

# Detecting inhibition of nitrogen removal



Handling interferences

Dr. Achim Leitzke, 11.08.2019

**MACHEREY-NAGEL**

[www.mn-net.com](http://www.mn-net.com)



# Agenda

---



**Company**



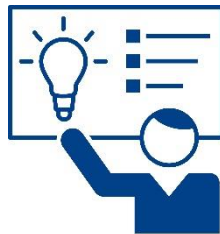
**Nutrient Removal**



**Nitrification-inhibition  
tests**



**Results**



**Case studies**



**Tips and Tricks**



**Summary**



# Company

---

MN Water Analysis

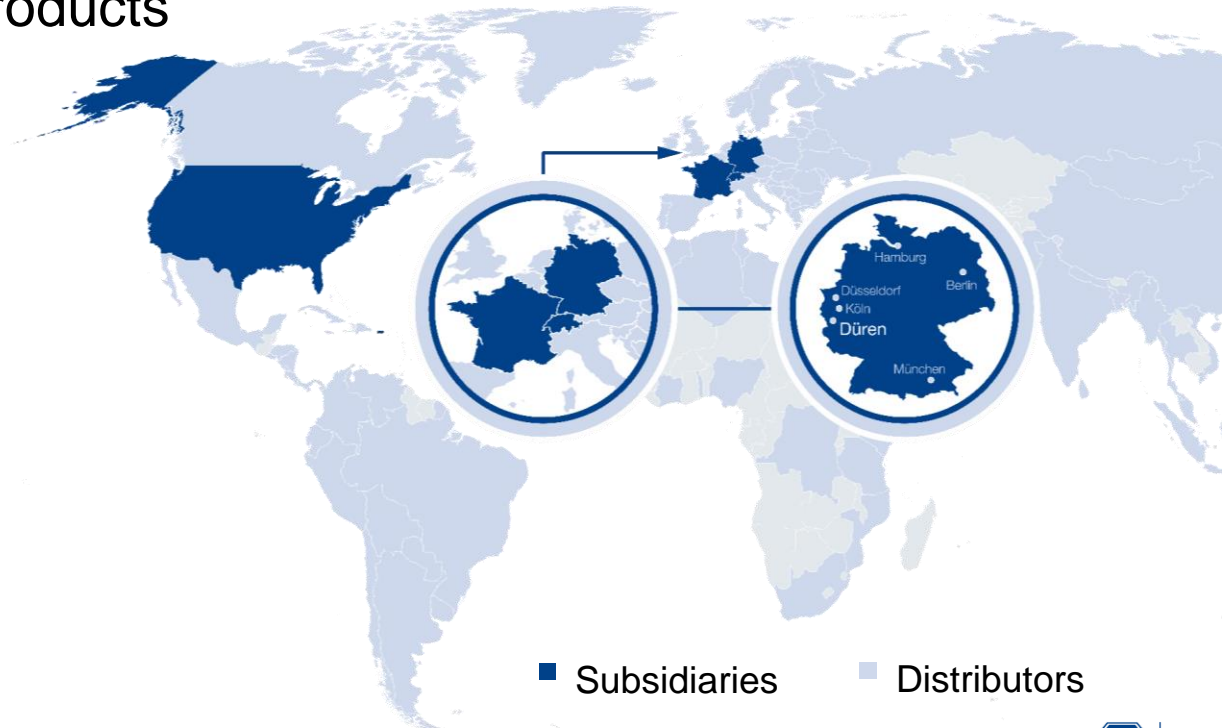




# Company

## MN today

- 4<sup>th</sup> Generation family owned
- More than 700 employees
- More than 25.000 products
- Turnover 120 Mio. €



# Company

---



## Business units

1911



Filtration

1959



Rapid Tests

1961



Chromatography

1970



Water Analysis

1989



Bioanalysis



# Nutrient removal

---





# Nutrient removal

## EPA Fact Sheet

- nitrogen and phosphorus are the primary causes of cultural eutrophication
- approximately 25% of all water body impairments are due to nutrient-related causes
  - oxygen depletion
  - algal growth
  - ammonia
- more stringent effluent limits lead to
  - P-removal
  - N-removal

