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ABSTRACT

The compounds 2-Methylisoborneol (2-MIB) and Geosmin are the primary source of the foul odor found in drinking water. Algal contamination is the principal cause of the formation of these compounds. Geosmin and 2-MIB have such a low odor threshold, that even the slightest amount can produce an unpleasant odor and taste in drinking water. Thus, developing a reliable sampling and analysis platform for very low level detection is important. Standard Method 6040D describes a procedure for the detection of 2-MIB and Geosmin using Solid Phase Micro Extraction (SPME) coupled with a Gas Chromatograph (GC) and Mass Spectrometer (MS). Selective Ion Monitoring (SIM) is used for compound detection down to part per trillion (ppt) levels. This examination will optimize the sampling and detection of 2-MIB and Geosmin.

INTRODUCTION

It has been found that the presence of blue green algae in water sources produces 2-MIB and Geosmin. Both Geosmin and 2-MIB are malodorous compounds that emit a musty earthy aroma. When the algae generates an abundance of these compounds in a drinking water reservoir, there are resulting taste and odor problems. Algae blooms are influenced by their climate, as a result, the formation of Geosmin and 2-MIB is more of a problem during the summer months and in warmer climates. Areas in the southwestern portion of the United States seem to have the most problems with 2-MIB and Geosmin.

Drinking waters are tested in order to determine water quality for prospective consumers. Two of the major complaints that water suppliers need to address are issues with taste and odor. Geosmin and 2-methylisoborneol (2-MIB), although non-toxic, both have very strong odors and can be detected at levels below 10ppt. Thus, developing a reliable sampling and analysis platform is important.

Standard Method 6040D addresses the sampling and analysis of 2-MIB and Geosmin utilizing SPME in conjunction with GCMS in Single Ion Monitoring (SIM) mode. The SPME fiber is placed in the headspace above the sample for a period of time and then the fiber is inserted into the GC injection port for analyte desorbtion onto the GC column. This application will develop and optimize Method 6040D experimental parameters for the determination of 2-MIB and Geosmin down to 5ppt.

EXPERIMENTAL

Headspace SPME is a non-exhaustive sampling technique so the experimental conditions required optimization in order to make the extraction technique both efficient and reproducible. For the automation of the sampling process, the EST Analytical FLEX autosampler was used. The FLEX suite software simplified the sample method development process with the ease of its drag and drop method builder.

The most efficient SPME fiber for this analysis was a Divinylbenzene/ Carboxen/Polydimethylsiloxane (DVB/CAR/PDMS) coated fiber with a 50/30µm film thickness. The Shimadzu QP2010 SE GCMS was run in SIM mode and was fitted with a SPME liner and a Restek Rxi-5 Sil MS column. The autosampler and GCMS experimental conditions developed for this analysis are listed in Tables 1 and 2.

Table 1: FLEX Autosampler Experimental Parameters

Method Type Gc Ready Gc Cycle Tim Constant Heat

Incubation T Incubation T

Fiber Guide D Sample Vial I **Extraction Til** Fiber Extracti Agitation Type Agitation Dela Agitation Du

Wait On Inpu Wait Input

Injection Port Fiber Guide S Fiber Guide D **Fiber Insertio Fiber Insertio** Fiber Desorbti **Injection Star**

Determination of 2-Methylisoborneol and Geosmin in Water Using Solid Phase Micro Extraction

itosampler	Flex
	General
е	SPME
	Continue
ne	21min
at Mode	Yes/Continue
Sample	Incubate Agitate
emp.	65°c
īme	1.0min
	Extraction
Depth	45%
Fiber Depth	1Cm
ime	30.1min
tion Agitate	Yes
pe	Oscillate
elay	0.1min
iration	30.0min
	Wait
ut	Yes
	GC Ready
D	esorbtion
rt	A
Speed	40%
Depth	50%
on Speed	75%
on Depth	1cm
otion Time	3min
art Output	Start

