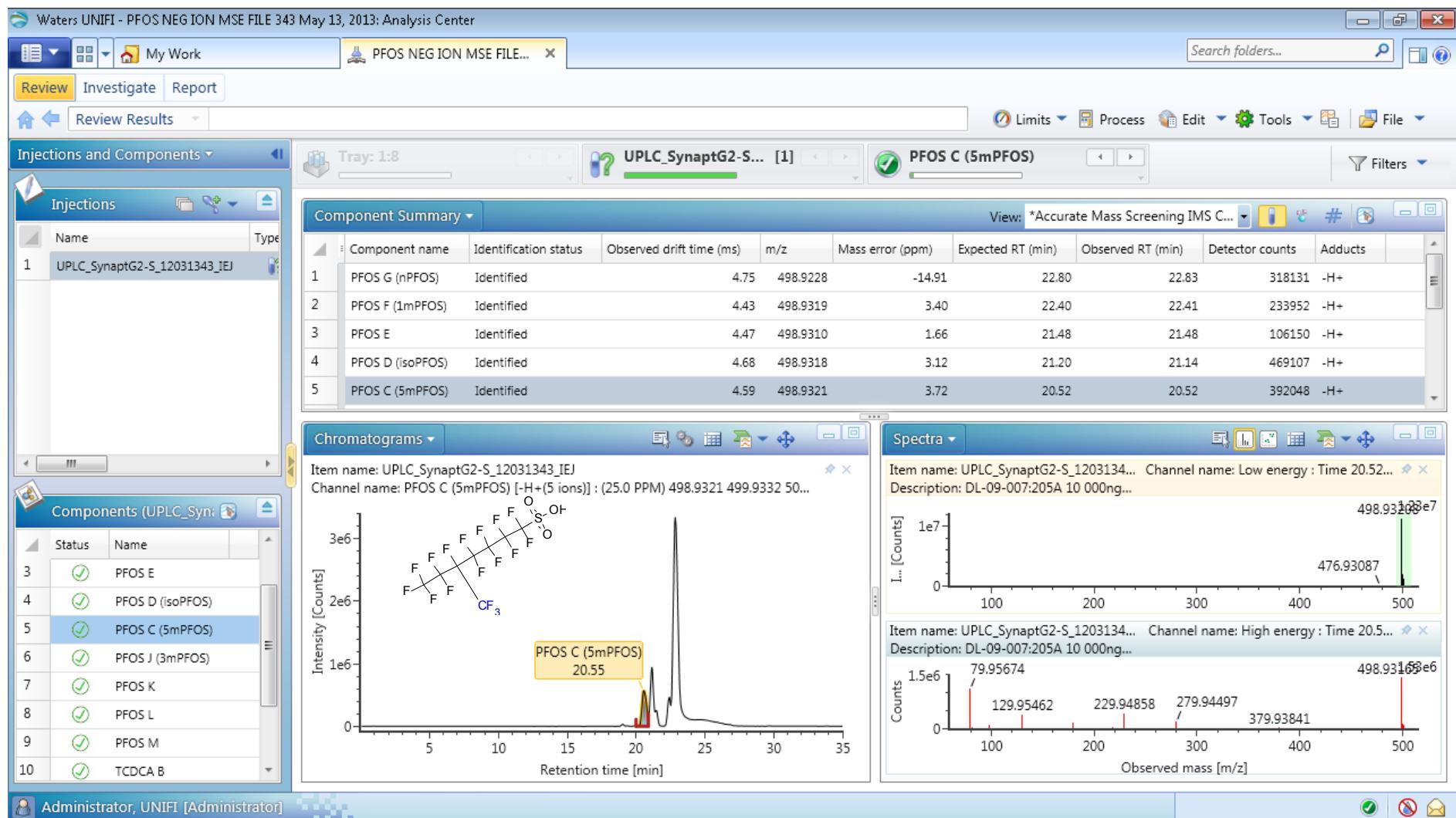
Ion Mobility Resolved TDCA (Interference A) with MS^E Precursor and Fragmentation Spectra