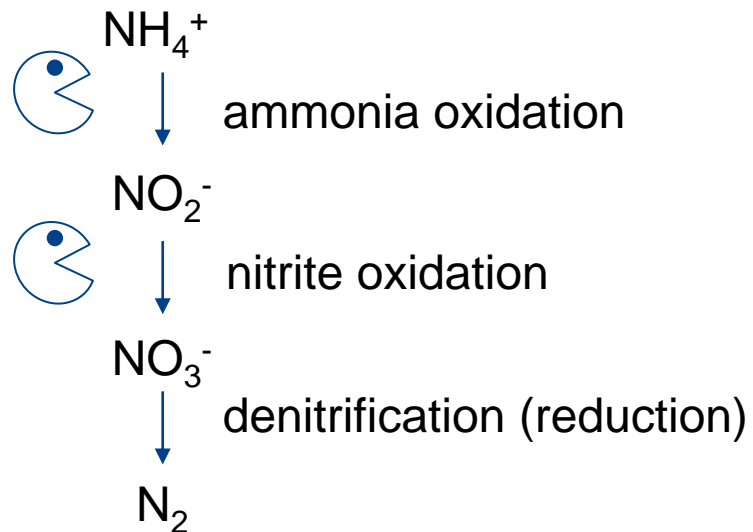




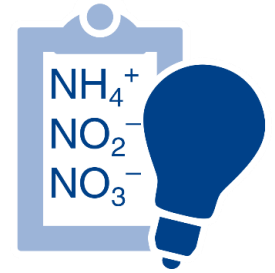


# Nutrient removal

## Denitrification process



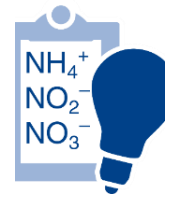




# Nitrification-inhibition tests

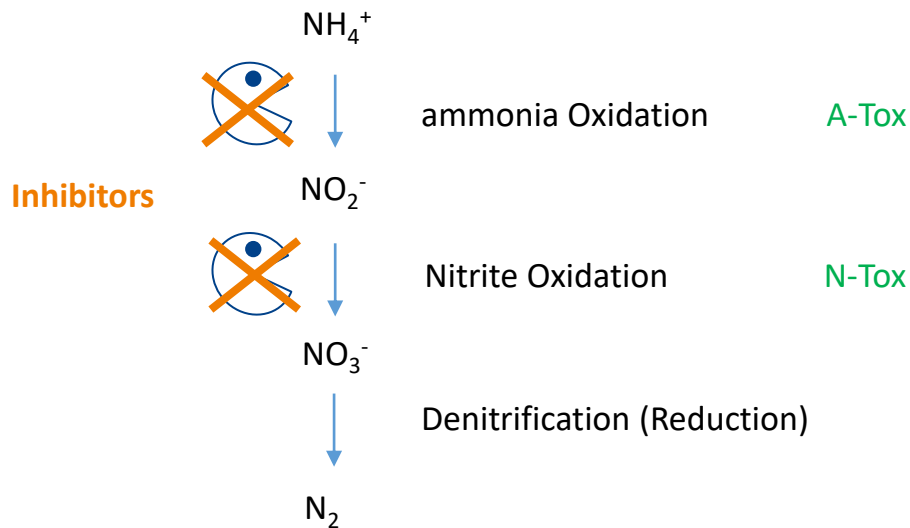
---



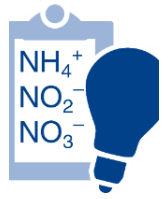


# Nitrification-inhibition tests

## Inhibition



 “(de)nitrification-inhibition” results in increased total N in outflow or down-time of plant

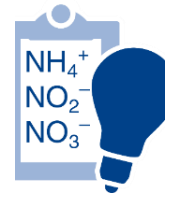


# Nitrification-inhibition tests

- ISO 9509
  - use active sludge
  - add sample
  - incubate 4-24 h
  - measure nitrate and ammonia



! Or...



