Keeping Up with Changing PCR Methods

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PROBLEM STATEMENT

- Growth based methods for monitoring recreational water too slow to protect swimmers
 - 18 to 72 hour incubation
 - Delay getting results leads to 70% error rate at California beaches

• EPA-approved QPCR method is faster

- Very sensitive to environmental interferences that inhibit PCR chemistry
- Highly variable, reliant on standard curve for quantification
- Travel time to lab means results not out until late afternoon

Digtal PCR more precise, less sensitive to inhibition

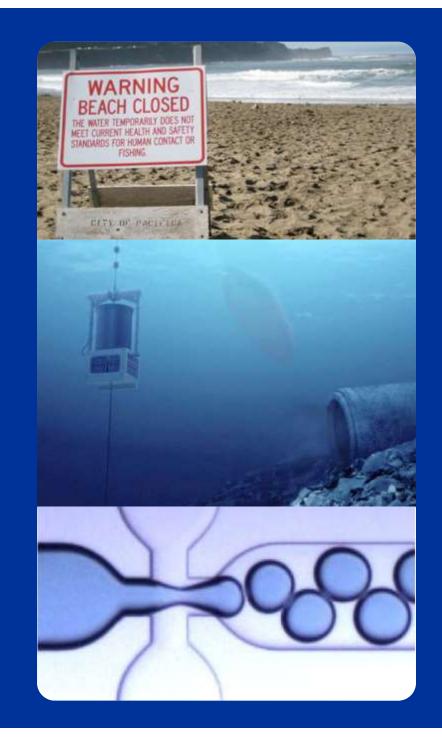
- Travel time still an impediment to timely notification of health risk

Need to begin processing water samples in the field to produce a meaningful answer

- Publicly disseminated before swimmers enter contaminated water

OUTLINE

- Current state of qPCR for water quality monitoring
- Proof of concept study for automating water quality monitoring
- Introduction to digital PCR
- Development of an automated dPCR instrument



QPCR HAS COME A LONG WAY

15 years of development

Validation in multiple epidemiology studies

Two EPA-approved methods for *Enterococcus*

- Methods 1611 and 1609
- E. coli method in use in Great Lakes

ARE WE DONE?

We have EPA-approved qPCR methods

Have trained 14 labs in California on Method 1609

– Using method routinely

Sample collection delays timely analysis

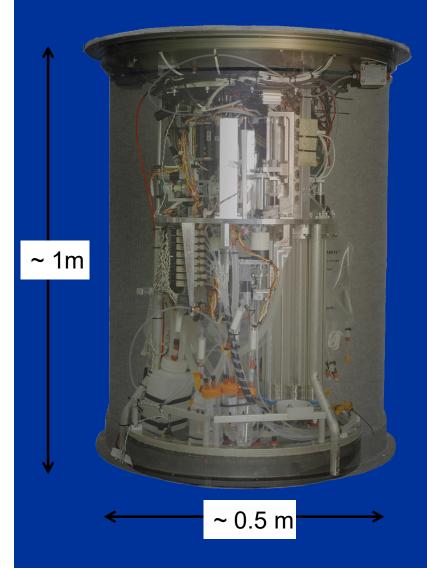
- Requires additional personnel for meaningful answer

WHERE DO WE GO FROM HERE?

Automation can help provide answers early in the day

 New digital PCR technology is more accurate and less susceptible to inhibition

2ND GENERATION ENVIRONMENTAL SAMPLE PROCESSOR (ESP)