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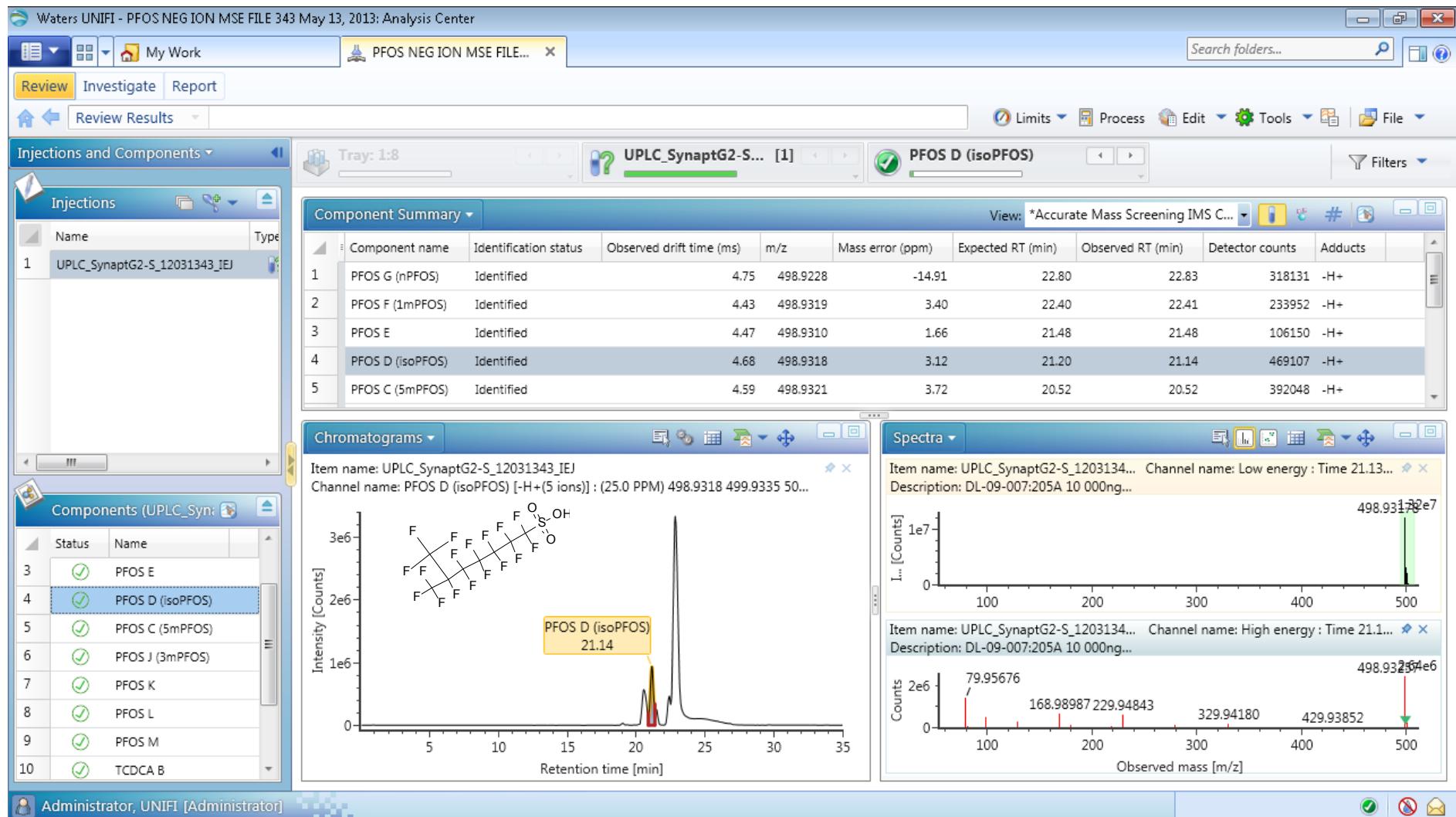
PFOS (C) isomer, ion mobility resolved from isobaric interference TCDCA (A) at retention time 20.55 mins

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PFOS (D) isomer, ion mobility resolved from isobaric interference TCDCA (A) at retention time 21.14 mins

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Unassigned PFOS (E), ion mobility resolved from isobaric interference TDCA (A) at retention time 20.88 mins

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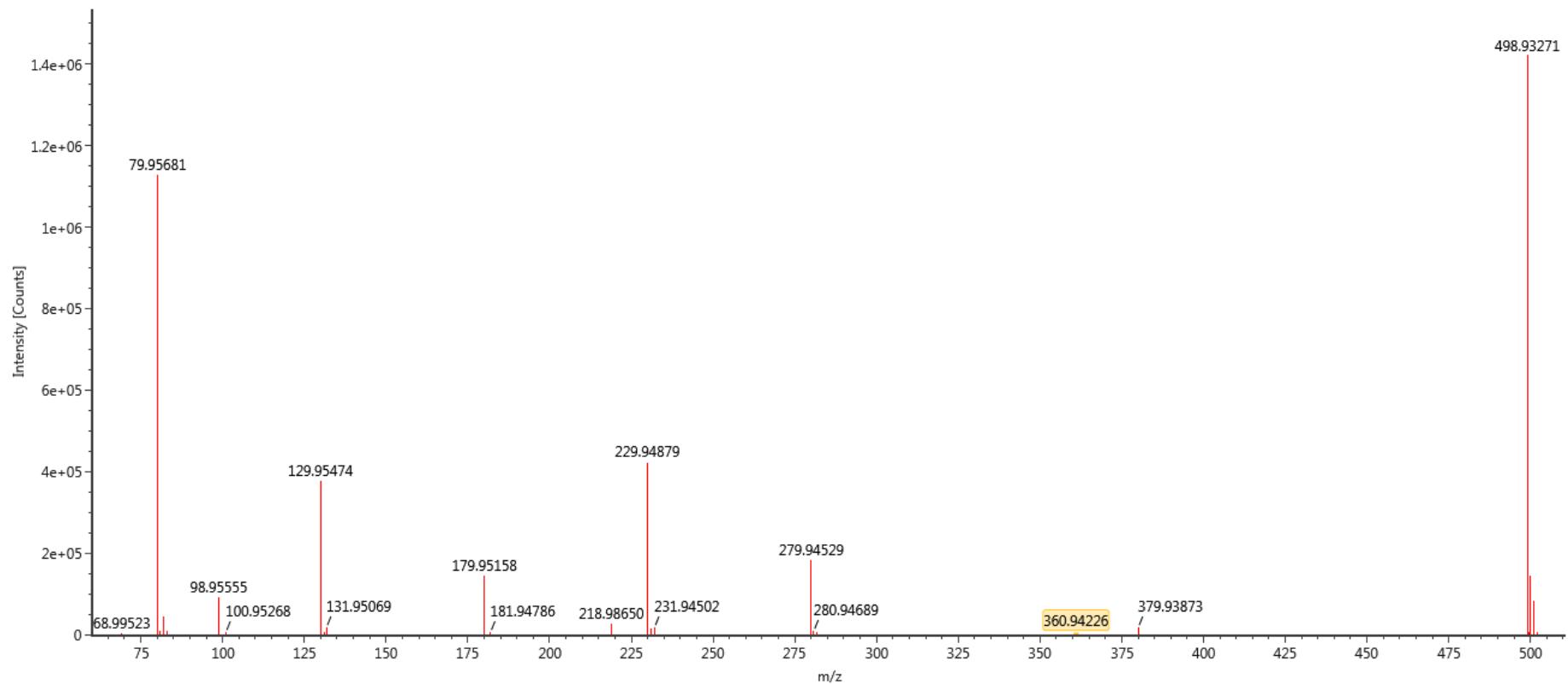


