

Assessing Calibration-Related Measurement Bias Near the Limit of Quantitation

Troy Strock¹, Wayne Whipple¹, Steve Reimer², Diane Gregg³

¹US Environmental Protection Agency, Region V Laboratory
536 S. Clark St. 10th floor (ML-10C), Chicago IL 60605

² US Environmental Protection Agency, Region X Manchester Laboratory
7411 Beach Drive East (LAB), Port Orchard, WA 98366

³ US Environmental Protection Agency, Region VI Laboratory, Houston Branch
10625 Fallstone Road (6MD), Houston, TX 77099

Disclaimer

The opinions expressed herein are the author's and (due to limited time for review and busy schedules) do not necessarily reflect the opinion of the co-authors, let alone the US EPA.

How low can you go?



Measurement sensitivity is an important consideration for many analytical chemistry applications

Environmental laboratories and instrumentation with lower sensitivity may have a competitive advantage

However, instrument sensitivity and background in reagents and standards can vary over time, depending on the analyte

Allowing intercept of calibration function to float (and weighting) enables regression line to fit lowest data points better; However, this can lead to measurement bias in applying the calibration curve, especially when extrapolated outside the calibration range

Definitions

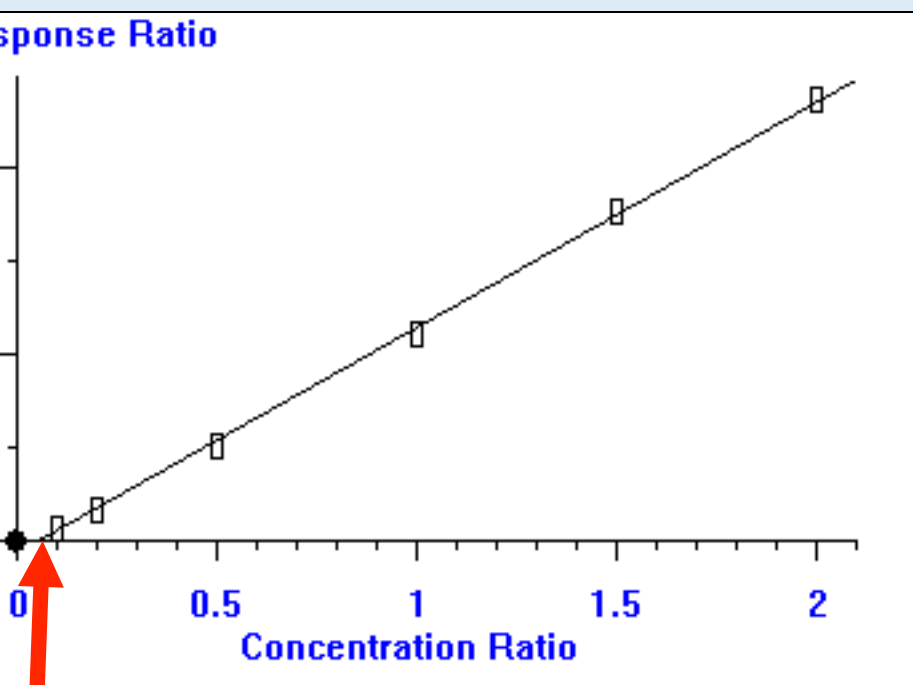
Calibration—The act of evaluating and adjusting the precision and accuracy of measurement equipment. Instrument calibration is intended to eliminate or reduce bias in an instrument's readings over a range of continuous values. http://www.chemwiki.ucdavis.edu/Analytical_chemistry/Data_analysis/Instrument_Calibration_Over_A_Regime/

Bias—a systematic error that contributes to the difference between the mean of a large number of test results and an accepted reference value. <http://www.astm.org/ILS/precisionbias.html>

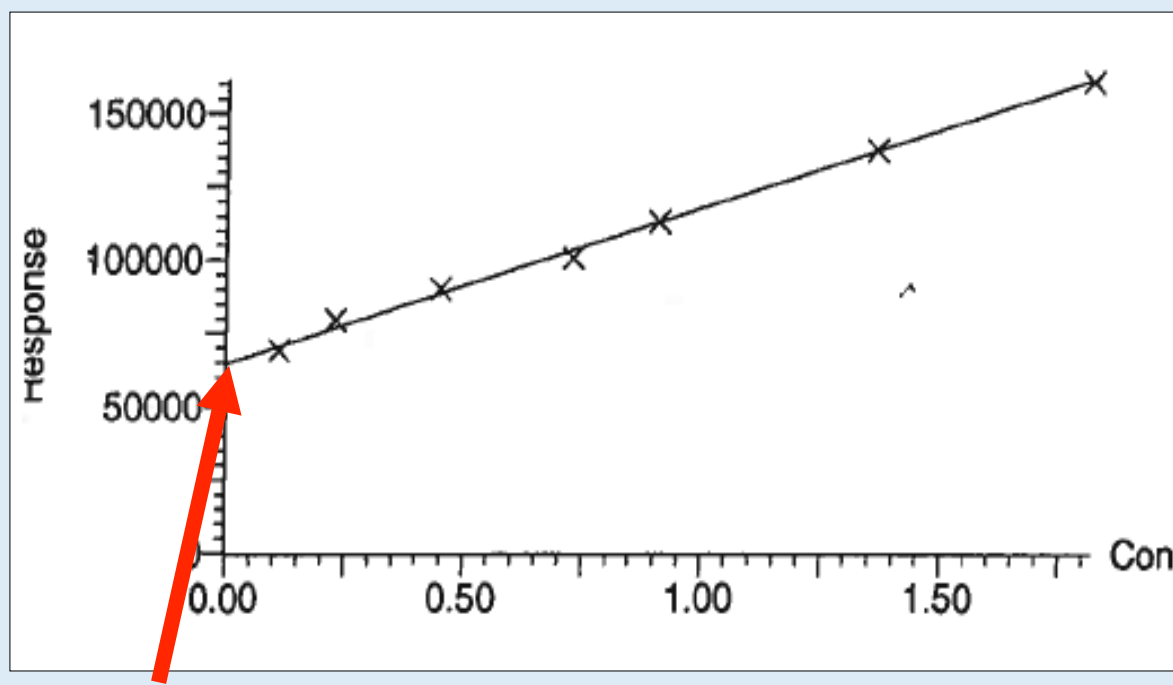
Limit of quantitation— The lowest concentration at which the analyte can not only be reliably detected but at which some predefined goals for bias and imprecision are met. <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC2556583/>

Calibration related measurement bias -- Loss of proportionality between measured response and calculated concentration due to application of the calibration model

Two types of calibration related measurement bias



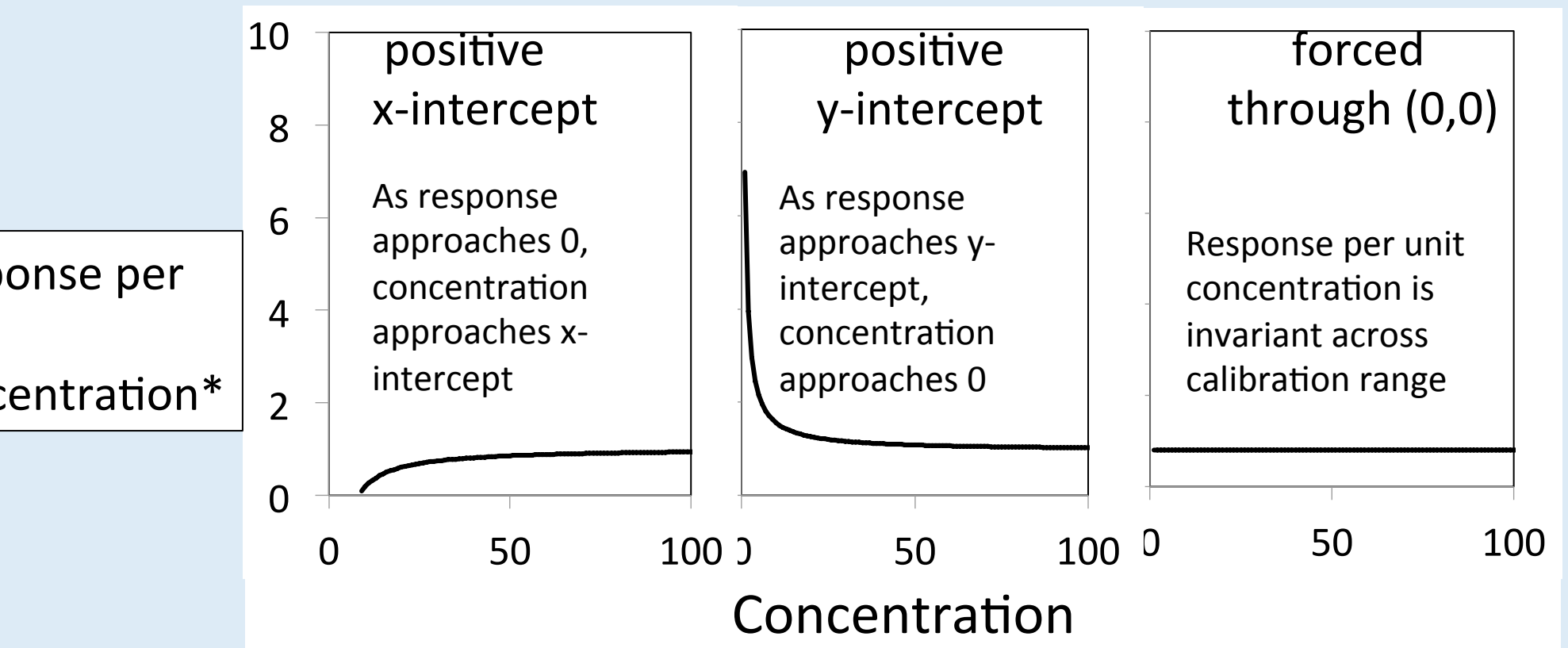
Positive x-intercept: Curve produces a positive calculated concentration at response = 0