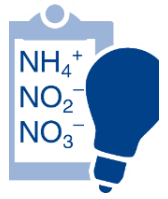
# Nitrification-inhibition tests

## How to detect inhibition?

- use ready-to-use kit
  - lyophilized bacteria
  - includes all necessary solutions...
  - ... and feed for bacteria
- How it works?
  - bacteria oxidize ammonia and nitrite
  - oxygen is consumed
  - oxygen consumption correlates with bacteria activity



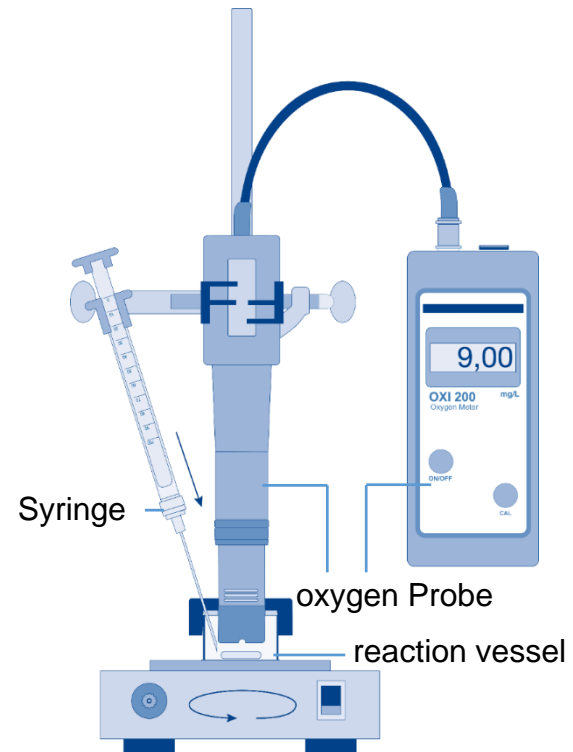
**bacteria inhibition leads to lower Oxygen Consumption**

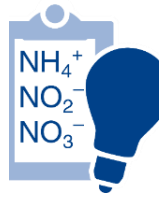


# Nitrification-inhibition tests

## How the test is done?

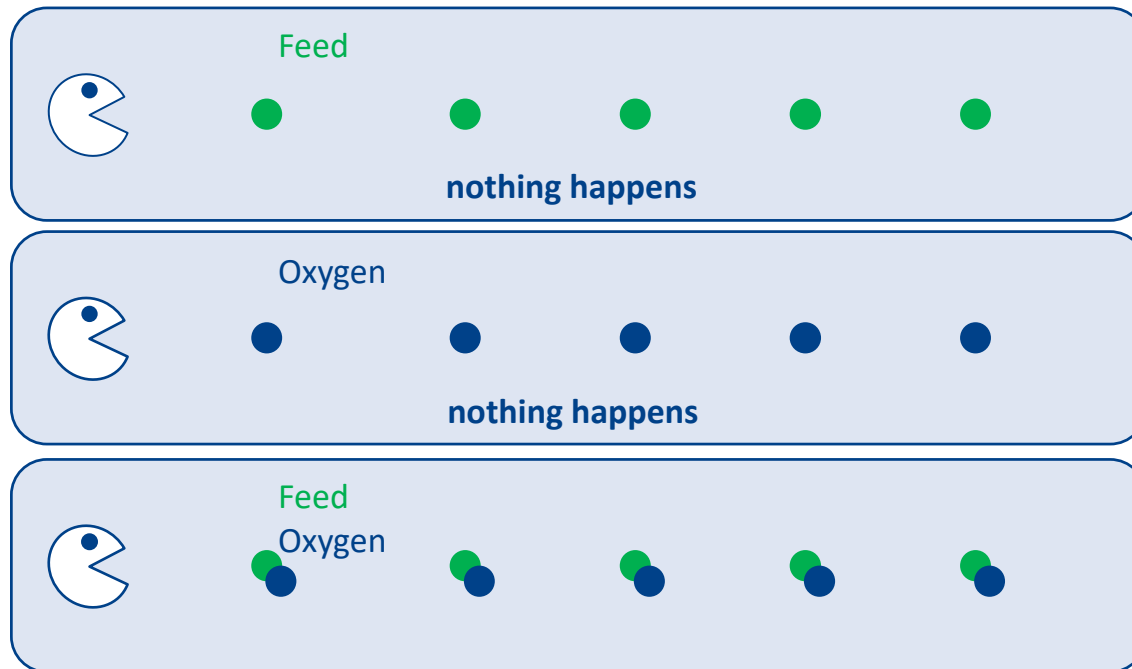
- use oxygen probe
- measure
  - Control
  - Sample
- calculate inhibition from difference