Table 2: GC/MS Experimental Parameters

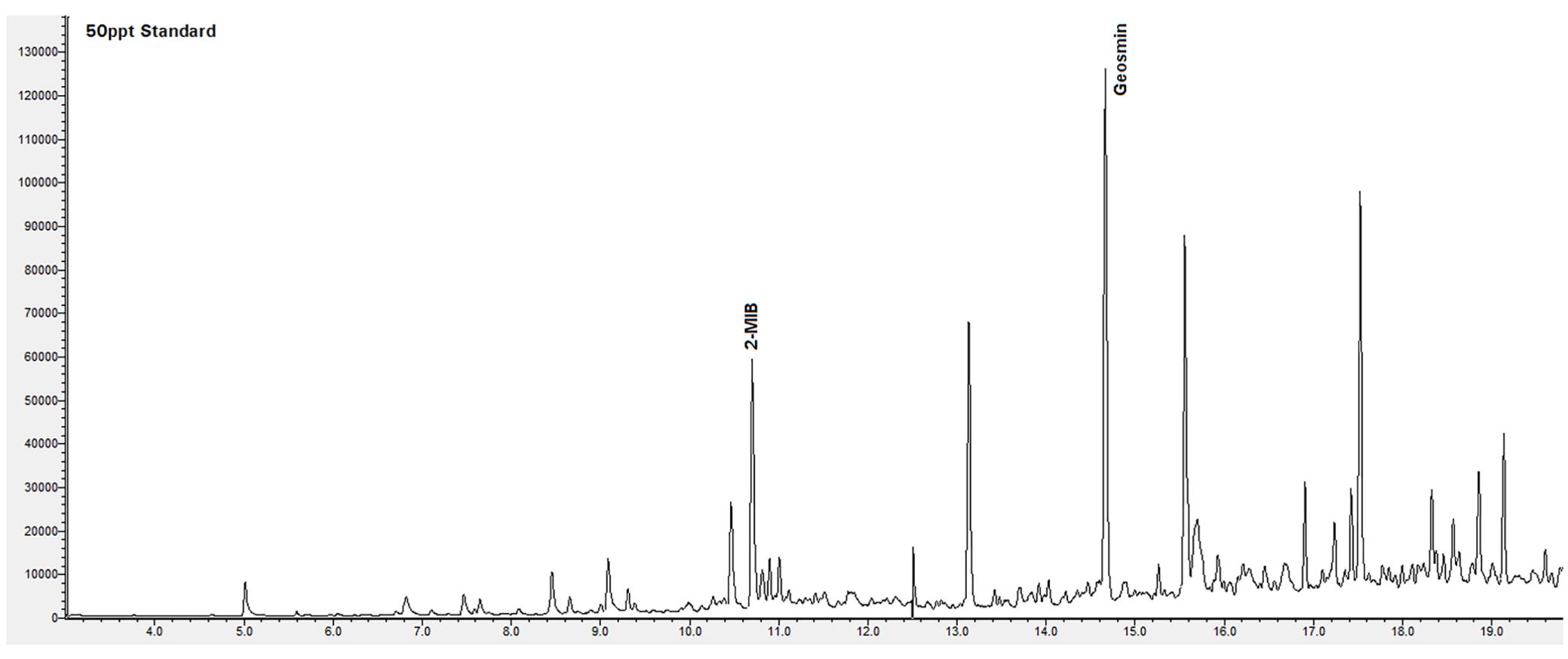
GC/MS	Shimadzu QP 2010 SE			
Inlet	Split/Splitless			
Inlet Temp.	270°C			
Inlet Head Pressure	40.7kPa			
Mode	Splitless			
Injection Pulse Pressure	50kPa for 2.0 min			
Carrier Gas Split Ratio	2			
Desorption	3.0min at 270°C			
Column	Rxi-5 Sil MS 30.0m X 0.25mm X 0.25µm			
Oven Temp. Program	60°C hold for 2.0 min., ramp 8°C/min to 200°C, hold for 0.5min, 20min run time			
Column Flow Rate	0.8ml/min			
Gas	Helium			
Linear Velocity	32.6ml/min			
Source Temp.	220°C			
MS Transfer Line Temp.	300°C			
Acquisition Mode	SIM			
SIM lons 3.01 to 12.50min	95, 107, 108			
SIM lons 12.51 to 20.00min	112, 125, 126			
Event Time	0.30sec			
Solvent Cut Time	3.0min			

The 2-MIB and Geosmin standard was ordered from Supelco while Sodium Chloride was purchased from Sigma Aldrich. A six point standard curve was prepared in water with a range of 5 to 100ppt. Ten milliliters of each curve standard was added to a prepared 20ml headspace vial and sealed. The prepared headspace vials contained 2.5g of Sodium Chloride. A linear curve was attained for each analyte, see Figure 1. After the curve was established, seven replicate samples of the 5ppt and the 50ppt standards were run in order to establish method detection limits and precision and accuracy data. Table 3 displays the experimental results and Figures 2 and 3 show chromatograms of both the 5 and 50ppt standards.