Positive y-intercept: Curve produces a calculated concentration of = 0 at a response > 0

types of calibration related measurement bi

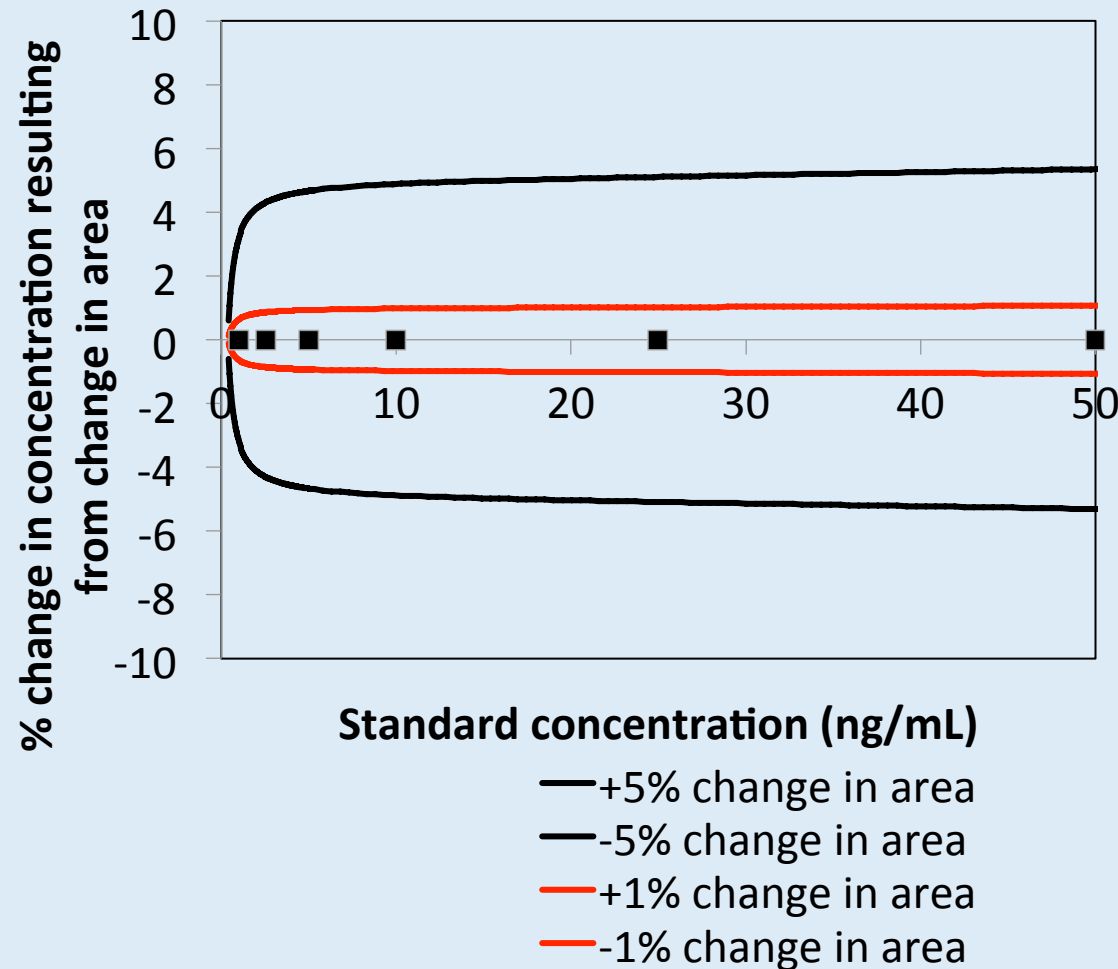
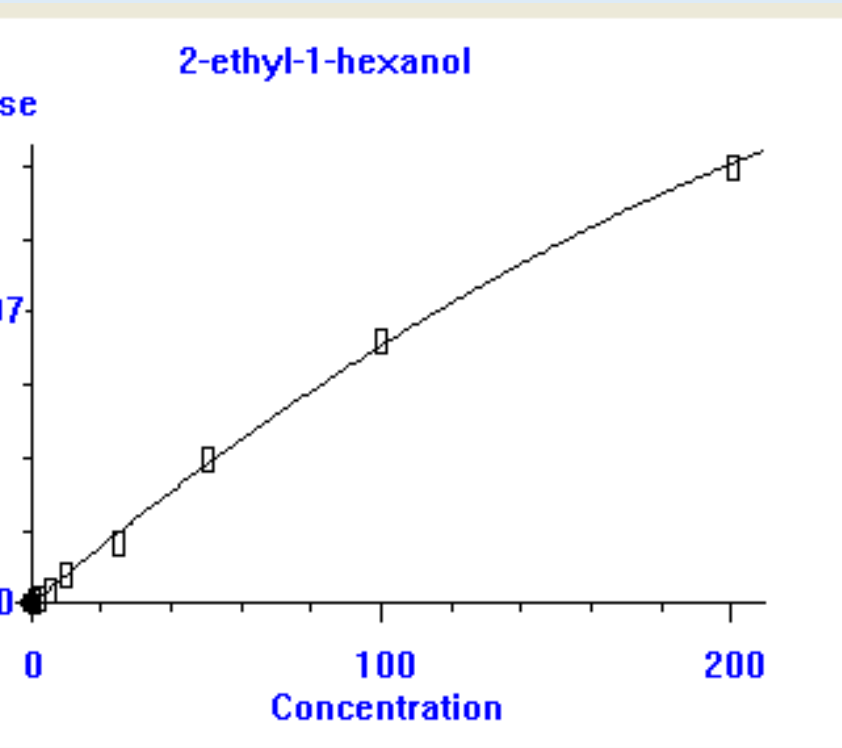
For calibration models that do not pass through (0,0), modeled response per unit concentration is not linear



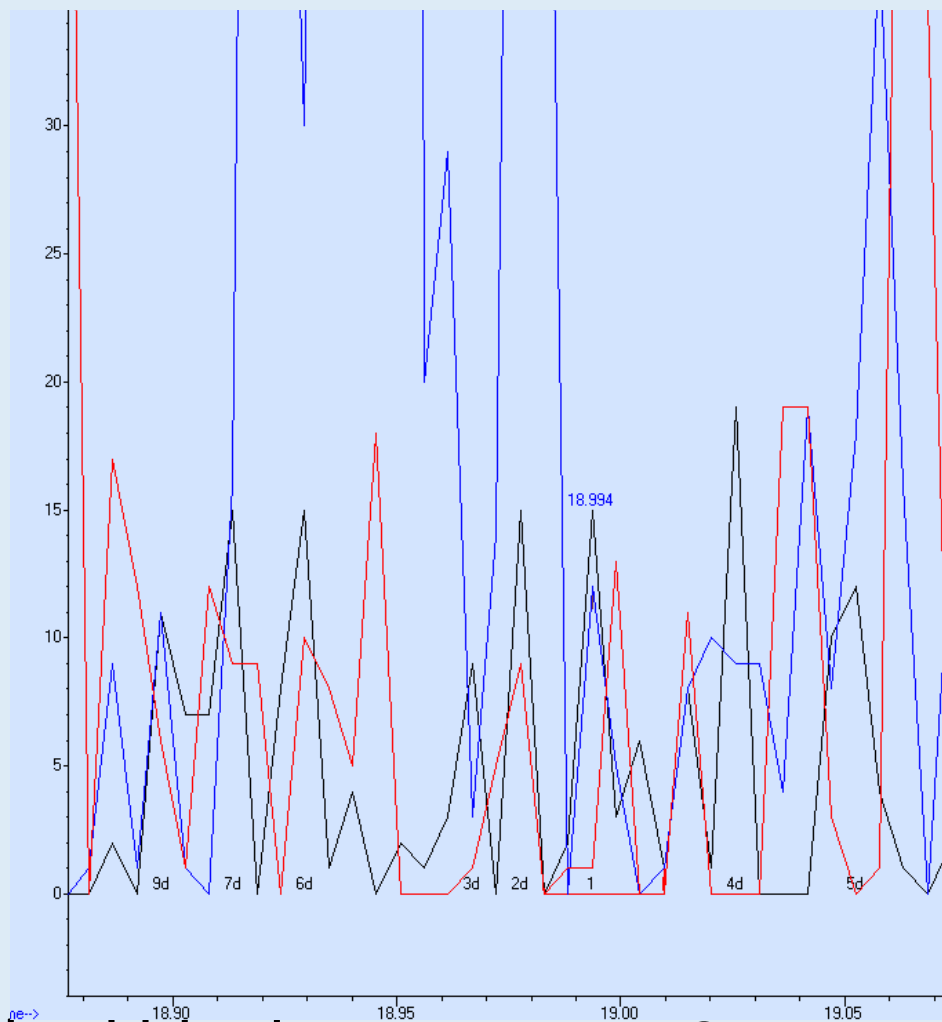
*Recalculated expected response across calibration range based on calibration function, then divide expected response by concentration

Curves with Positive X-Intercepts

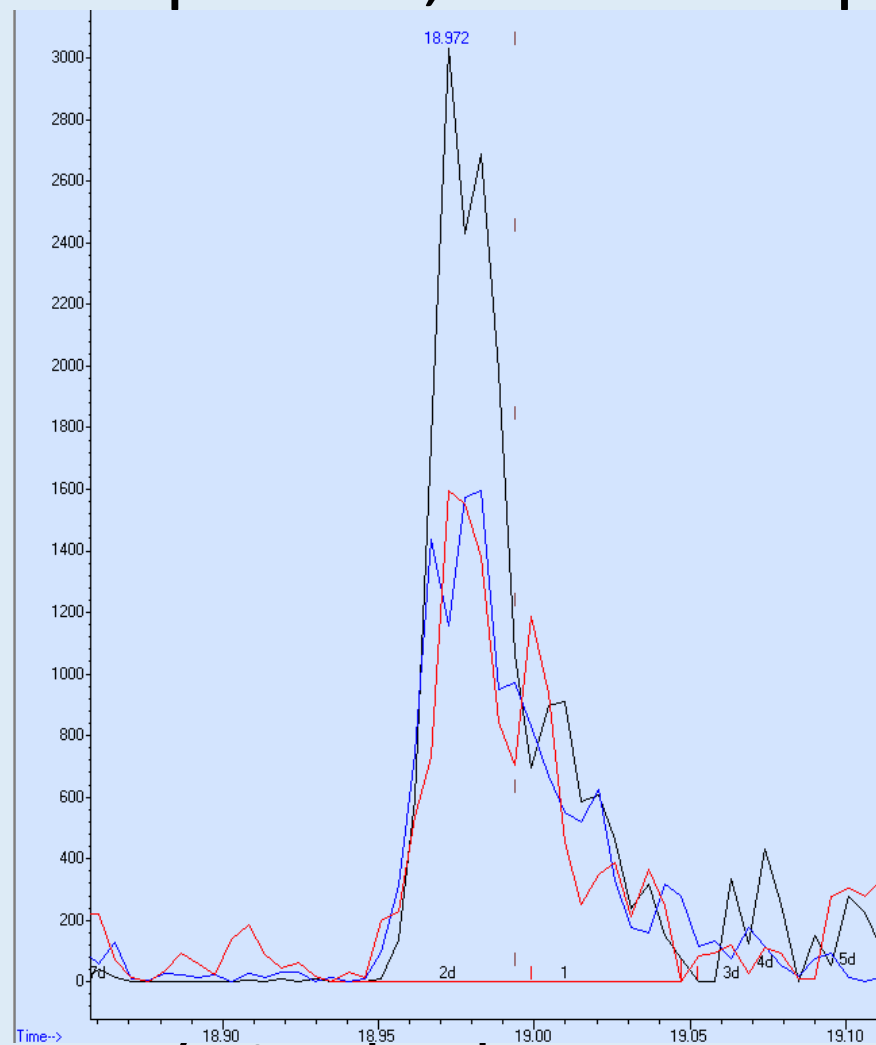
2-ethyl-1-hexanol by 5030/8260, 80 deg C
(x-intercept is ~60% of 1 ppb standard concentration)



Curves with Positive X-Intercepts: 2,4-dinitrophenol



Method blank, response = 9
Calc concentration = 2.05 ug/mL



2.5 ug/mL cal std, response = 5962
Calc concentration = 3.05 ug/mL

Curves with Positive x-Intercepts: Pentachlorophenol

Concentration (ug/mL)	Calc RRF
0.5	0.031
1	0.031
2.5	0.056
5	0.076
10	0.093
25	0.119
50	0.128
75	0.137
100	0.134