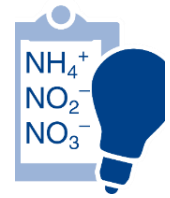




# Nitrification-inhibition tests

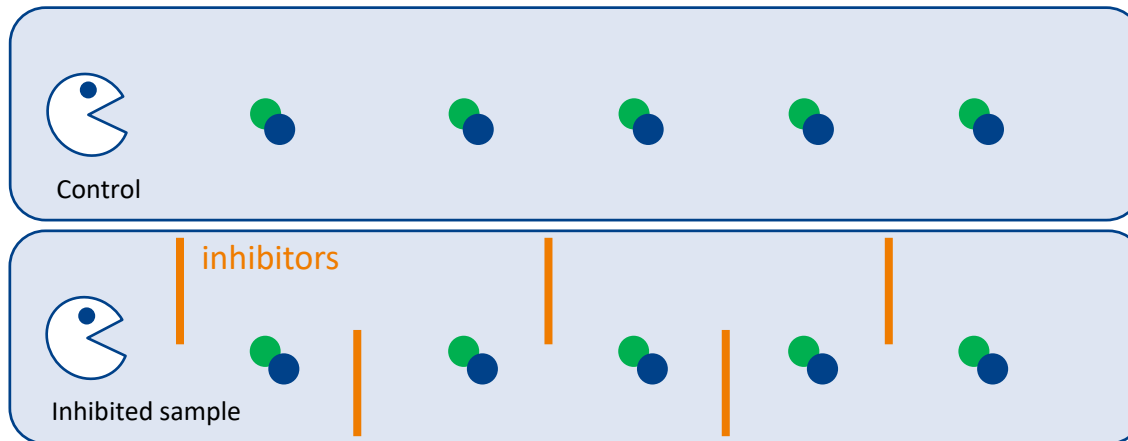
## How the tests work?





# Nitrification-inhibition tests

## Control and sample



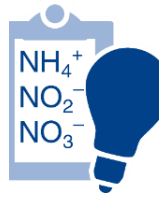
$\Delta O_C : 4 \text{ O}_2$

$$\Delta O_C - \Delta O_S = 1 \text{ O}_2$$

$\Delta O_S : 3 \text{ O}_2$

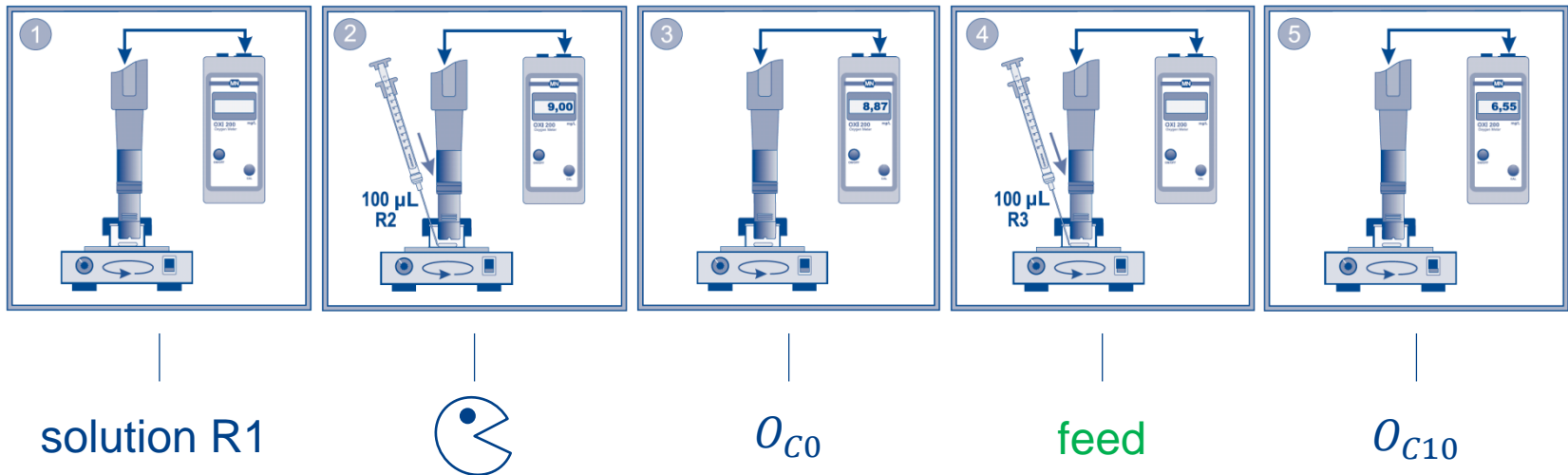
 **inhibition:**  $\frac{\Delta O_C - \Delta O_S}{\Delta O_C} * 100 = \frac{1}{4} * 100 = 25 \%$