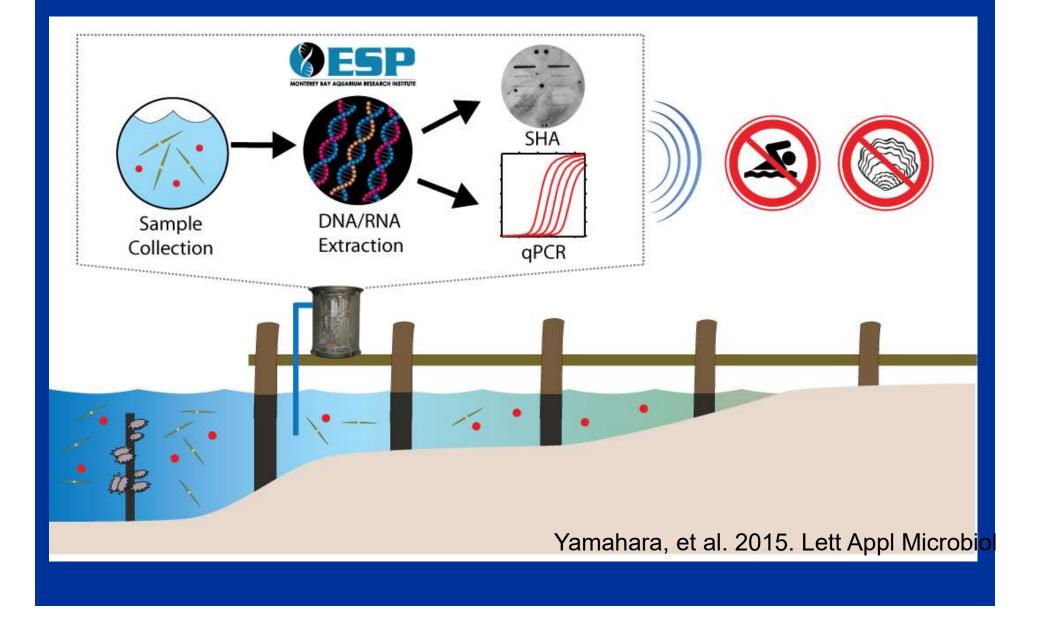
Collection

Concentration

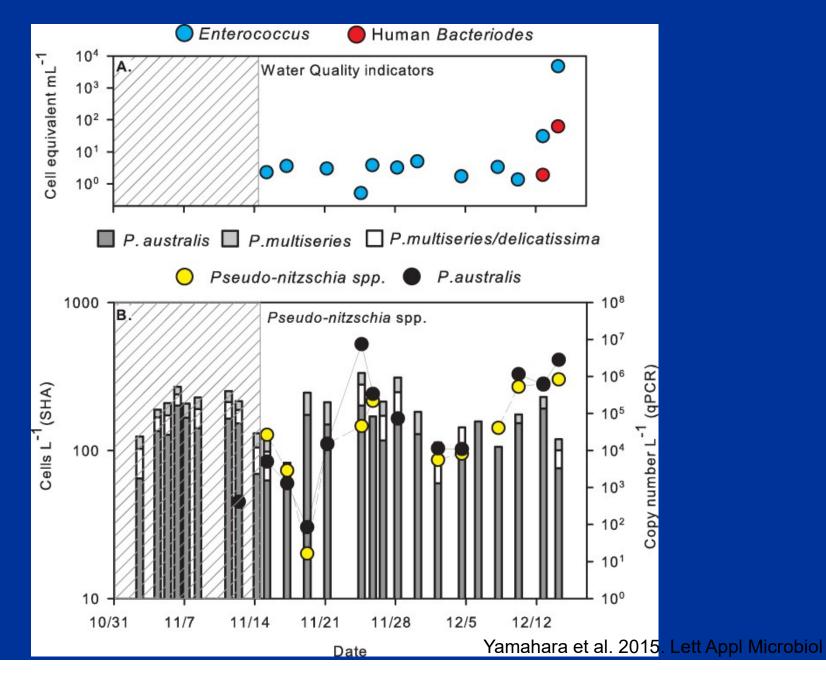
Extraction

Detection

PROOF OF CONCEPT



SANTA CRUZ WHARF, CA, DEPLOYMENT



CRITIQUE OF 2ND GENERATION ESP

Pros-

- Quantification of BOTH fecal indicators and harmful algae from the same sample
 - Sample to results in 4 hours

Cons-

Extremely complicated

- 1980's technology
- Requires highly skilled technicians to maintain and operate
- qPCR very susceptible to environmental interferences

Not portable

- Similar in size to a 55 gallon drum
- Weighs about 300 lbs.