Calibration range (ug/ mL)	LLOQ standard* conc (ug/mL)	Blank concentration (ug/mL)	Blank an low stan area
0.5-100	1	0.80	24%
1-100	2.5	1.28	14%
2.5-100	2.5	2.03	2.1%
5-100	5	2.77	0.7%

*LLOQ: Lower Limit of Quantitation, set as lowest standard whose calculated concentration was within $\pm 50\%$ of expected. From SW-846 method 8000D.

ives with Positive x-intercepts: Methylated -dinitro-2-sec-butyl phenol

ad 1/x weighted

-125 ug/L

= 0.998

=9.1%

nt) (IS conc) = 1.7 ug/L

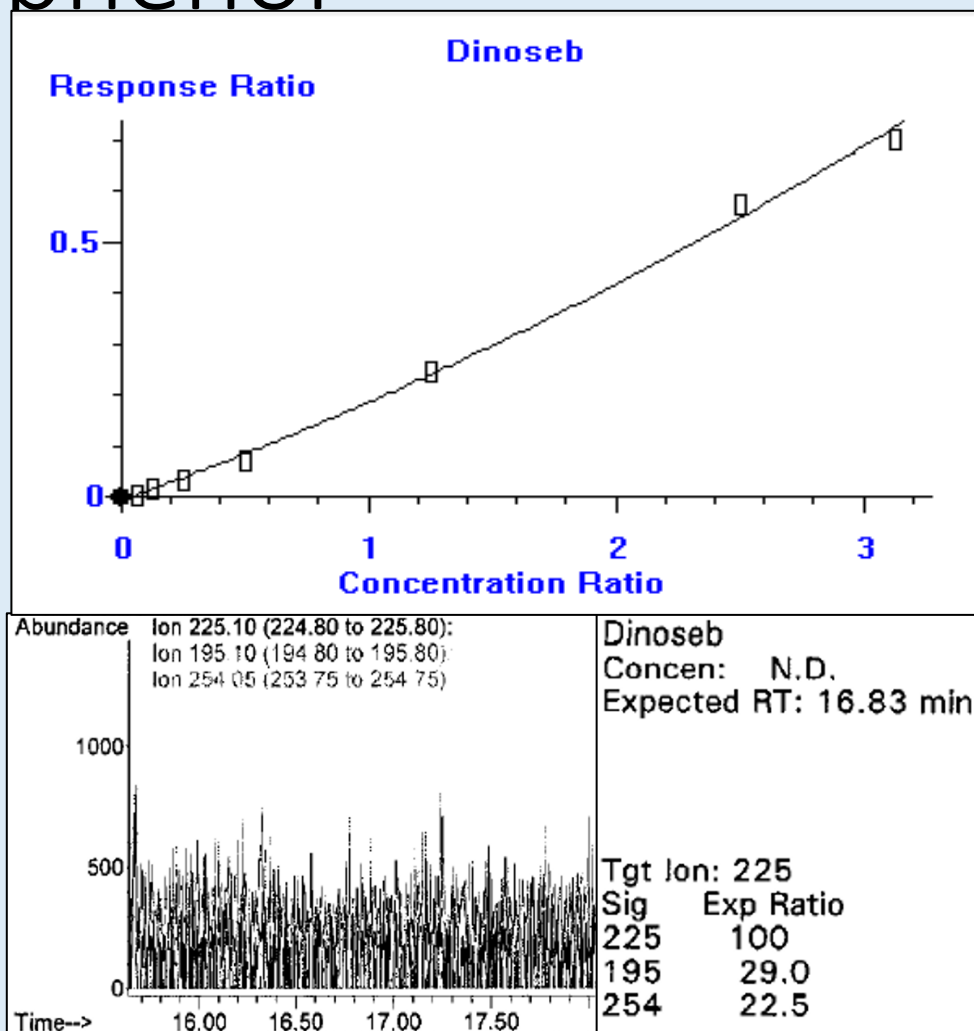
ethod Detection Limit Study
(2.5 ug/L spike level, n=7)

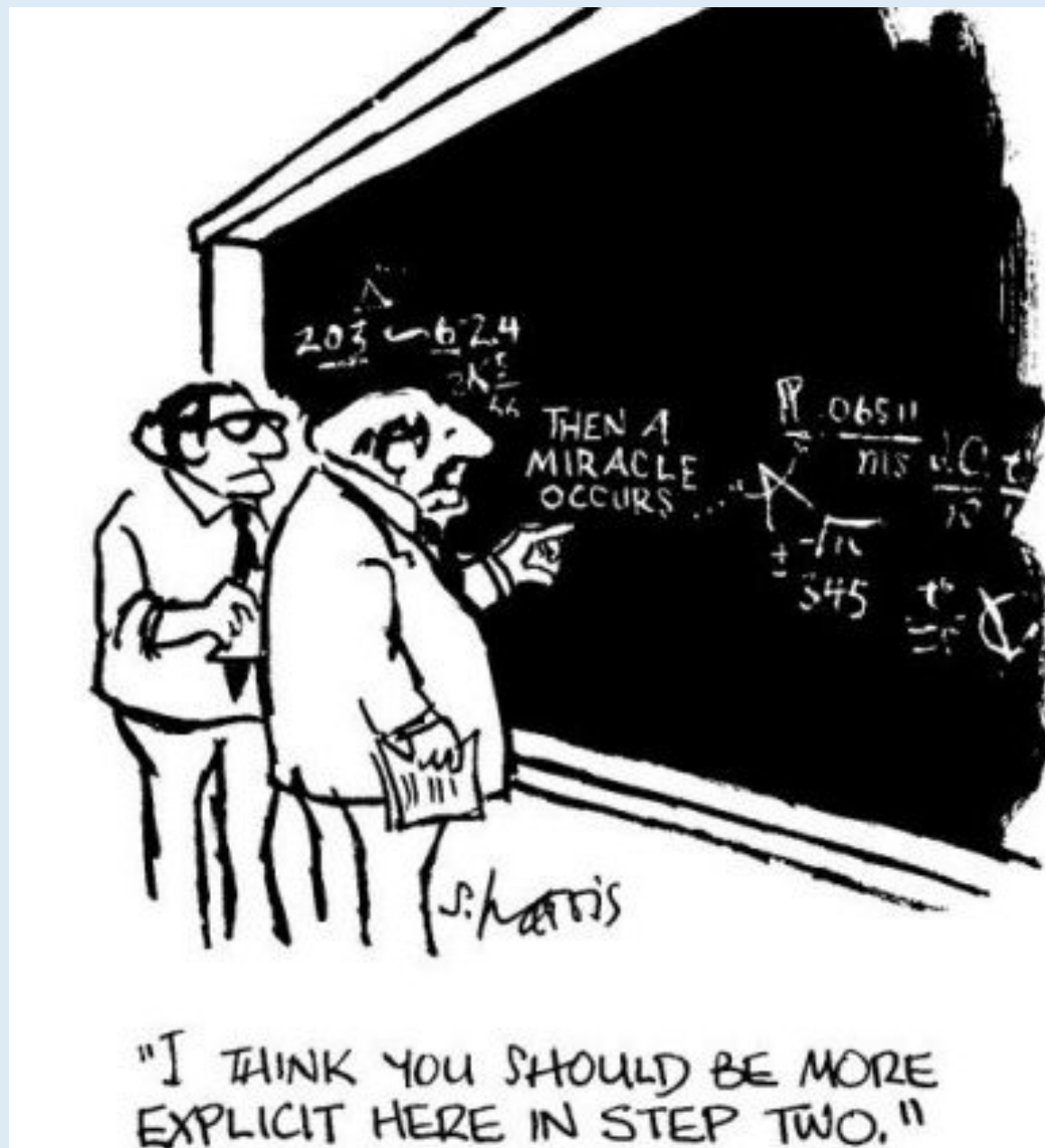
d Dev (ug/L)

0.086

DL ($s \cdot t_{\alpha=0.01, df=6}$)

0.27





Curves with Positive x-Intercepts – Measurement Quality Considerations

Concentration below the x-intercept can be calculated with the curve, unless the data system recognizes no signal and software returns 'ndetect'.

Responses close to the x-intercept can change by orders of magnitude without much change in concentration

When concentrations of interest are near or below the LOQ, use care in establishing integration parameters and signal thresholds, otherwise low responses that produce (biased) blank concentrations potentially near the LOQ will be ignored