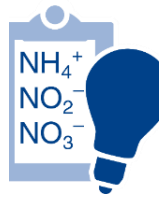


# Nitrification-inhibition tests

## Control

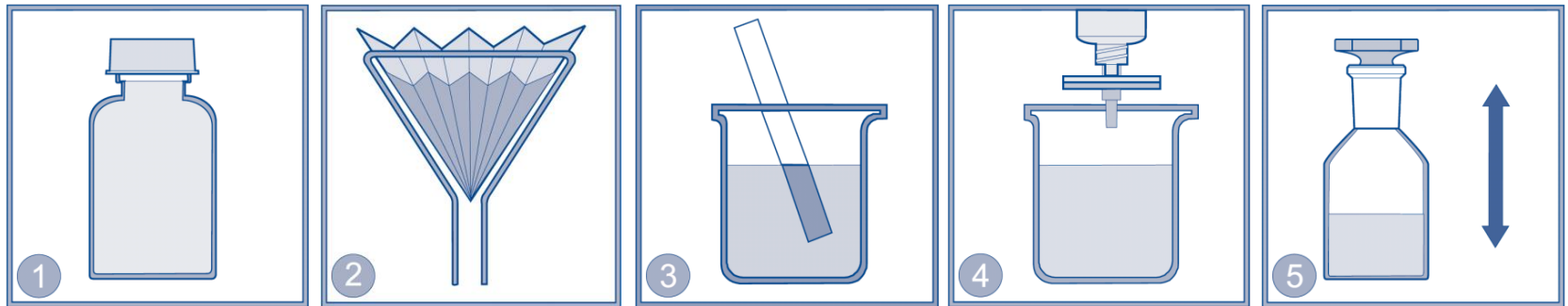


calculate Oxygen Consumption in control as :  $\Delta O_C = O_{C0} - O_{C10}$

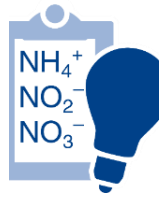


# Nitrification-inhibition tests

## Sample preparation

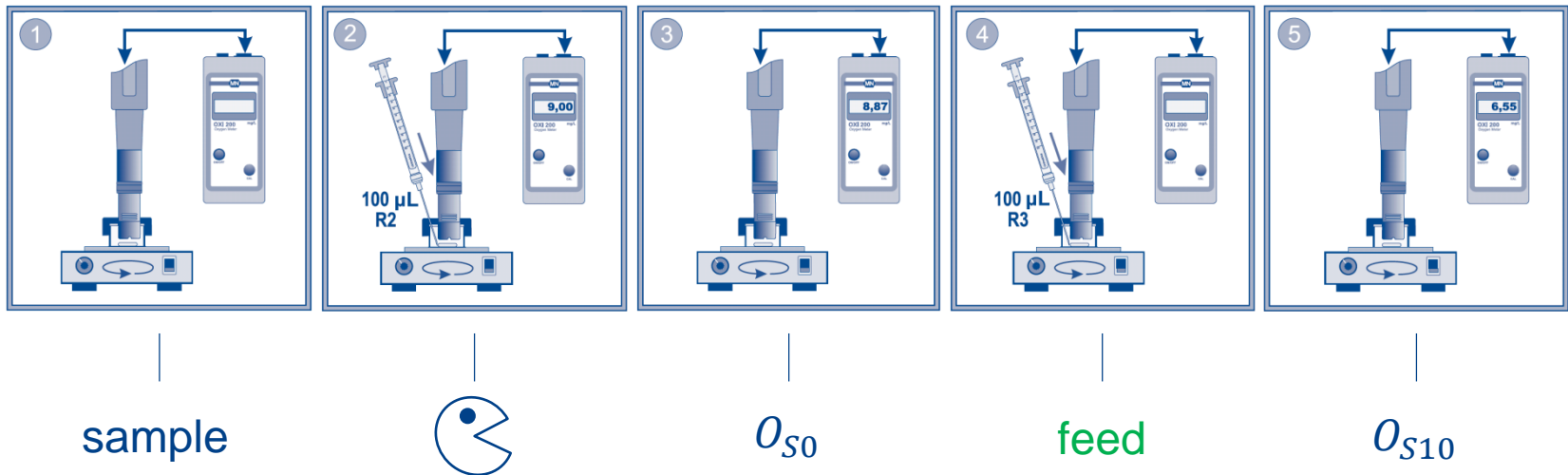


reactivate *A-Tox-Bacteria* by shaking 4-6 time one day before performing test



# Nitrification-inhibition tests

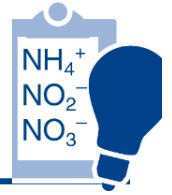
## Sample



calculate Oxygen Consumption in sample as :  $\Delta O_S = O_{S0} - O_{S10}$