PFOS ISOMER C

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Item name: UPLC_SynaptG2-S_12031343_IE
Description: DL-09-007:205A 10 000ng/ul

Channel name: High energy : Retention Time 20.5245+/-0.0647 minutes : Drift Time 118.56+/-8.49 bins : 3D mass peak list

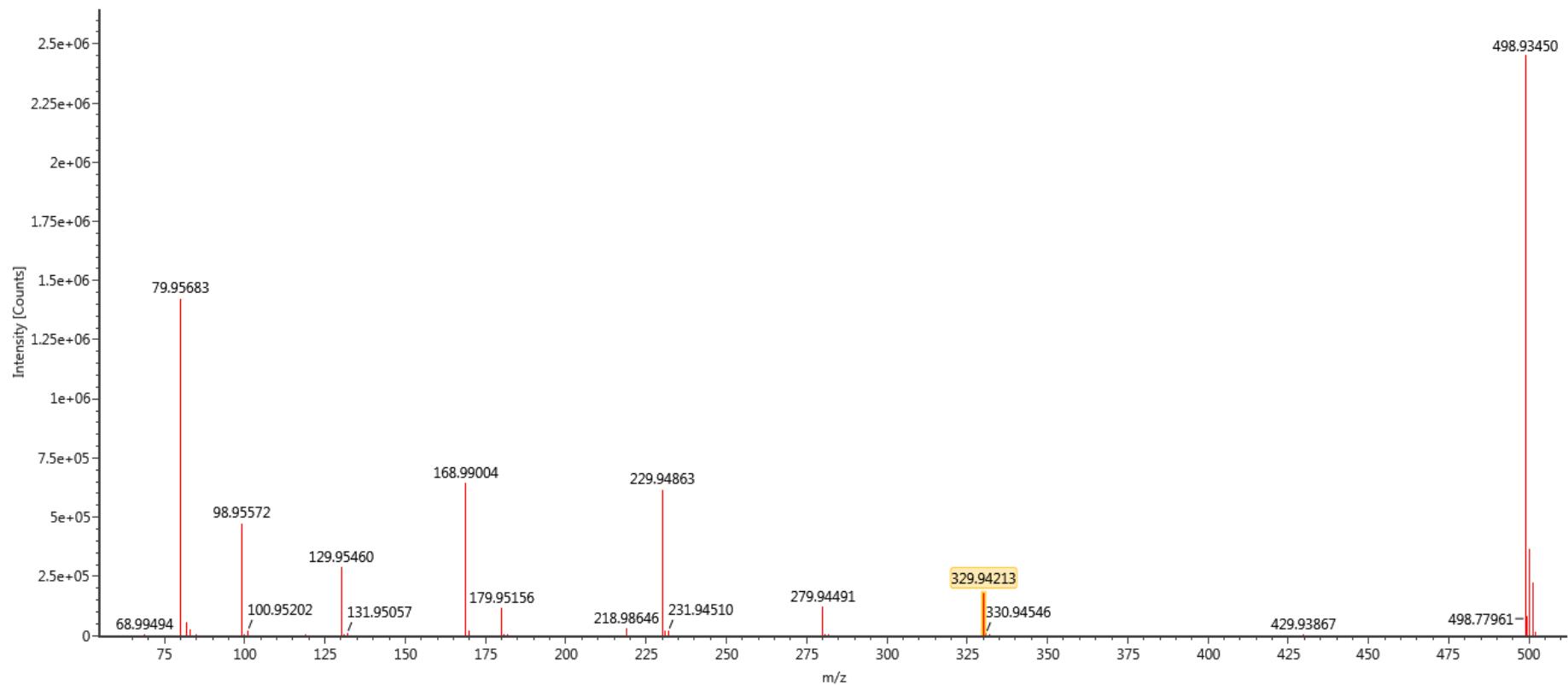


PFOS ISOMER D

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Item name: UPLC_SynaptG2-S_12031343_IE
Description: DL-09-007:205A 10 000ng/ul