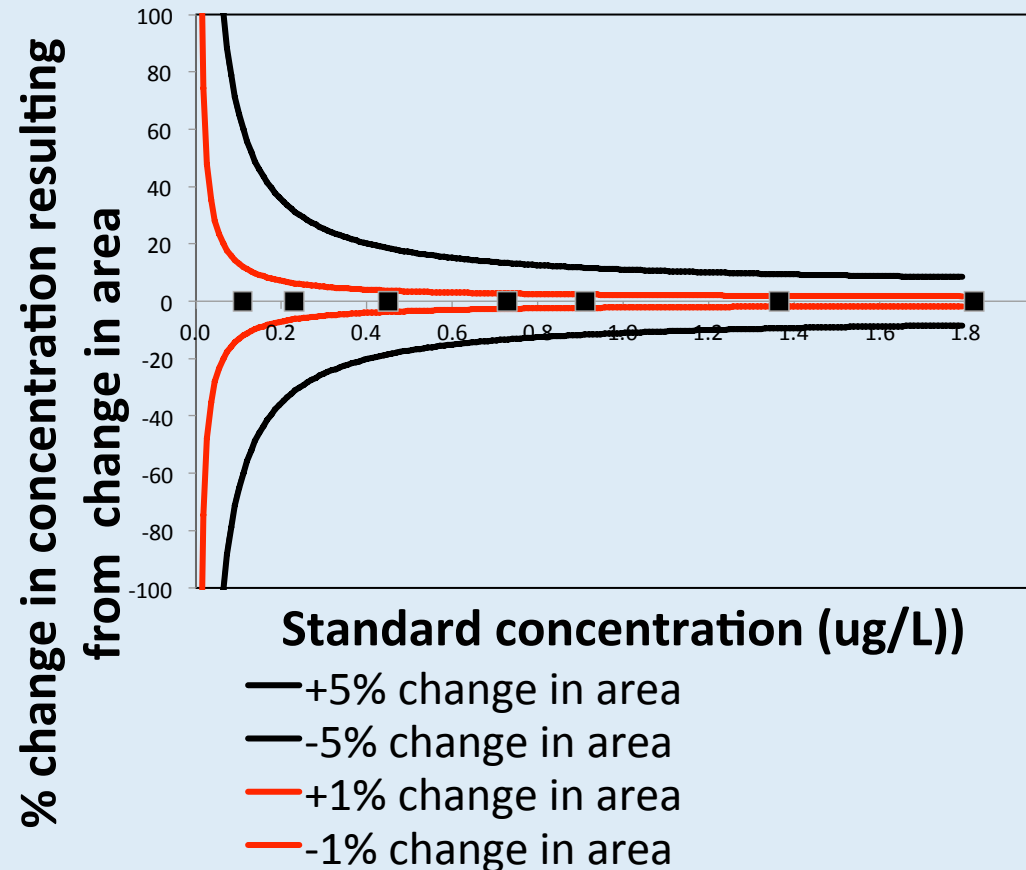
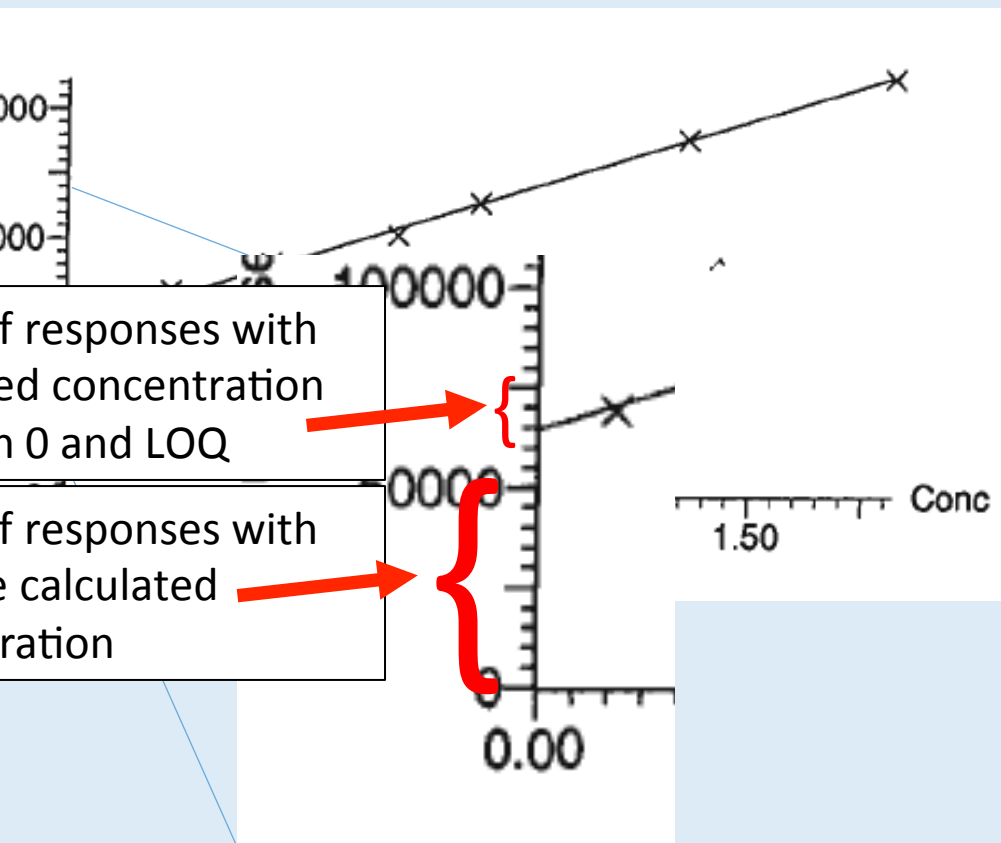
Curves with Positive x-Intercepts – Measurement Quality Considerations (cont.)

Calibrating to a lower concentration and weighting regression tend to push the x-intercept or the regression line closer to the origin, in turn decreasing positive bias in calculated concentration for extrapolated responses (i.e., method blanks)

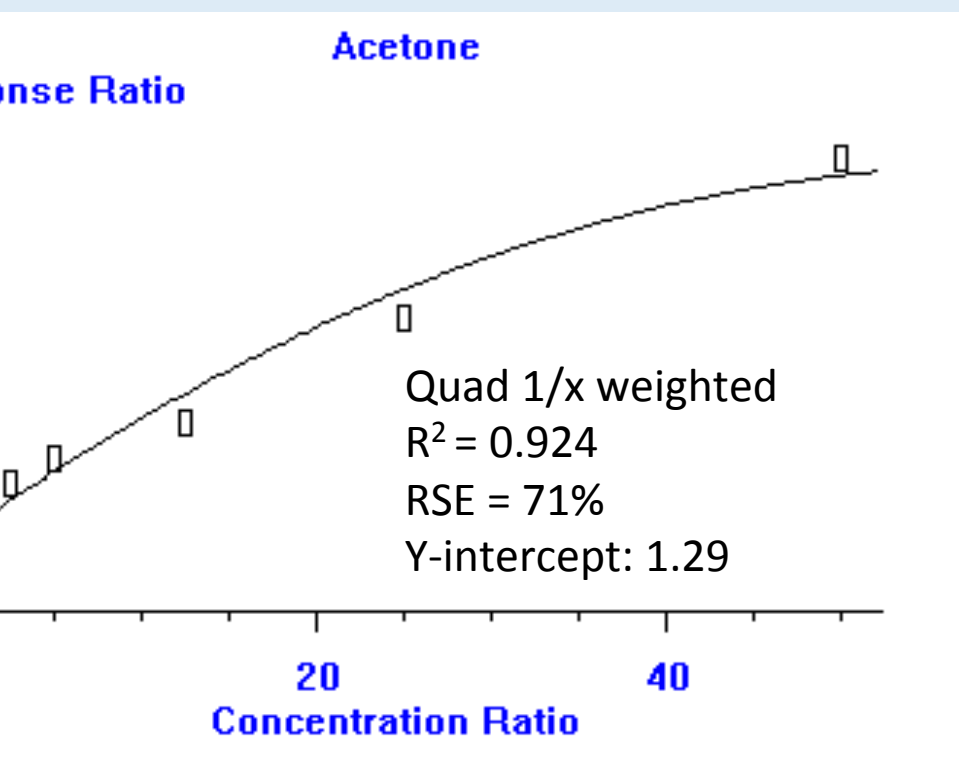
Response is useful for data evaluation (e.g., comparing blanks are to samples

Curves with Positive y-intercepts

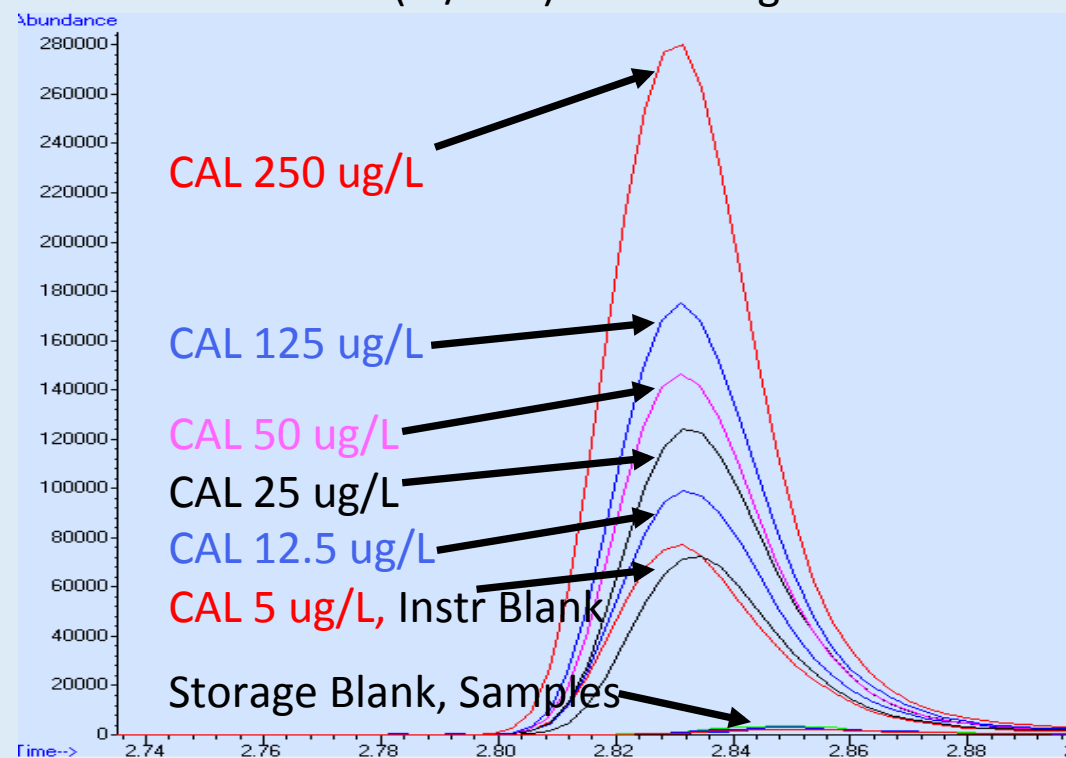
Octylphenol 12-ethoxylate by LC/MS/MS
(y-intercept is 93% of low standard response)



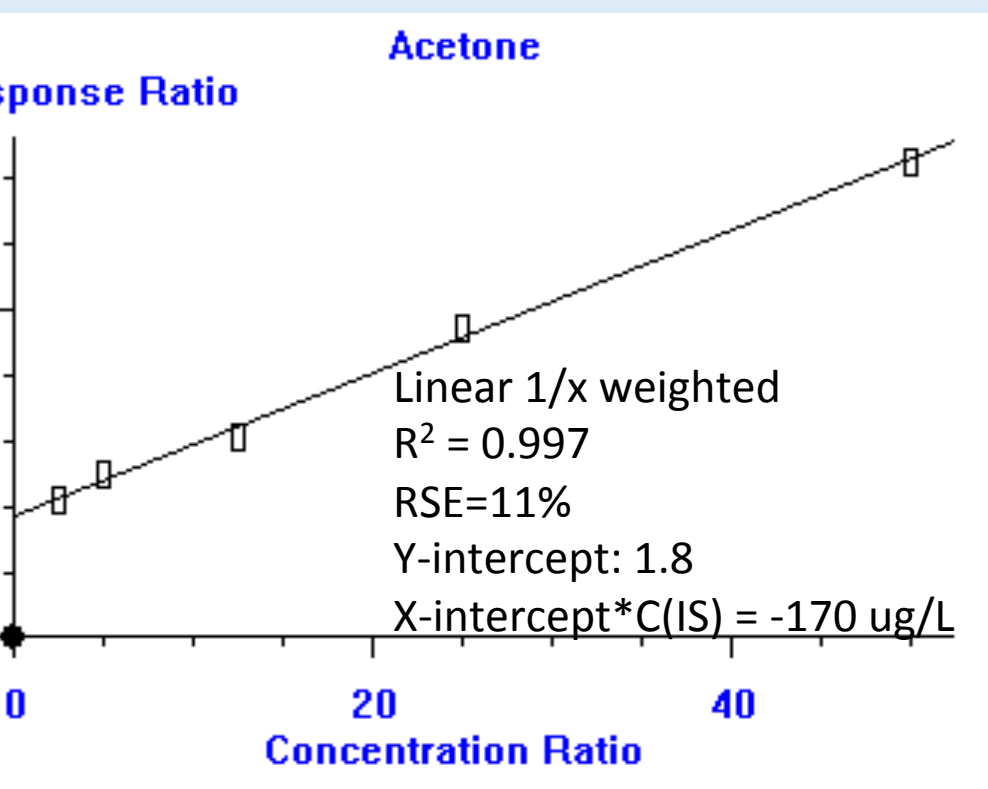
Curves with Positive Y-intercepts: Acetone in water by 5030/8260