QPCR VS. DIGITAL PCR

LIMITATIONS OF QPCR

Low precision when target concentration low

Susceptible to inhibition

May result in underestimation and false negatives

Difficult to implement cost-saving strategies

– Often difficult to measure multiple targets simultaneously in one reaction

→ Digital PCR has the potential to overcome these limitations!

CHARACTERISTICS OF DIGITAL PCR

- Direct quantification of target by counting positive droplets
 - No standard curve needed
- Can provide precision estimate even with one reaction

- More resistant to inhibition
- Simultaneous measurement of multiple targets in one reaction

HOW DROPLET DIGITAL PCR WORKS

• "MPN" PCR

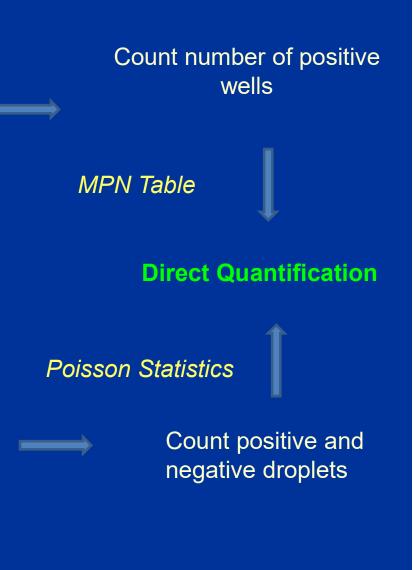


100ml water



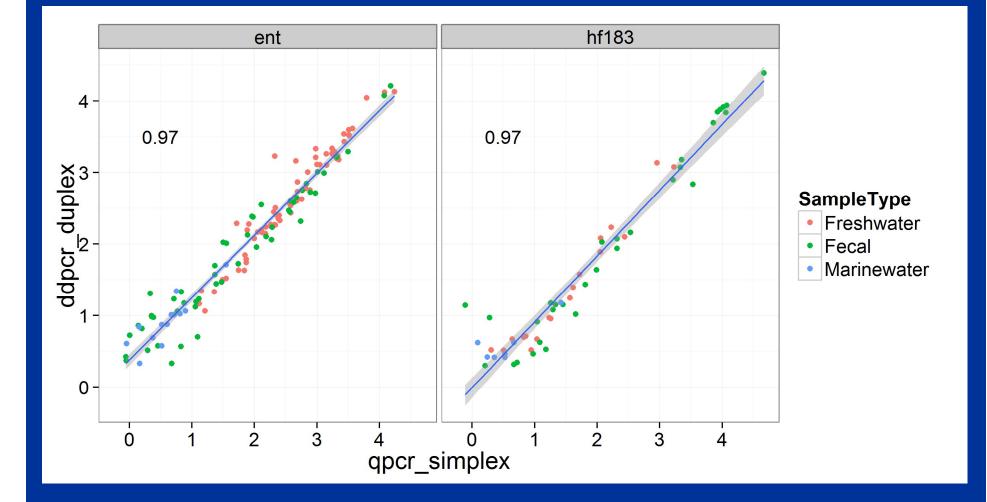
96 wells

 \leq 20,000 droplets



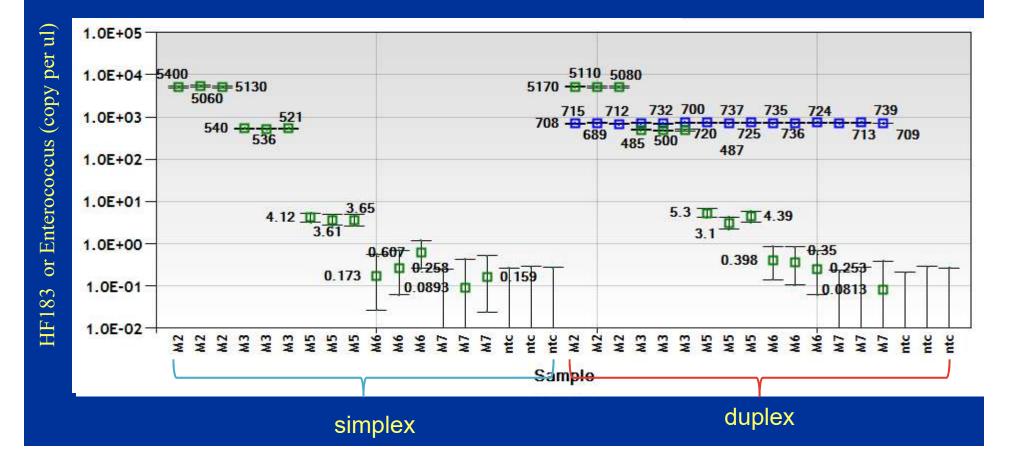
20ul PCR (tube/well)

HIGHLY CORRELATED WITH EPA APPROVED QPCR METHOD FOR ENTEROCOCCUS



MEASURING ENTEROCOCCUS AND HF183 SIMULTANEOUSLY

 Quantification of *Enterococcus* (blue squares) and HF183 (green squares) are not affected by each other



INHIBITION

More robust

- qPCR signal disappears with increased inhibitor concentration
- ddPCR signal remains nearly constant

Humic acid concentration (ng/ul)	qPCR (HF183 copy/rxn)	ddPCR (HF183 copy/rxn)
0	1810	1810
1	1165	1680
2.5	184	1700
5	0	1870

WISH LIST FOR AUTOMATED DPCR INSTRUMENT

Fast

Results telemetered to decision makers in < 4 hours of sampling event

• Flexible

- Able to detect multiple targets (indicators, pathogens, source markers)

Easy to operate

- Able to be operated by a field technician or lifeguard

Portable

- Vehicle mounted or hand carried to sampling sites

Reliable

- Not susceptible to environmental interferences
- Robust

NEW INSTRUMENT DESIGN CRITERIA



• Detecting and Tracking sources of contamination requires mobility

- Engineering design for a hand-carry instrument
- Modular design separate sample collection and detection