Channel name: High energy : Retention Time 21.1390+/-0.0647 minutes : Drift Time 120.88+/-8.48 bins : 3D mass peak list

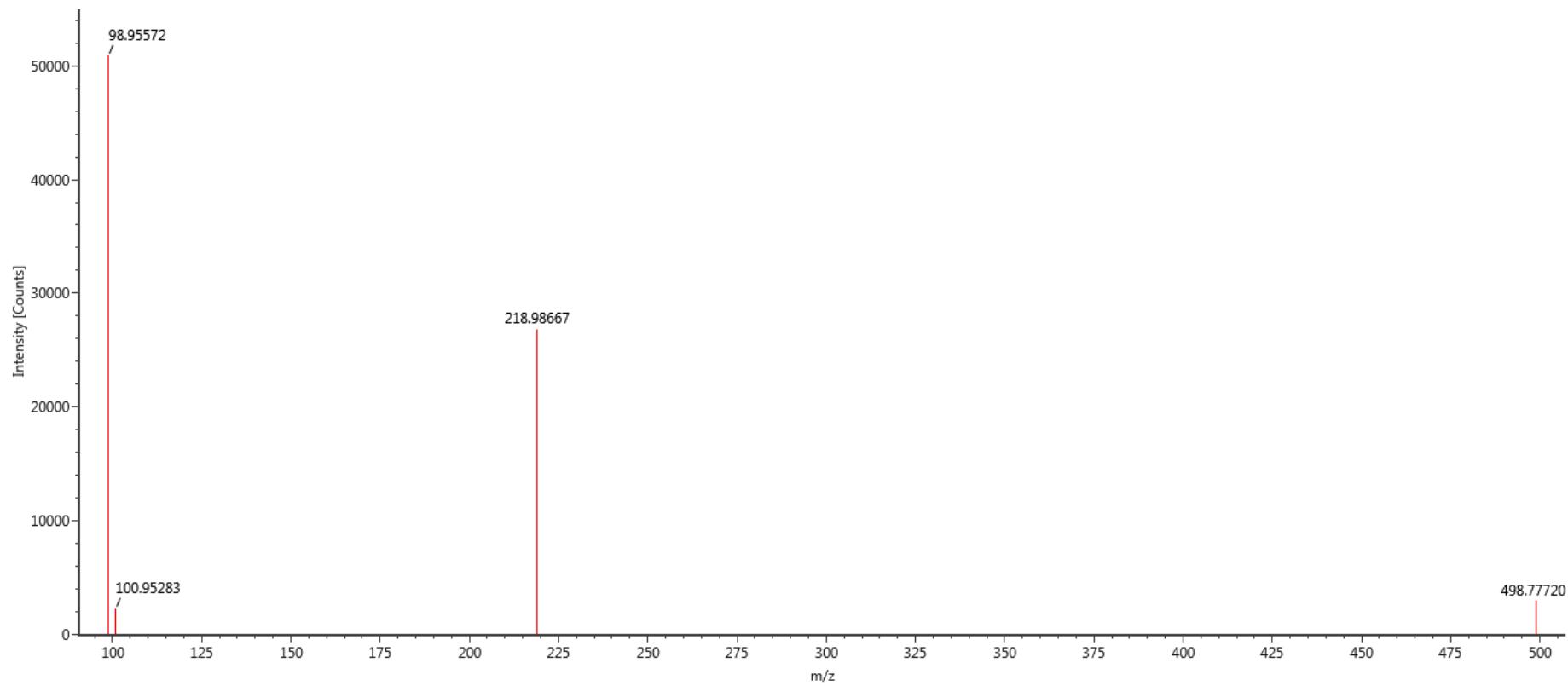


PFOS ISOMER E

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Item name: UPLC_SynaptG2-S_12031343_IE
Description: DL-09-007:205A 10 000ng/ul

Channel name: High energy : Retention Time 21.4786+/-0.0647 minutes : Drift Time 115.56+/-8.50 bins : 3D mass peak list