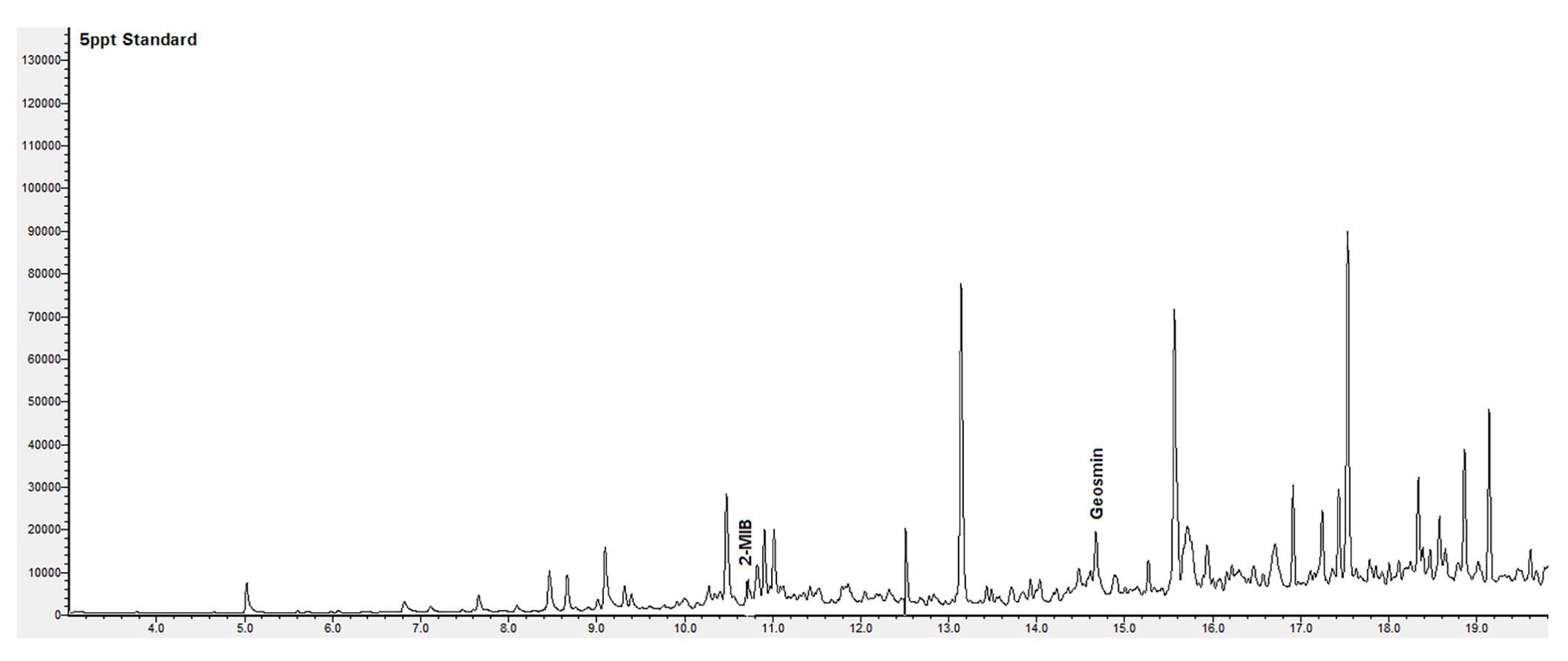
Table 3: Experimental Results Summary

Compound	Curve %RSD	Curve R ²	MDL (5ppt)	Precision (5ppt) %RSD	Accuracy (5ppt)	Precision (50ppt) %RSD	Accuracy (50ppt)
2-MIB	12.36	0.999	2.14	16.22	83.94	10.39	95.03
Geosmin	11.46	1.000	1.06	7.07	95.22	5.40	101.85

Figure 2: Chromatogram of 50ppt Standard







Conclusion

The SPME analysis of 2-MIB and Geosmin more than met the method requirements of Standard Method 6040D. The curve linearity from 5 to 100ppt had an R² of 0.999 or greater and a %RSD of better than 12.5. The resulting precision at 5ppt was about 16% for 2-MIB and 7% for Geosmin while the recoveries were 84% for 2-MIB and 95% for Geosmin. At 50ppt both compounds displayed improved precision and accuracy. Geosmin had 5.4% precision and 102% recovery while 2-MIB had 10.4% precision and 95% recovery. Once the optimum parameters were established the FLEX autosampler proved to be an exceptional system for the SPME sampling of drinking water samples.

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