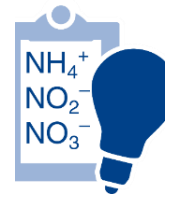
# Nitrification-inhibition tests

---



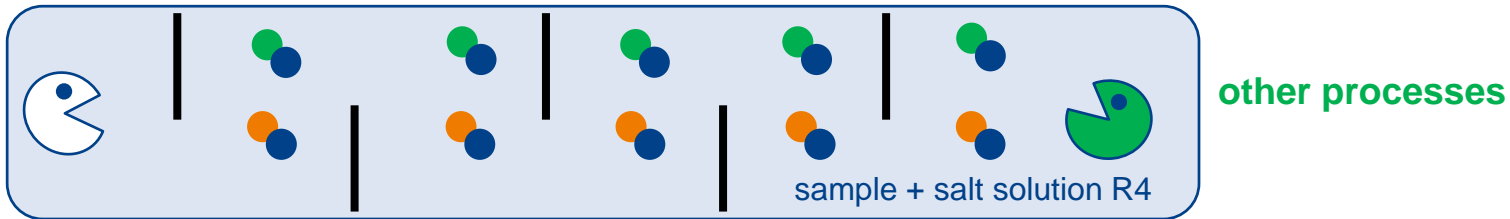
## Calculate inhibition

- oxygen demand Control:  $\Delta O_C$
- oxygen demand Sample:  $\Delta O_S$
- inhibition [%] =  $\frac{\Delta O_S - \Delta O_C}{\Delta O_C} * 100$



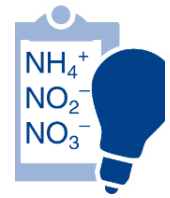
# Nitrification-inhibition tests

Correct for other oxygen consuming processes



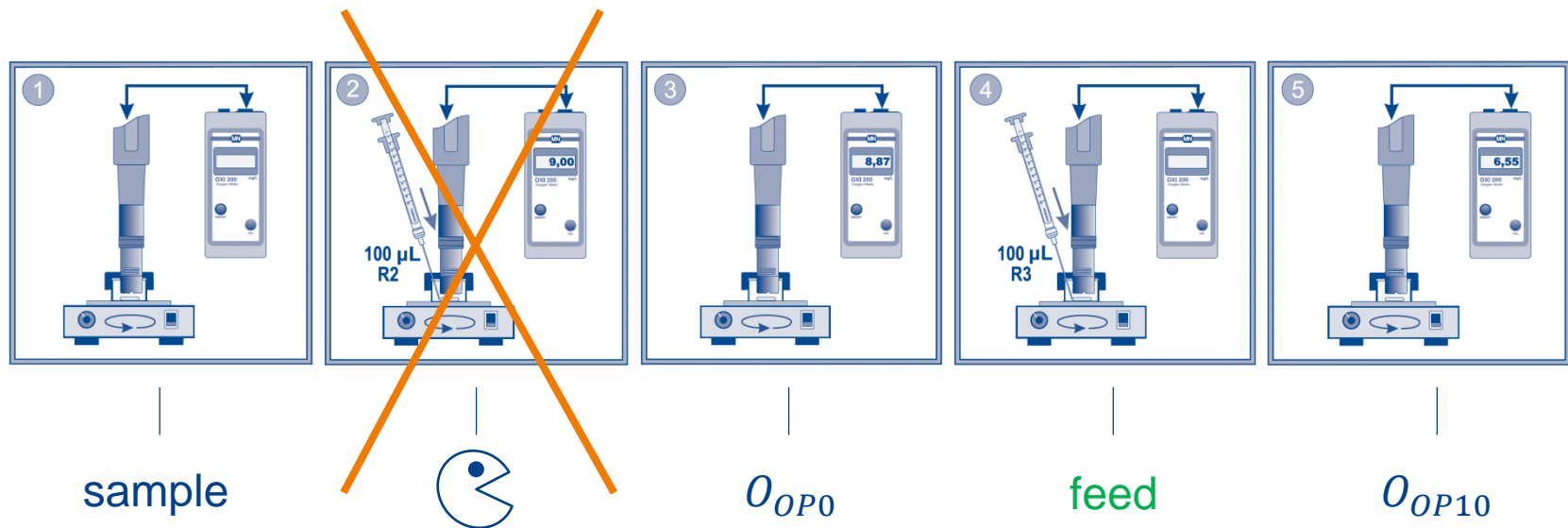
$$\begin{array}{ll} \text{nitrification + other processes} & = 8 \text{ O}_2 \\ \text{other processes alone } (\Delta O_{OP}) & = 5 \text{ O}_2 \end{array}$$