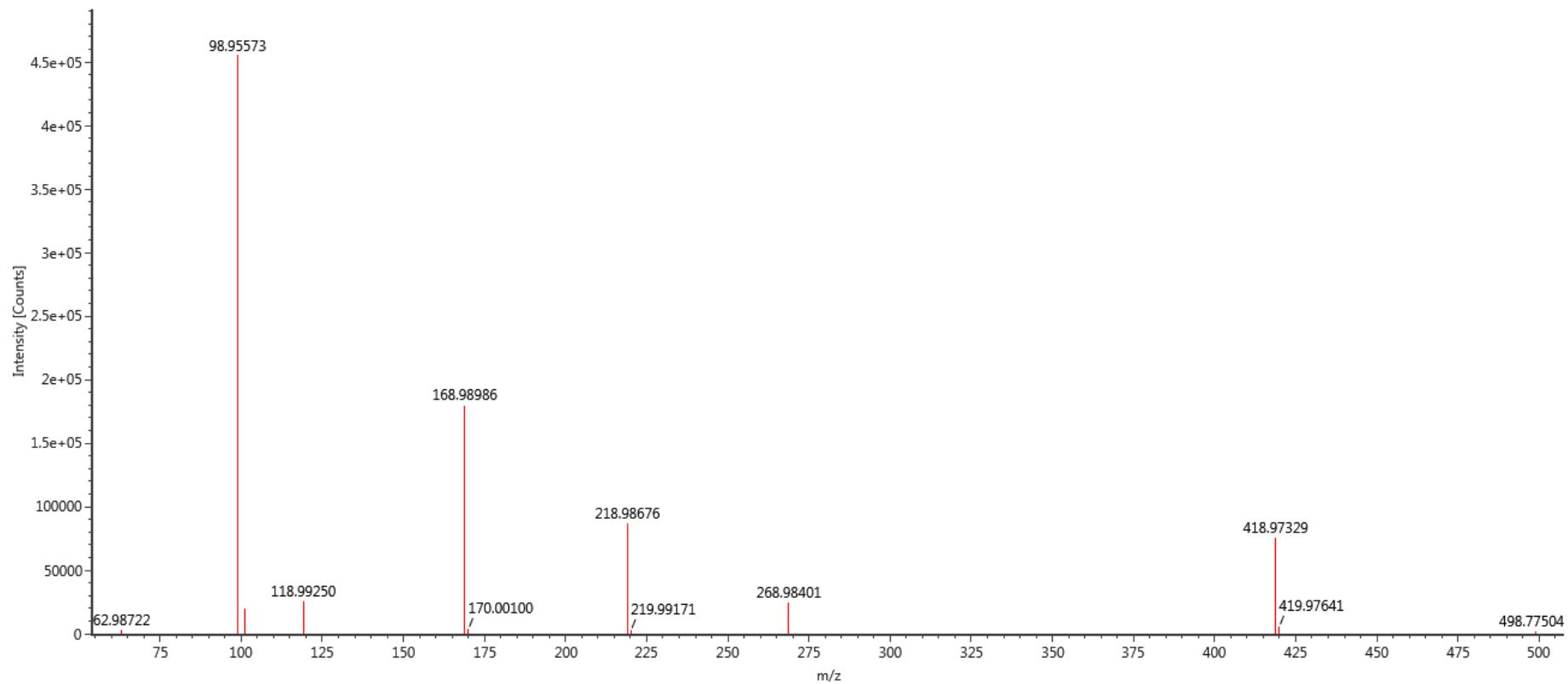


PFOS ISOMER F

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Item name: UPLC_SynaptG2-S_12031343_IE
Description: DL-09-007:205A 10 000ng/ul

Channel name: High energy : Retention Time 22.4055+/-0.0647 minutes : Drift Time 114.55+/-8.51 bins : 3D mass peak list

