## **BHIMADZU**

# Rapid Quantification of Perfluorinated Compounds in Drinking and Surface Water Using LC-MS/MS

Jeremy Post, Christopher Gilles, William Lipps Shimadzu Scientific Instruments, Columbia, MD

Novel Aspect

Fast, low chemical background, high sensitivity measurement of perfluorinated compounds in water.

#### Introduction

Perfluorinated compounds (PFCs) are synthetic molecules with carbon chains to which fluorine is bound by high strength carbon-fluorine covalent bonds. Useful due to their surfactant and hydrophobic physicochemical properties these materials are used broadly since the 1950s in industrial applications such as the conveyance of stain and stick resistance to a wide variety of materials and products including clothing, cookware, carpet, and food containers. These molecules, now classified as persistent organic pollutants, are highly stable and resistant to biological degredation, therefore they persist in the environment, bioaccumulate, and due to their ready transport by water are found even in remote regions of Earth. As contaminants of emerging concern, research on PFCs is ongoing to determine the impacts of these materials on human health and the environment. A CDC study detected four of these analytes in >98% of serum samples representative of the general US population above age twelve. Low level analysis of these compounds is challenging due to their ubiquitous presence in the laboratory and analytical instrumentation components



Figure 1: Central Maryland surface water source locations

> Figure 2: Direct method surface water sampling collection technique.



### Method

Surface water samples were collected and stored according to document SOP #EH-01 Surface Water Collection using the Direct Method. As the mobile phase transfer lines, degasser membrane, pump inlet block, and sample vial septa all utilize PTFE, made from a prominent PFC, perfluorooctanoic acid (PFOA), these components were either exchanged or bypassed to eliminate all sources of extraneous analyte from the instrumentation. The calibration curve solutions were prepared in glass vials using glass syringes with a metal plunger. PFC standards were purchased from Wellington Laboratories. A Shimadzu LCMS-8050 was used for analyte separation and mass spectrometry. Analytes were ionized using negative mode electrospray ionization. All samples were passed through 0.2 um glass fiber filter prior to analysis. The method required only 3.15 minutes. Table 1 provides optimized transitions and retention times for the PFCs measured in this method.

Chromatography Parameters

| Flow Rate: 0.35 mL/min  | Mobile Phase A: 10 mM ammonium acetate in water          |
|-------------------------|--|
| Column oven: 35°C       | Mobile Phase B: 10 mM ammonium acetate in 80:20 MeOH:ACN |
| Injection volume: 20 µL | Column: Supelco Titan C18 (50 x 2.1mm 1.9 um)            |
|                         | Gradient: Linear 45% - 97% B 0-2.6 minutes               |
|                         |  |

Mass Spectrometry Parameters

| Nebulizing Gas 2.2 L/min | Interface Temperature 230 C       |
|--------------------------|-----------------------------------|
| Heating Gas 8 L/min      | Desolvation Line Temperature 95 C |
| Drying Gas 10 L/min      | Heat Block Temperature 225 C      |
|                          | Column Oven 35 C                  |

#### Table 1: Method information for the perfluorinated compounds measured in surface and drinking water

| Name                                  | Abbreviation | Structure  | Retention Time | Transitions        |
|---------------------------------------|--------------|--|----------------|--------------------|
| Perfluoro-n-hexanoic acid             | PFHxA        |  | 1.28           | 313>119<br>313>269 |
| Perfluoro-n-heptanoic acid            | PFHpA        |  | 1.63           | 363>169<br>363>319 |
| Perfluoro-n-octanoic acid PFOA        |              |  | 1.91           | 413>169<br>413>369 |
| Perfluoro-n-nonanoic acid PFNA        |              |  | 2.15           | 463>219<br>463>419 |
| Perfluoro-n-decanoic acid             | PFDA         | $F \xrightarrow{F} F \xrightarrow{F} F \xrightarrow{F} F \xrightarrow{F} F \xrightarrow{F} F \xrightarrow{F} H \xrightarrow{F} $ | 2.36           | 513>269<br>513>469 |
| K Perfluoro-n-butanesulfonate         | L-PFBS       | F<br>F<br>F<br>F<br>F<br>F<br>F<br>F<br>F<br>F<br>F<br>F<br>F<br>F<br>F<br>SO <sub>3</sub> <sup>-</sup> K <sup>+</sup>   | 1.02           | 299>80<br>299>99   |
| Na Perfluoro-n-hexanesulfonate        | L-PFHxS      | $F \xrightarrow{C} C \xrightarrow{C} C \xrightarrow{C} C \xrightarrow{C} C \xrightarrow{SO_3 \cdot Na^+}$  | 1.70           | 399>80<br>399>99   |
| Na Perfluoro-n-octanesulfonate L-PFOS |              | F = F = F = F = F = F = F = F = F = F =  | 2.18           | 499>80<br>499>99   |



Figure 3: PFC calibration curves demonstrating excellent linearity at low analyte concentration matched with spectra showing the analyte LOQ and a chromatogram with 60 fg on column.

sample preparation.

|     | Lab Blank | Centennial<br>Lake | Liberty<br>Lake | Little   | Middle   | Drinking<br>Water | Table 2 : Calculated amounts o       |
|-----|-----------|--------------------|-----------------|----------|----------|-------------------|--------------------------------------|
|     |           |                    |                 | Patuxent | Patuxent |                   | materials in the surface and dr      |
|     |           |                    |                 | River    | River    |                   | water samples. The measured          |
| кА  | 0.34      | 1.19               | 0.59            | 0.68     | 1.17     | 0.51              | amounts were averaged (n=4)          |
| Ac  | 0.1       | 0.36               | 0.33            | 0.33     | 0.33     | 0.16              | duced by the amount of each a        |
| А   | 0.79      | 1.09               | 0.92            | 1.04     | 1.05     | 0.37              | data at a line that a b Dlawle (100) |
| А   | 0.22      | 0.24               | 0.24            | 0.25     | 0.26     | 0                 | detected in the Lab Blank (18iv      |
| 4   | 0.71      | 0.43               | 0.16            | 0.07     | 0.08     | 0                 | purified water). These values v      |
| BS  | 0         | 0                  | 0               | 0        | 0        | 0                 | then divided by the injection v      |
| HxS | 0         | 0                  | 0               | 0.06     | 0.06     | 0.05              | to determine the final concent       |
| S   | 0         | 0                  | 0.13            | 0.19     | 0.1      | 0                 | ng/L.                                |
|     |           |                    |                 |          | Discus   | ssion             |                                      |