$$\text{corrected consumption } (\Delta O_S) = 3 \text{ O}_2$$



# Nitrification-inhibition tests

## Sample



calculate consumption by other processes as:  $\Delta O_{OP} = O_{OP0} - O_{OP10}$



# Results

---







# Nitrification-inhibition tests

## ISO 9509 vs. BioFix<sup>®</sup> Tox Tests, IC<sub>50</sub> values

| Substance                           | ISO 9509   | A-Tox      | N-Tox     |
|-------------------------------------|------------|------------|-----------|
| <b>Pure Substances and Mixtures</b> |            |            |           |
| N-Allylthiourea                     | 0,5        | 0,2        | 16        |
| Cu-Sulfate                          | 0,02 g/L   | 0,4 g/L    | 0,04 g/L  |
| Palegal SF                          | 0,1 ml/L   | 3 ml/l     | 0,15 ml/l |
| Dishwashing liquid                  | 0,1 ml/l   | 1,6 ml/l   | 0,3 ml/L  |
| <b>Real Samples</b>                 |            |            |           |
| Metall industry                     | 0,004 ml/l | 0,005 ml/l | 0,05 ml/L |
| Tannery                             | 105 ml/l   | >125 ml/l  | 75 ml/l   |
| Food industry                       | 2 ml/l     | 4 ml/l     | 77 ml/l   |



**Tox-tests compare very well to ISO 9509**



# Nitrification-inhibition tests

## IC<sub>50</sub> values for A-Tox / N-Tox

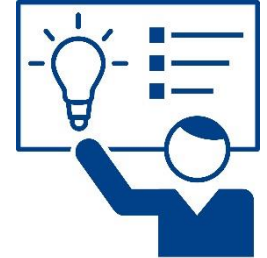
| Classification                                    | Reference  | IC <sub>50</sub> -Values   |  |
|---|--|--|--|
|   |  | BioFix® A-Tox  | BioFix® N-Tox  |
| Nitrification inhibitors in the chemical industry | <i>N</i> -Allylthiourea  | 0,89 mg/L NATU   | 19,8 g/L NATU  |
|   | Thiourea   | 0,99 mg/L TU   | 48,8 g/L TU  |
|   | Potassium chlorate   | 185 g/L KClO <sub>3</sub>  | 42,5 g/L KClO <sub>3</sub>   |
| Nitrification inhibitors in agriculture           | „N-Serve“ / „Nitrpyrine“<br>2-Chloro-6-(trichloromethyl)pyridine | 72 mg/L <i>N</i> -Serve  | 80 mg/L <i>N</i> -Serve  |
|   | „Didin“ (Cyanoguanidine)   | 450 mg/L Cyanoguanidine  | >100 g/L Cyanoguanidine  |
| Chlorophenols                                     | 2-Chlorophenol   | 2,9 mg/L 2-Chlorophenol  | 71 mg/L 2-Chlorophenol   |
|   | 3,5-Dichlorophenol   | 7,1 mg/L 3,5-DCP   | 6,3 mg/L 3,5-DCP   |
| Heavy Metals                                      | Cu <sup>2+</sup> (as copper(II)sulfate x 5 H <sub>2</sub> O)     | 407 mg/L CuSO <sub>4</sub> x 5 H <sub>2</sub> O<br>104 mg/L Cu <sup>2+</sup> | 52 mg/L CuSO <sub>4</sub> x 5 H <sub>2</sub> O<br>13,2 mg/L Cu <sup>2+</sup> |
|   | Cd <sup>2+</sup> (as cadmium chloride x H <sub>2</sub> O)        | 246 mg/L CdCl <sub>2</sub> x H <sub>2</sub> O<br>137 mg/L Cd <sup>2+</sup>   | 190 mg/L CdCl <sub>2</sub> x H <sub>2</sub> O<br>106 mg/L Cd <sup>2+</sup>   |
|   | Zn <sup>2+</sup> (as zink sulfate x 7 H <sub>2</sub> O)          | 1,2 g/L ZnSO <sub>4</sub> x 7 H <sub>2</sub> O<br>0,27 g/L Zn <sup>2+</sup>  | 430 mg/L ZnSO <sub>4</sub> x 7 H <sub>2</sub> O<br>98 mg/L Zn <sup>2+</sup>  |