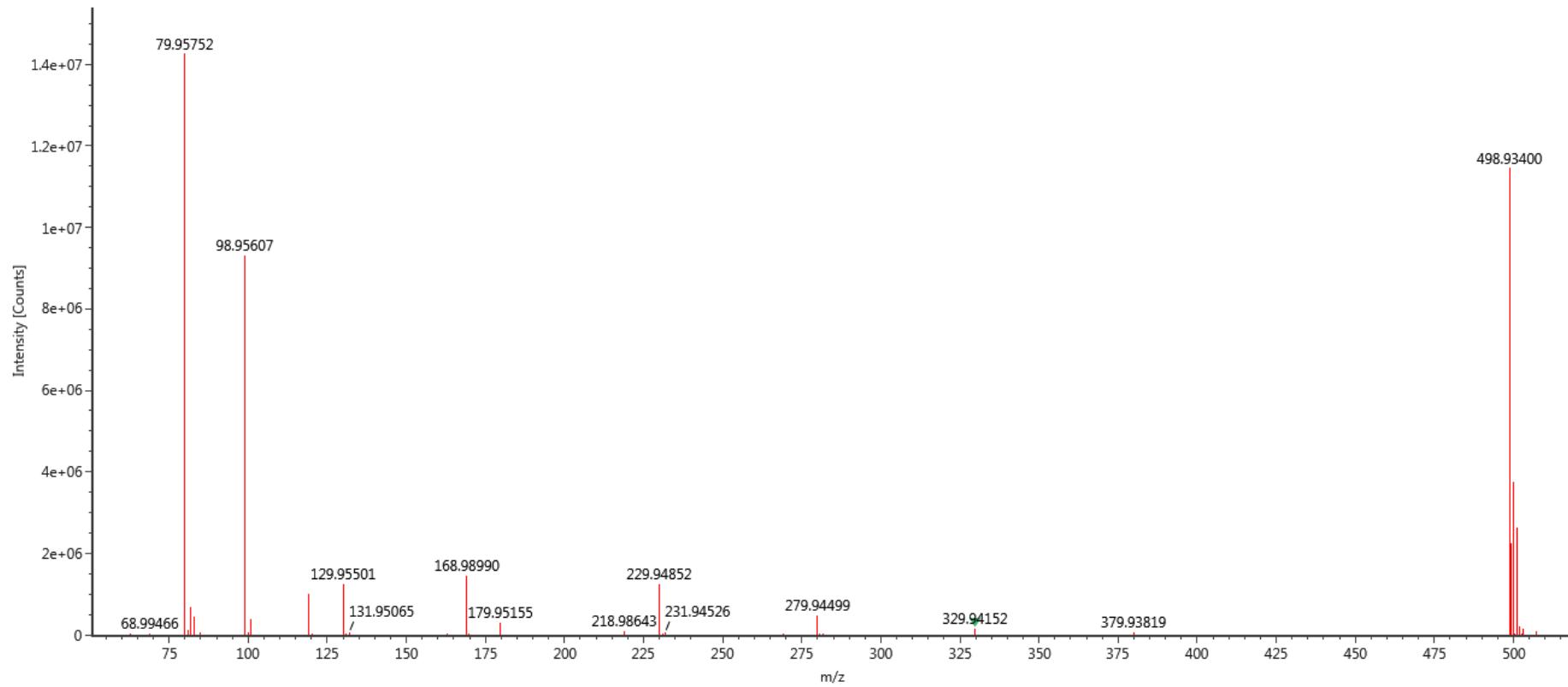


PFOS ISOMER G

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Item name: UPLC_SynaptG2-S_12031343_IE
Description: DL-09-007:205A 10 000ng/ul

Channel name: High energy : Retention Time 22.8319+/-0.0647 minutes : Drift Time 122.61+/-8.47 bins : 3D mass peak list



Summary for major PFOS isomers and matrix interferences

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	PFOS ISOMER IDENTIFICATION				
PFOS Isomers	C 5mPFOS J 3mPFOS	D IsoPFOS	E 2,2-perfluoro-methyl PFOS (tentative)	F 1mPFOS	G nPFOS
Drift Time (ms)	4.59 4.27	4.68	4.47	4.43	4.75
Mass Measurement Error	3.4 ppm -0.23 ppm	3.66 ppm	3.12 ppm	3.72 ppm	-14.91 (2.68ppm HE)
Retention Time (mins)	20.55	21.14	21.48	22.40	22.80
TDCA Interferences	A TDCA	B TCDCA			
Drift Time (ms)	6.65	6.64			
Mass Measurement Error	3.59 ppm	1.64 ppm			
Retention Time (mins)	20.88	22.52			

A summary of drift times, retention times and isomer assignments for major PFOS isomers and co-eluting matrix.