SAMPLE COLLECTION/PROCESSING

Sample Collection/ Processing





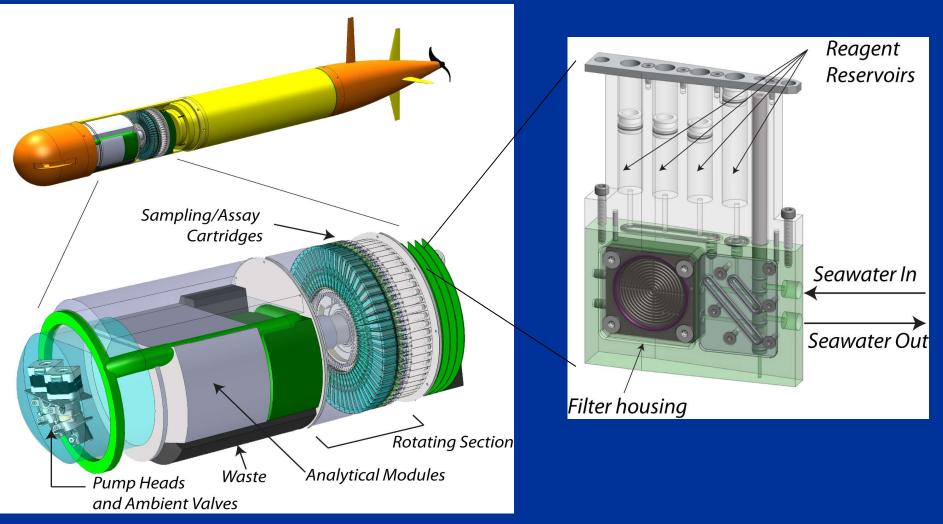


Analyte Detection





3RD GENERATION ESP SOLUTION



Same engineering concepts, different form factor

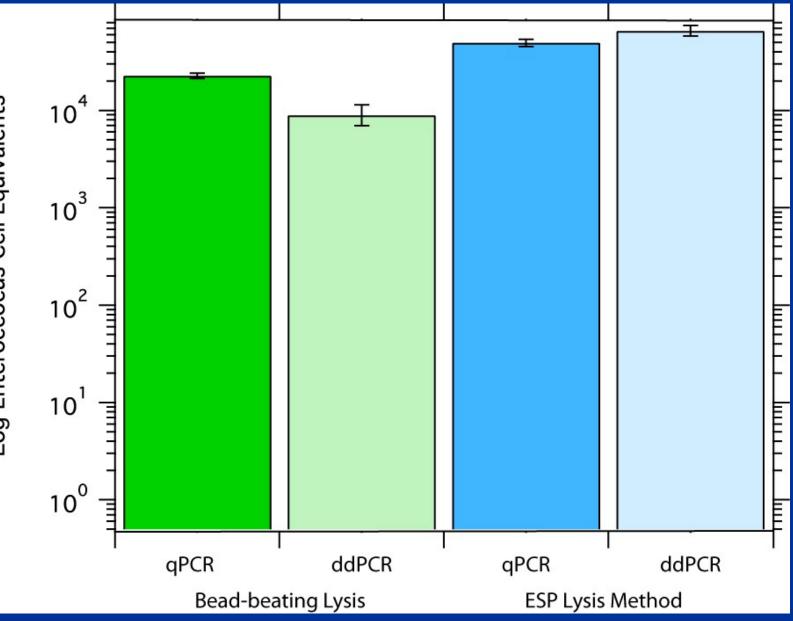
PROTOTYPE 3RD GENERATION ESP

• 3rd Generation (3G) ESP technology

- Sample Collection and Processing
 - Preservation and In-situ Lysis
- Digital PCR (ddPCR)



ESP DNA EXTRACTION COMPARISON



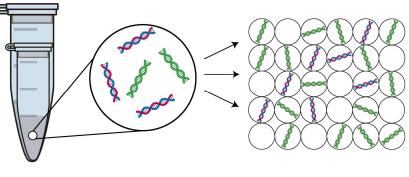
-og Enteroccocus Cell Equivalents