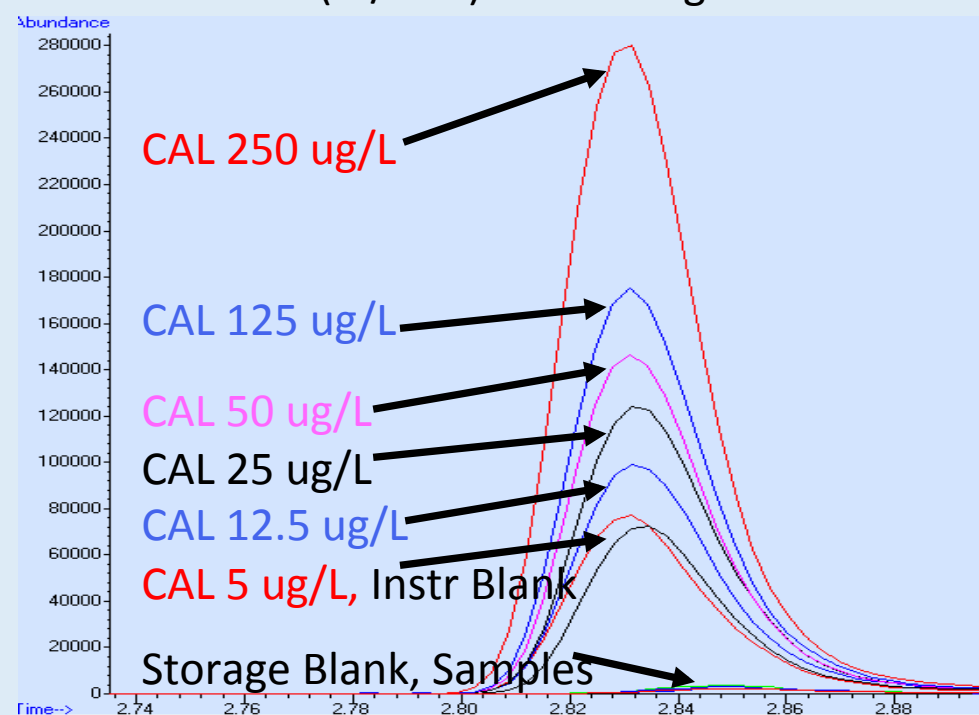
Extracted ion (m/z 43) chromatogram



Curves with Positive Y-intercepts: Acetone in water by 5030/8260

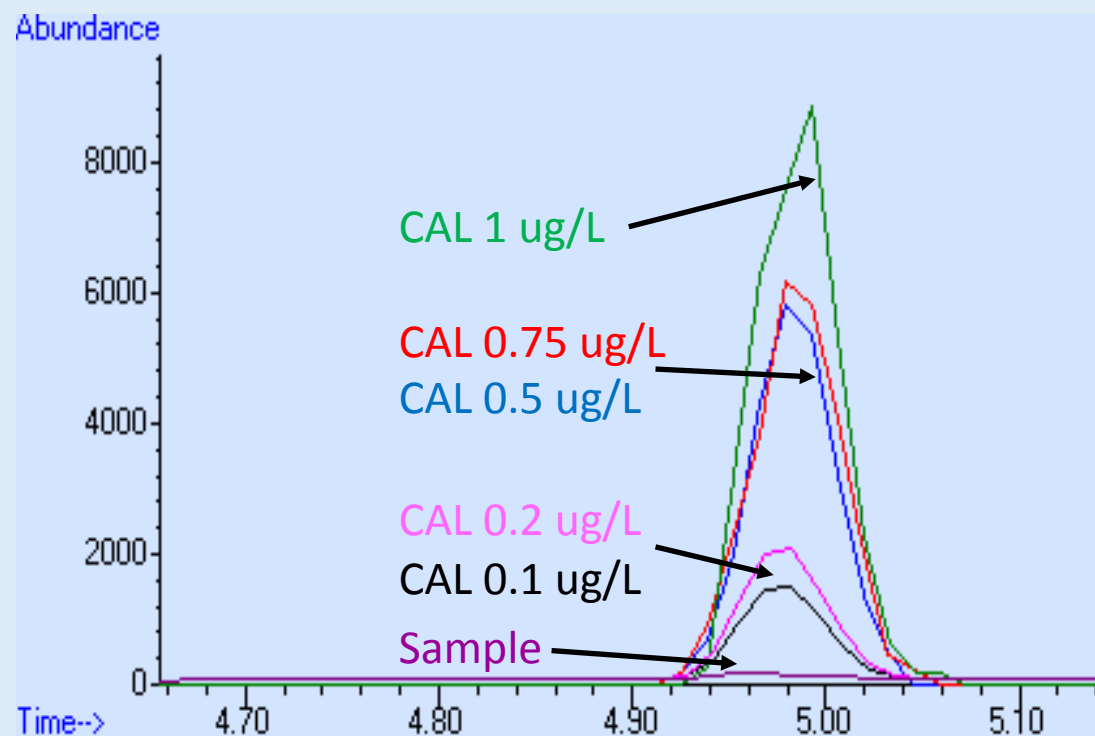
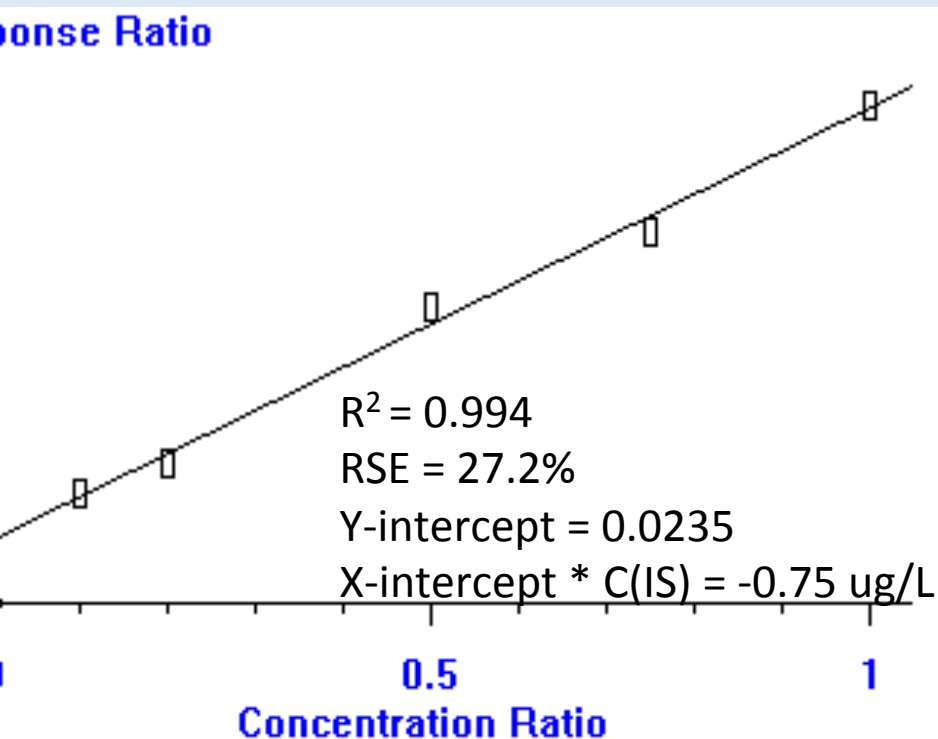


Extracted ion (m/z 43) chromatogram



Curves with positive Y-intercepts: Vinyl chloride water by 5030/8260 (Single Quad SIM)

Extracted ion m/z 62 response



Positive Y-Intercepts – Measurement Quality Considerations

Background present in calibration standards can be calibrated out, which may create a measurement bias problem. Matrix spikes can reveal this bias, but depends on matrix spike level relative to the LOQ

When necessary for data application, limit bias by raising effective LOQ to a calibration standard level clearly distinguishable from the y-intercept.

For example, raising LOQ to lowest calibration standard with response $>$ twice the y-intercept of calibration function limits bias at the LOQ to factor of 2 (true concentration of 10 = measured concentration of 5)

Carefully evaluate sources of background, and minimize any sources associated with standards that are not also in samples, if possible

Use response instead of or in addition to concentration for data evaluation (e.g., comparing blanks to samples).

Other Approach: FDA Method for Preparation and MS/MS Analysis of Honey for Fluoroquinolone Residues (enrofloxacin, ciprofloxacin)

External standard calibration in blank honey matrix

Linear calibration model, not forced through zero.

If needed, a $1/x$ weighting may be used to more accurately quantitate low concentrations. Acceptable curves have correlation coefficients of 0.99 or greater.

Calibration range: 2.5-50 ng/g

Concentration limit: 5 ng/g

< 2.5 ng/g reported as non-detect

2.5 ng/g - 5.0 ng/g reported as positively identified, but below quantification limit

≥ 5.0 ng/g reported as a numerical concentration value

<http://www.fda.gov/Food/FoodScienceResearch/LaboratoryMethods/ucm071495.htm>

Recommendation to instrument software developers

Proximity of calibration intercept (x or y) to low calibration standard concentration or response is not always apparent

Need: a software option that lets analysts calculate and display the proximity of the LOQ standard response or concentration to the intercept of calibration function

With this data, we could track proximity of LOQ standard to X-intercept or Y-intercept of calibration function over time, which would provide an indication of trends in measurement bias near the LOQ that might otherwise be hard to identify

Conclusions

For calibration models not forced through the origin, the intercept can be an important indicator of the potential for measurement bias near the LOQ.

Background in reagents and standards used for instrument calibration and instrument sensitivity can vary by over time, and the cause is not always apparent.

Evaluating the calibration intercept and comparing responses of blanks, samples and standards during data evaluation will result in more defensible decisions about how to address non-linear behavior near the LOQ.

Conclusions (cont.)

When calibration fit is acceptable, calibration models forced through the origin avoid sticky problems associated with the intercepts

Concentration estimates extrapolated outside the calibration range should be used with caution due to potential for measurement bias.