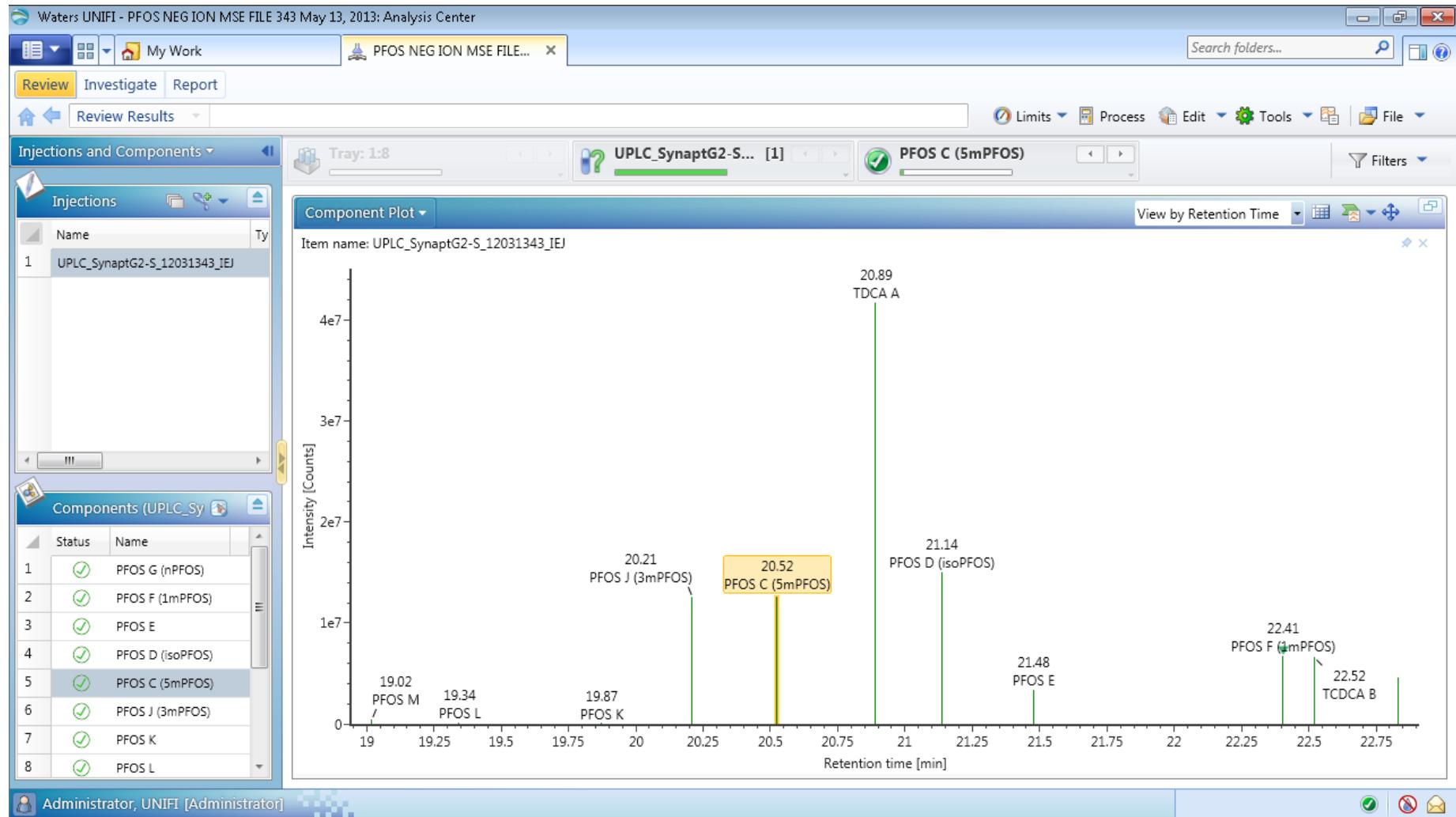
Screening database



- A screening database was created for branched PFOS isomers based on observed retention times and unique product ions
 - 1m-PFOS, 3m-PFOS, 4m-PFOS, 5m-PFOS, 6m-PFOS, 4,4dim-PFOS, 3,5-dim-PFOS, 4,5-dim-PFOS, 5,5-dim-PFOS
- Environmental samples and a technical blend was analysed and compared against the database.
- The software filtered the results to include only isomers having a positively identified product ion.

Component summary for PFOS screening of mink liver extracts

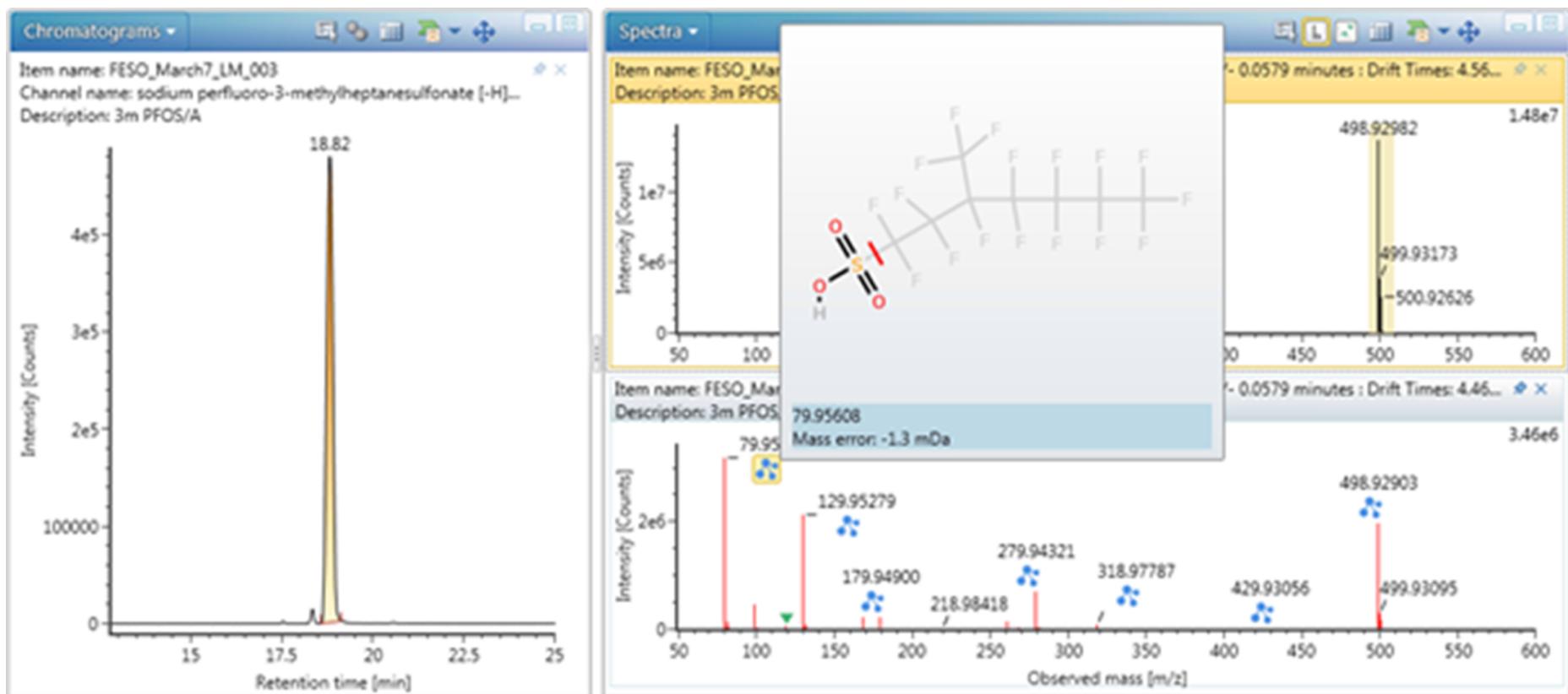
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Software enabled identification