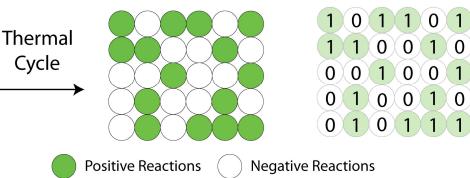
ANALYTE DETECTION



ASU DROPLET DIGITAL PCR MODULE

Partition a normal PCR reaction with many DNA templates into many individual PCR reactions Digital readout of positive and negative reactions provides an absolute quantification

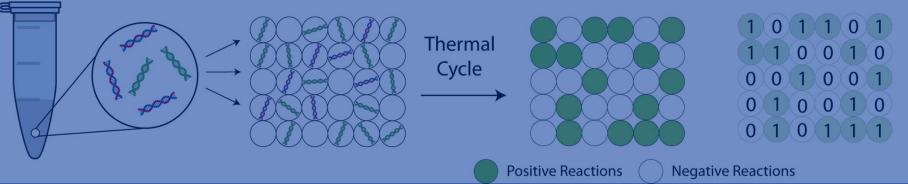




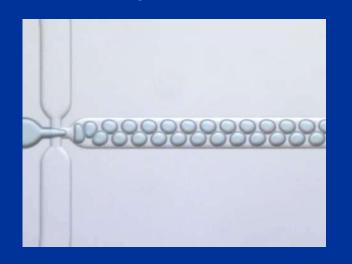


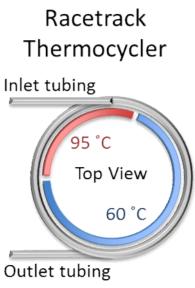
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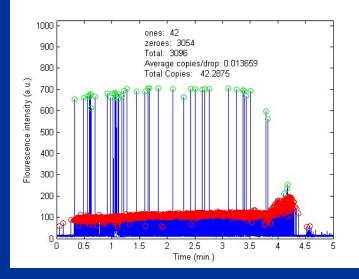