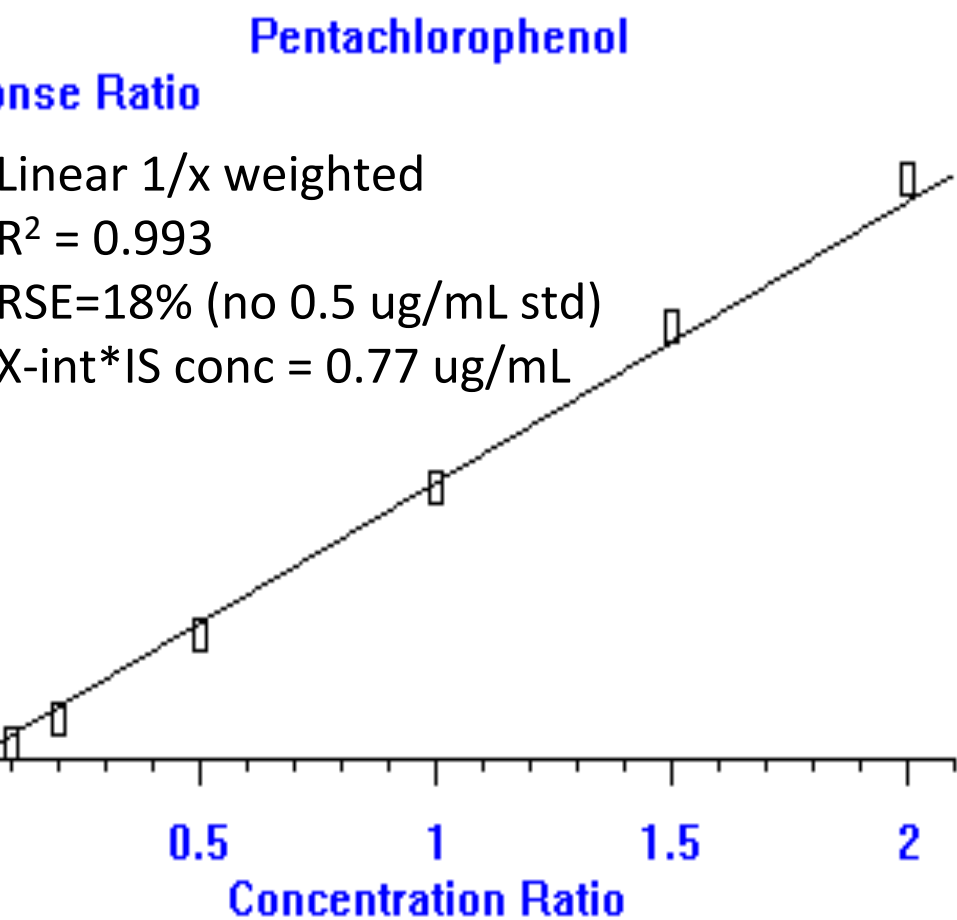
However, even for curves with non-zero intercepts, as long as target analytes responses are not close to the intercept (i.e., where the relationship between response and concentration is proportional), concentration estimates below the LOQ may still be useful.

Most important: Define data quality needs first. Then consider how measurement bias or uncertainty near the LOQ may impact them.

Thanks for listening!

sitive x-intercept: Pentachlorophenol by 8270

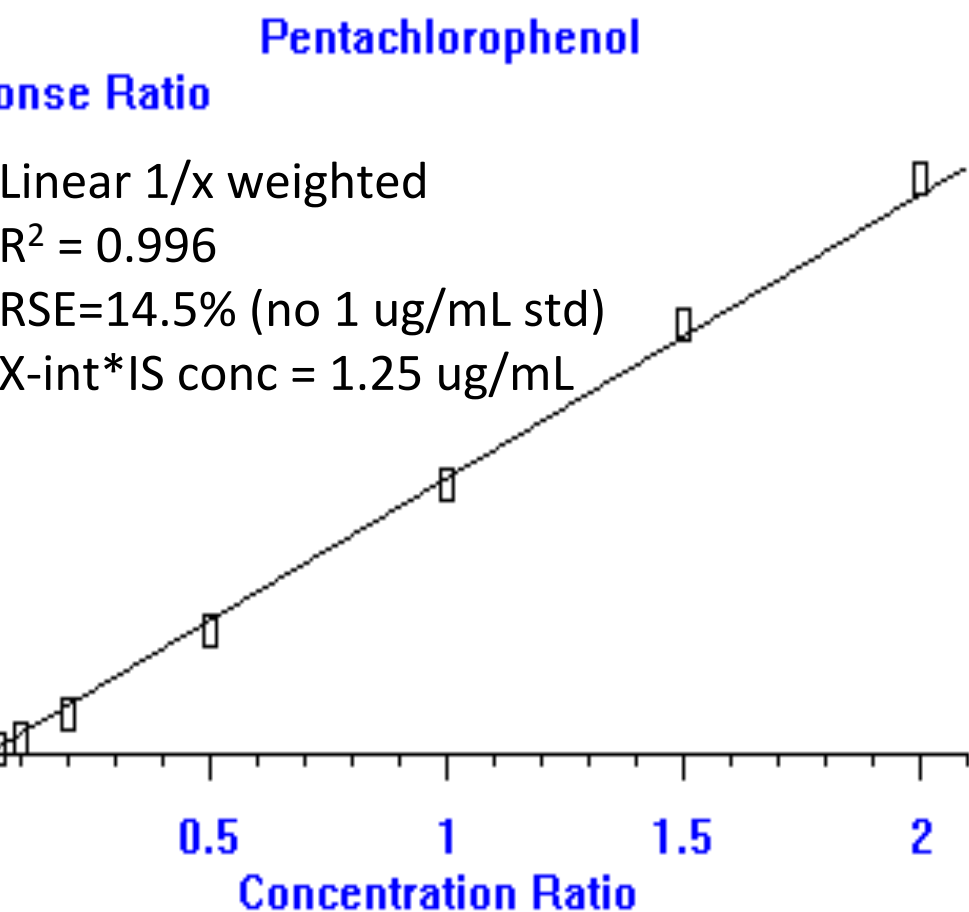
Calibration Range: 0.5-100 ug/mL



Conc (ug/mL)	Calc RRF	Calc conc (ug/mL)	% e
0.5	0.031	0.9	8
1	0.031	1.0	4
2.5	0.056	2.0	-2
5	0.076	4.1	-1
10	0.093	8.9	-1
25	0.119	26.7	6
50	0.128	56.8	13
75	0.137	90.7	2
100	0.134	118	17

sitive X-intercept: Pentachlorophenol by 8270

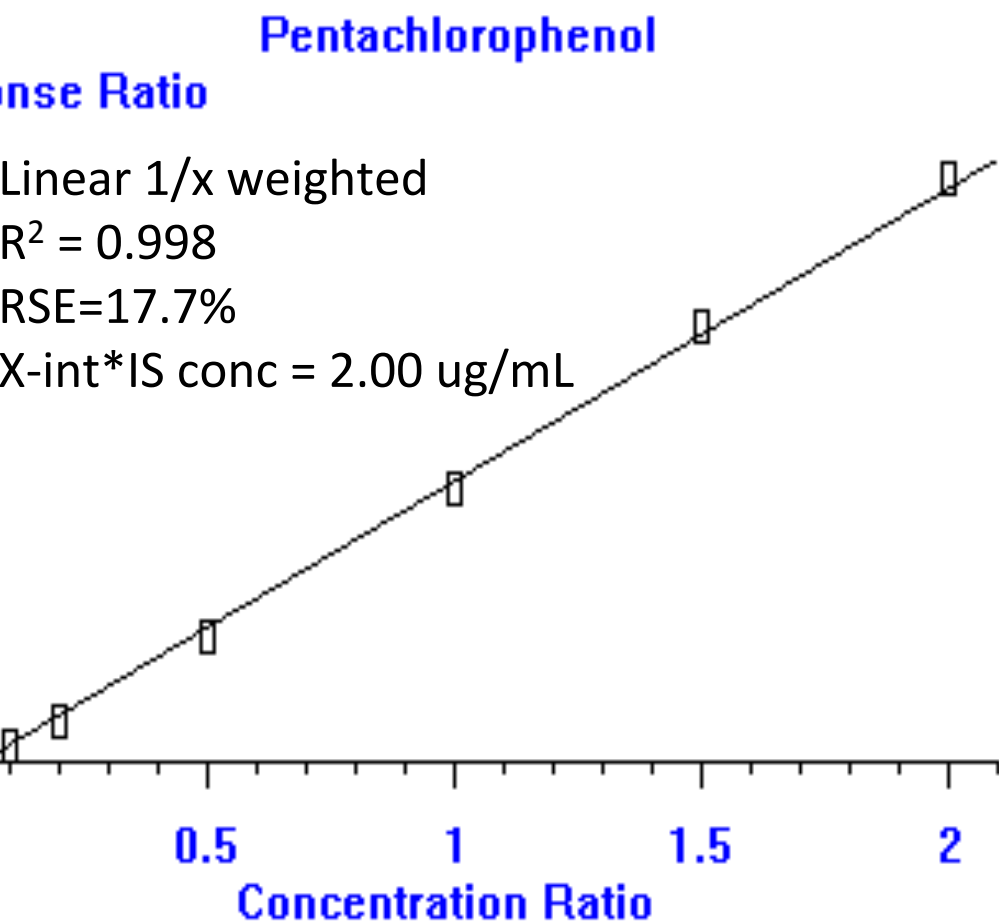
Calibration Range : 1-100 ug/mL



Conc (ug/mL)	Calc RRF	Calc conc (ug/mL)	% e
0.5	0.031	1.38	17
1	0.031	1.52	5
2.5	0.056	2.45	-
5	0.076	4.51	-9
10	0.093	9.30	-
25	0.119	26.8	7
50	0.128	56.5	1
75	0.137	90.0	2
100	0.134	117	1

sitive X-intercept: Pentachlorophenol by 8270

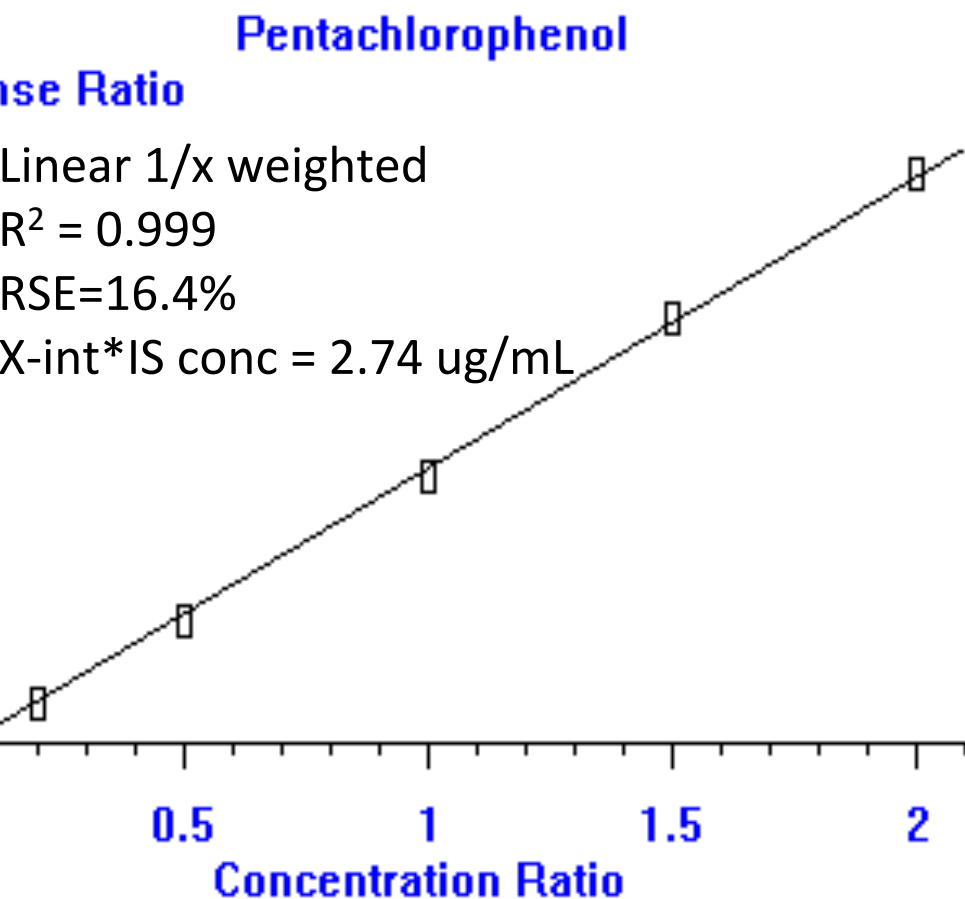
Calibration Range : 2.5-100 ug/mL



Conc (ug/mL)	Calc RRF	Calc conc (ug/mL)	% e
0.5	0.031	2.1	32
1	0.031	2.3	12
2.5	0.056	3.2	2
5	0.076	5.2	3
10	0.093	9.9	-
25	0.119	27.1	8
50	0.128	56.1	1
75	0.137	89.0	1
100	0.134	115	1