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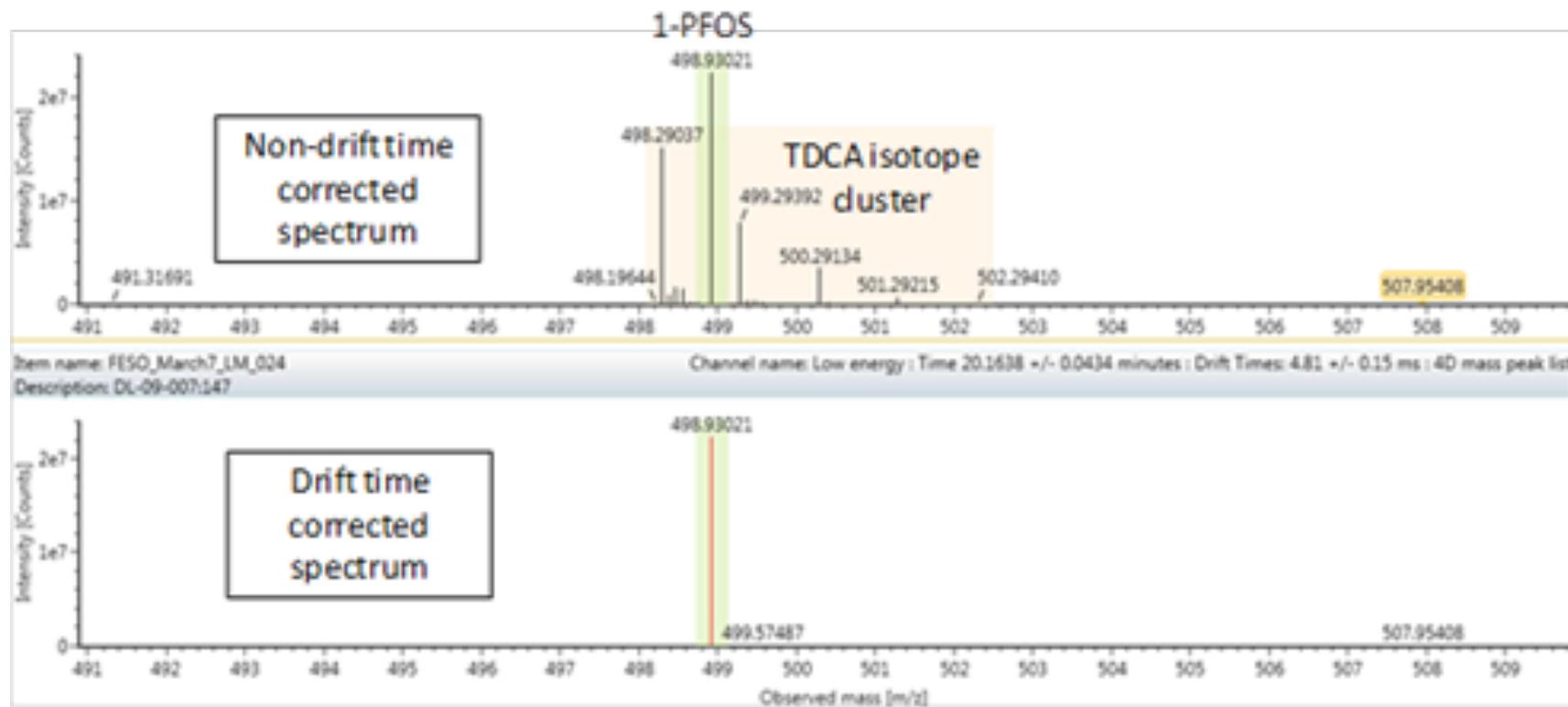
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Extracted ion chromatogram and spectrum for both low energy (top) and high energy (bottom) collision states. Using the structure, proposed product ions were deduced from the observed spectral peaks, as indicated by the blue molecule icons.

Drift time corrected spectrum

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Spectrum for the same chromatographic peak, 1-PFOS, with interference from TDCA evident in the top spectrum. The bottom spectrum is from the same chromatographic peak and injection, but with the removal of any ions that do not share the same drift time as 1-PFOS.

Conclusions

- Co-eluting isobaric biological interference's TDCA and TCDCA have been resolved from PFOS isomers using ion mobility.
- Utilising CO₂ as a mobility drift gas further enhanced the mobility resolution between PFOS isomers and isobaric TDCA / TCDCA interference isomers.
- The enhanced mobility resolution induced using CO₂ has a drift gas also enabled distinctive drift times to be obtained for the PFOS isomers.
- Single component precursor ion and fragmentation spectra have been generated for PFOS isomers and TDCA / TCDCA interference
- PFOS isomers can be characterised without contribution from isobaric interference isomers of TDCA / TCDCA.
- UPLC IMS MS^E facilitates accurate and informative data generation by isomer-specific analysis. This will be increasingly important as regulatory, scientific, and public awareness of the environmental impact of perflouralkylsubstances increases.



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BENGT LUNDQVIST MINNE

Waters Centre of Innovation

Contact:

ingrid.erickson@oru.se



Thank You!!!

Questions???

Ken Rosnack

ken_rosnack@waters.com

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