Partitioning to 1-nL Reactions

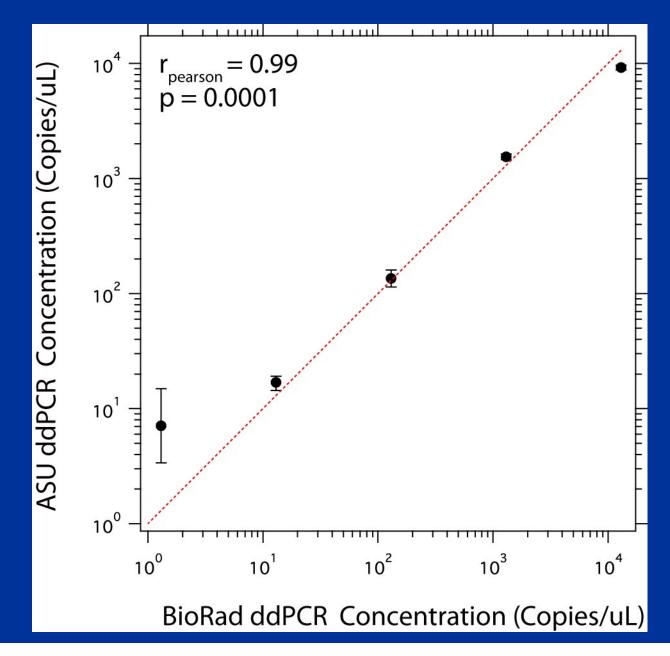




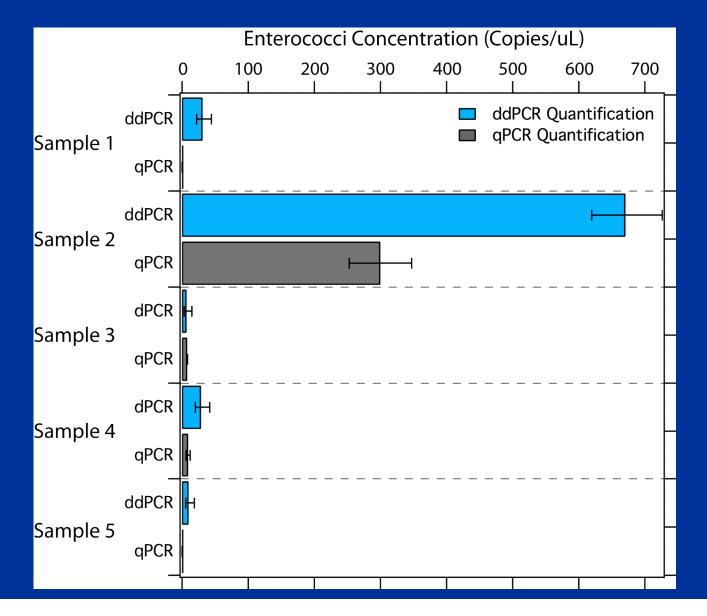
Digital Positive & Negative Droplets



DDPCR QUANTIFICATION OF ENTEROCOCCUS



QUANTIFICATION OF ENVIRONMENTAL SAMPLES USING ESP METHODS



CONCLUSIONS AND NEXT STEPS

- The challenge of portable biological sensors for water quality monitoring is sample acquisition and processing for downstream analyses
- Modular microfluidic design makes this technology extremely adaptable to new applications
 - Shape form extremely compact and malleable
 - Currently exploring drinking and reclaimed water applications
- Field sampling trials will commence in late 2016 or early 2017

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