sitive X-intercept: Pentachlorophenol by 8270

Calibration Range : 5-100 ug/mL

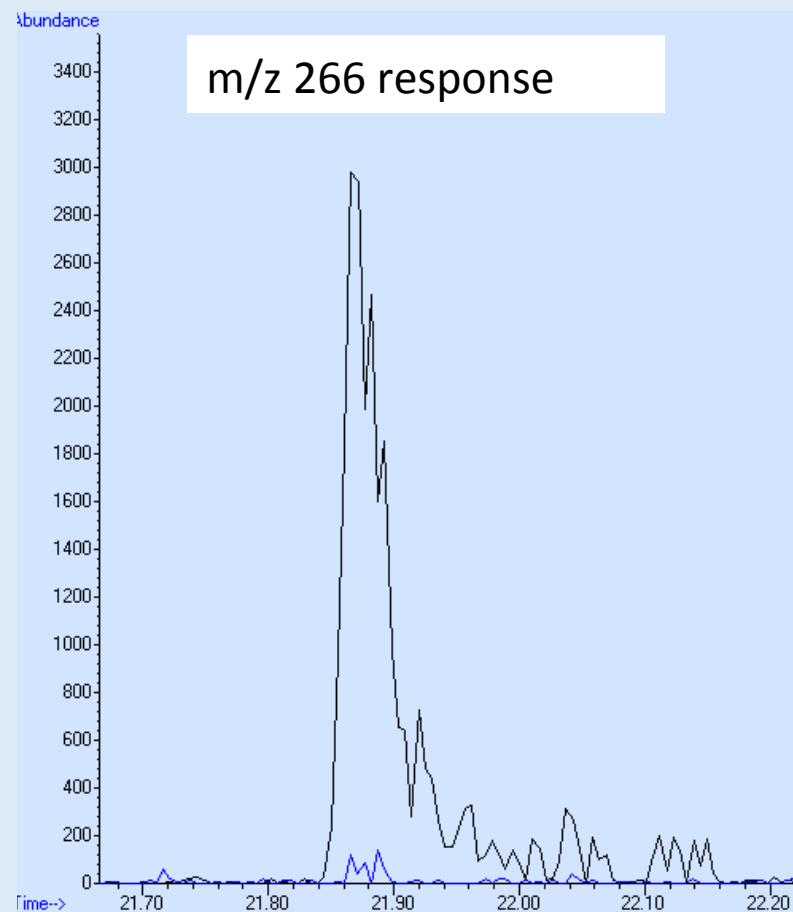


Conc (ug/mL)	Calc RRF	Calc conc (ug/mL)	% e
0.5	0.031	2.9	47
1	0.031	3.0	20
2.5	0.056	3.9	5
5	0.076	5.9	1
10	0.093	10.5	5
25	0.119	27.4	9
50	0.128	56.0	1
75	0.137	88.3	1
100	0.134	113.9	1

Positive x-intercept: Pentachlorophenol by 8270D

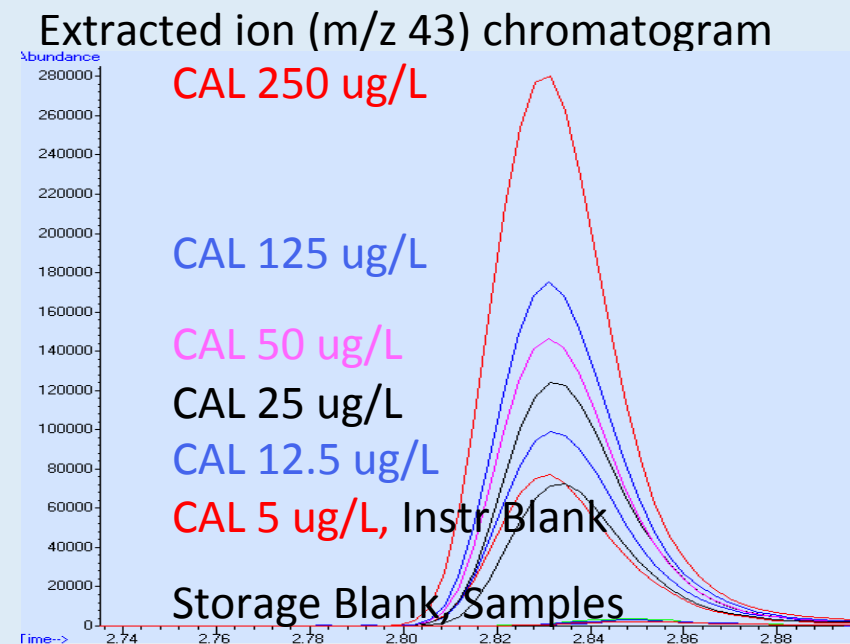
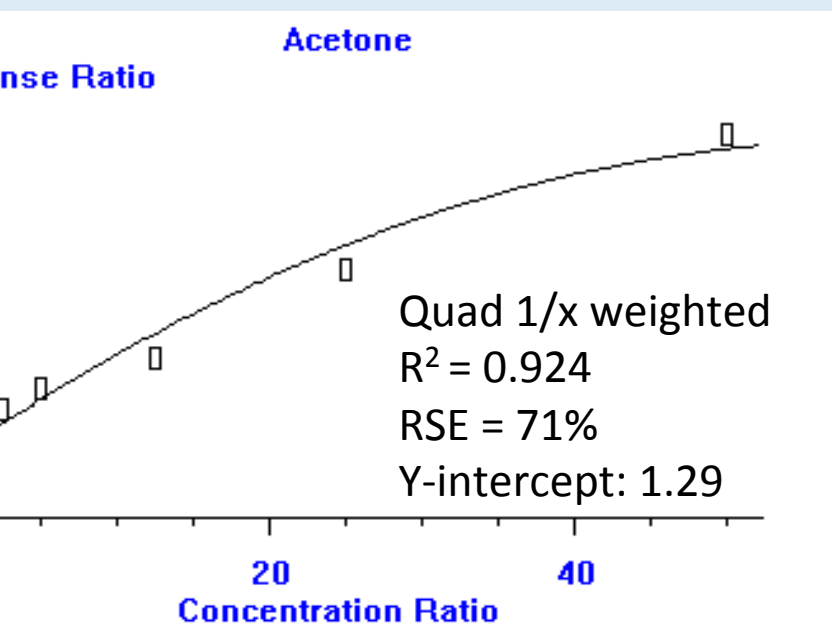
Concentration (ug/mL)	LLOQ standard* conc (ug/mL)	Blank conc (ug/mL, at area of 149)	Blank area / low standard area
100	1	0.80	24%
10	2.5	1.28	14%
100	2.5	2.03	2.1%
10	5	2.77	0.7%

LLOQ: Lower Limit of Quantitation, set as lowest standard whose calculated concentration was within 10% of expected. From SW-846 method 8000D.



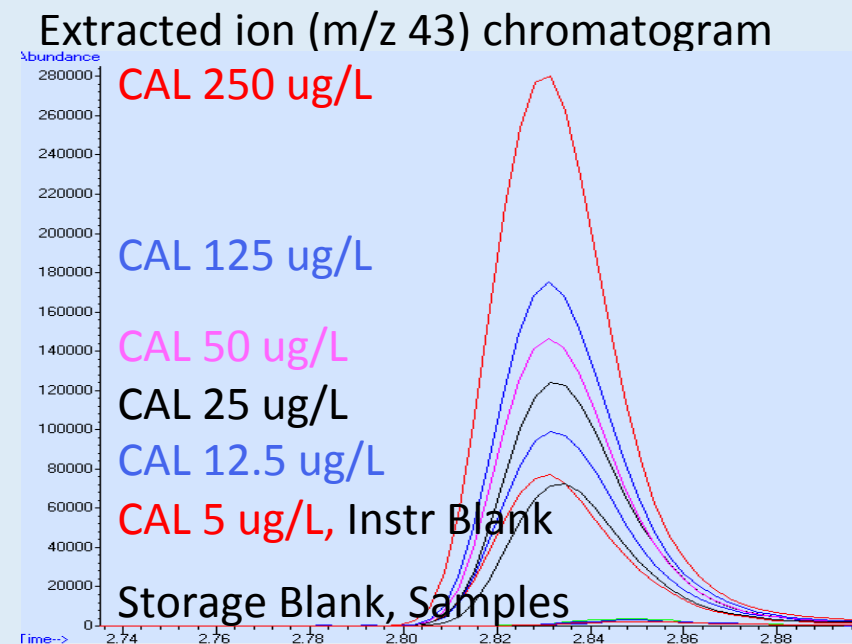
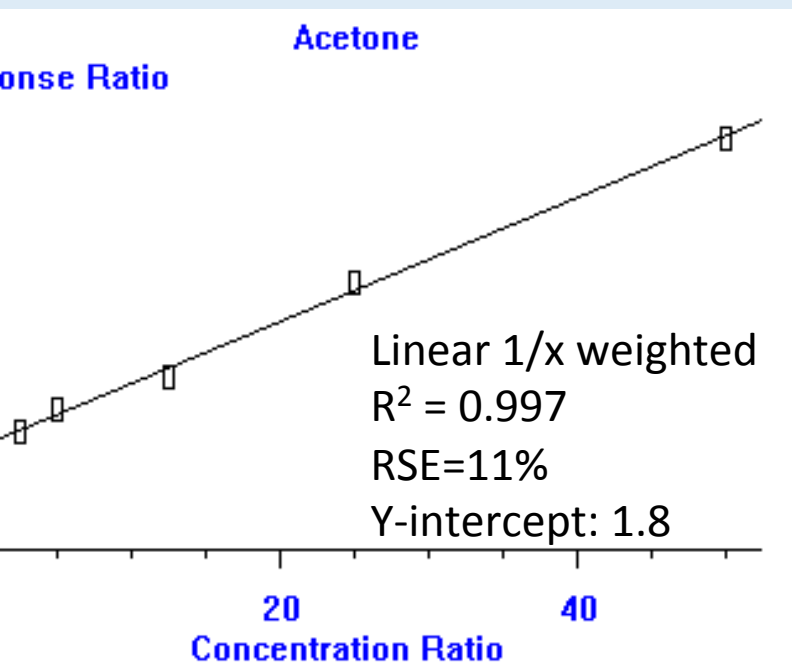
Black: 2.5 ug/mL cal standard
Blue: method blank

Positive Y-intercept: Acetone in water by 5030/8260



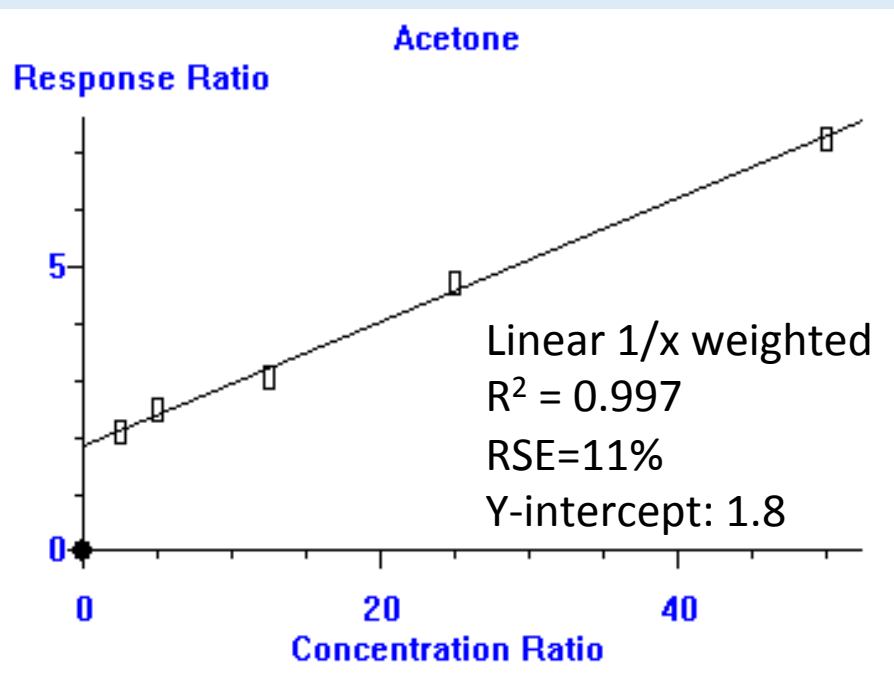
	Acetone (m/z 43) Area	Fluorobenzene (m/z 96) area	Response ratio	Calc (µg/L)	% of expected
5 ug/L	1435462	1134699	1.27	-1.4	
12.5 ug/L	1886403	1122258	1.68	20.4	163.1
25 ug/L	2343751	1126067	2.08	42.2	168.7
50 ug/L	2711673	1099304	2.47	64.0	127.9
125 ug/L	3221382	1065216	3.02	97.2	77.7
250 ug/L	5139976	1095717	4.69	213.1	85.2
500 ug/L	7685080	1063448	7.23	undefined	

Positive Y-intercept: Acetone in water by 5030/8260

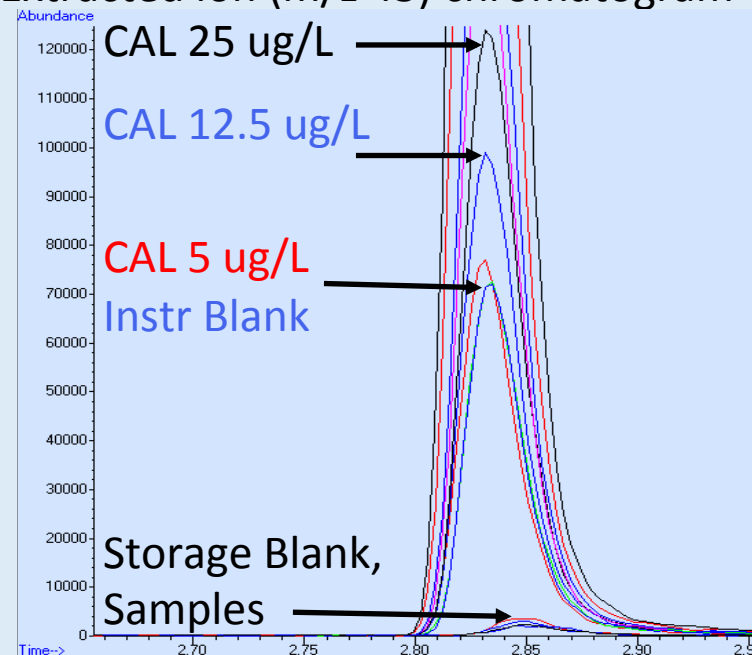


	Acetone (m/z 43) Area	Fluorobenzene (m/z 96) area	Response ratio	Calc (µg/L)	% of expected
5 ug/L	1435462	1134699	1.27	-52.8	
12.5 ug/L	1886403	1122258	1.68	-14.5	
25 ug/L	2343751	1126067	2.08	22.4	89.7
50 ug/L	2711673	1099304	2.47	57.9	115.9
125 ug/L	3221382	1065216	3.02	109.3	87.5
250 ug/L	5139976	1095717	4.69	262.9	105.2
500 ug/L	7685080	1063448	7.23	496.6	99.3

Positive Y-Intercept: Acetone in water by 5030/8260



Extracted ion (m/z 43) chromatogram



	Acetone (m/z 43) Area	Fluorobenzene (m/z 96) area	Response ratio	Calc (µg/L) Quad 1/x	Calc (µg/L) Linear 1/x
25 ug/L	2343751	1126067	2.08	42.2	22.4
ment Blank	1373321	1090711	1.26	-1.7	-53.4
age Blank	80798	1006630	0.08	-59.5	-162
mple 1	67660	936560	0.07	-59.9	-163
mple 2	48775	941101	0.05	-60.8	-165
mple 3	46067	889486	0.05	-60.8	-165

Positive Y-Intercept: Acetone in water by 5030/8260

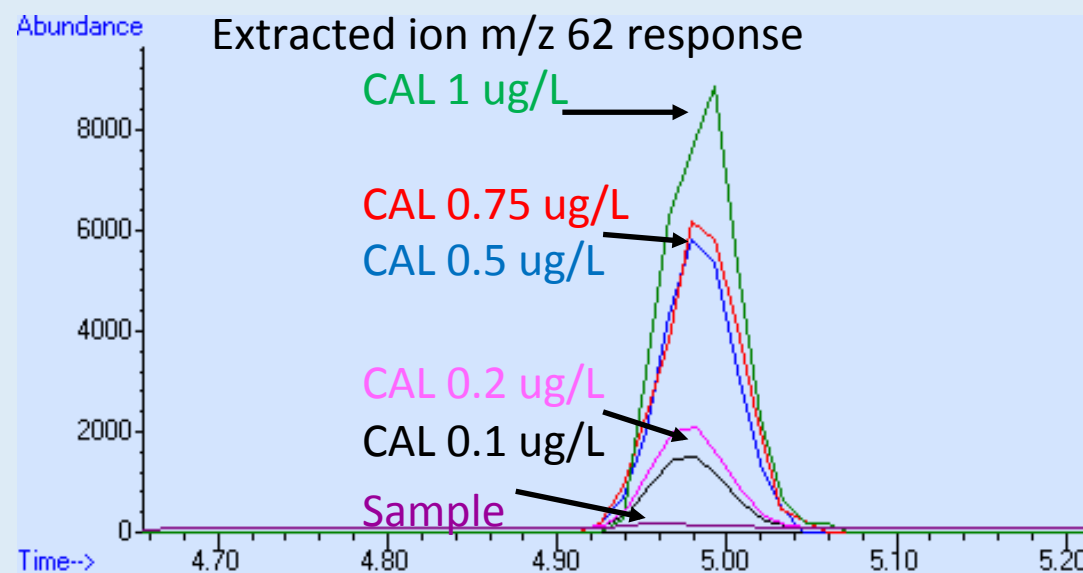
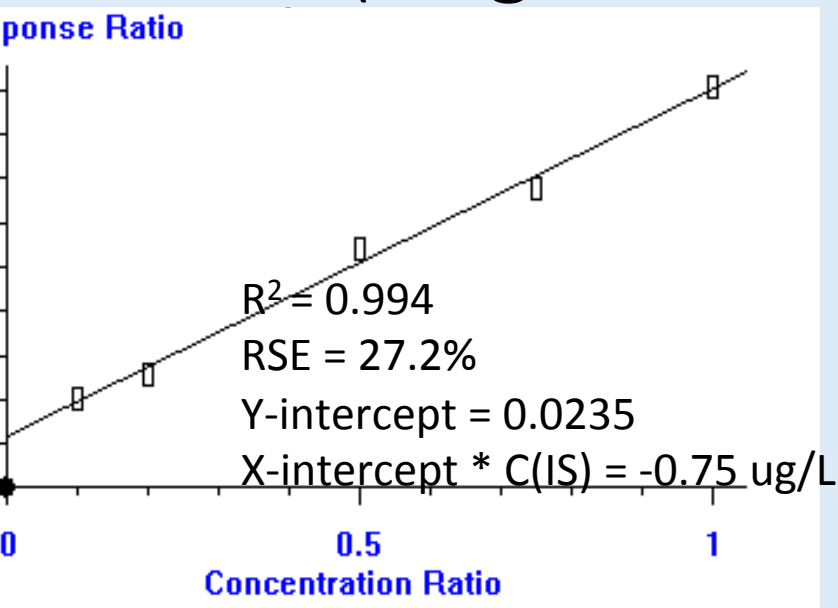
Added acetone sample (ug/L)	Actual (including background) (ug/L)	10% variability (Meas \pm 10% True)	Best case for true value		Worse case for true	
			% of Measured (no variability)	% of Measured (10% variability)	% of Measured (no variability)	% of M (10% va
0	170	-17 - 17		-	-	
25	195	5.5 - 44.5	100%	45-178%	13%	3-2
125	295	95.5 - 128	100%	76-124%	42%	32-
500	670	433 - 567	100%	86-113%	75%	65-

Using method of standard additions, $-(x \text{ intercept}) = \text{Calculated acetone background in the calibration standards} = \sim 170 \text{ ug/L}$

Best case: Background in the calibration standards is also present in samples.

Worst case: Background in calibration standards is absent from sample

sitive Y-intercept: Vinyl chloride in water by 30/8260 (Single Quad SIM)



	Vinyl Chloride m/z 62 Area	Chloroethane-d5 m/z 69 area	Relative response	Calc (µg/L)	% of expected
CAL 0.1 ug/L	4512	117771	0.038	0.09	94%
CAL 0.2 ug/L	5526	126996	0.044	0.09	43%
CAL 0.5 ug/L	12844	120084	0.107	0.53	107%
CAL 0.75 ug/L	20846	149072	0.140	0.74	99%
CAL 1.0 ug/L	24426	132886	0.184	1.03	103%
Sample 1	311	116725	0.003	-0.13	

tive Y-Intercept: Vinyl Chloride in water by 5030/8

Measured chloride le (ug/)	Actual (including std addition backgrnd) (ug/L)	Measured, with 10% Variability in Response (Meas ±10% Actual)	Best case		Worst case	
			% of Measured (no variability) (=meas/meas)	% measured (with 10% variability) =(meas)±10%(actual)	% of Measured (no variability) =(meas/actual)	% meas (with 10% v =meas/act (actu
	0.75	-0.075-0.075		-	-	-
1	0.85	0.02-0.17	100%	15-185%	12%	1.8-2
2	0.95	0.11-0.29	100%	53-148%	21%	11-3
5	1.25	0.38-0.66	100%	75-125%	40%	30-5
	1.75	0.83-1.31	100%	83-118%	57%	47-7

Calculated equivalent background in calibration standards (based on method of standard additions) = 0.75 ug/L

Best case: Background in the calibration standards is also present in samples.

Worst case: Background in calibration standards is absent from samples.