# Nitrification-inhibition tests

## IC<sub>50</sub> values for A-Tox / N-Tox

| Classification                                    | Reference  | IC <sub>50</sub> -Values               |  |
|---|--|--|--|
|   |  | BioFix® A-Tox                          | BioFix® N-Tox                          |
| Additives in textile industry / textile finishing | Palegal SF   | 3,8 mL/L Palegal SF                    | 0,23 mL/L Palegal SF                   |
|   | Gleyzin A  | 15,3 mL/L Gleyzin A                    | 223 mL/L Gleyzin A                     |
| Disinfectants, Detergents, Preservatives          | Hydrogen peroxide (H <sub>2</sub> O <sub>2</sub> ) | 5,5 mg/L H <sub>2</sub> O <sub>2</sub> | 4,7 mg/L H <sub>2</sub> O <sub>2</sub> |
|   | Benzyldimethyl-dodecylammonia chloride             | 45,2 mg/L                              | 76,2 mg/L                              |
|   | 4-Dodecylbenzenesulfonic acid                      | 0,55 mL/L                              | 0,84 mL/L                              |
|   | Dimethyldioctadecylammonia chloride                | 18,6 mg/L                              | 5,0 g/L                                |
| Solvents  | DMSO   | 4,68%                                  | (> 100%)                               |
|   | Ethanol  | 0,74%                                  | 43,0%                                  |



# Case studies

---





# Case study 1

---

## Indirect discharger in Germany

- Discharge of sludge waters
- Discharger needs to check with BioFix<sup>®</sup> A-Tox
- Inhibitions >20% → Discharge is not permitted
- Waste must be disposed at costs of 35.000 EUR (39.000 USD)
- 1 mg/L NATU is used regularly to check the status of the bacteria



## Case study 2

---

### Sewage plant in Dresden, Germany

- Analysis of waste waters from industrial indirect dischargers
- Food industry, galvanics, chemical industry
- Water of each discharger is analyzed every 4-6 weeks
- Determination of the concentration with non significant inhibition
- Inhibition of >50% is considered as toxic
- One discharger creates a stimulation through a high  $\text{NH}_4$ -load



# Tips and Tricks

---





# Tips and Tricks

---



When doing the test...



**Be careful to treat control and sample identically**

# Tips and Tricks

---



## Recommended Oxygen Level

> 8 mg/L O<sub>2</sub>



increase oxygen level by shaking or air flow through sample

# Tips and Tricks



## Ensure Minimum Oxygen Consumption of Control

- > 1,0 mg/L O<sub>2</sub>
- Depends on fitness of bacteria
- Initial Consumption when shipped:
  - *A-Tox*: > 3,0 mg/L O<sub>2</sub>
  - *N-Tox*: > 1,5 mg/L O<sub>2</sub>



# Tips and Tricks



## Take care of stirring speed

- Optimal 200 – 400 /min
- Use same speed for all tests



# Tips and Tricks



## Storage of bacteria

*A-Tox*



+ 2 to + 8 °C  
+35 to +46 °F

*N-Tox*



-20 ± 2 °C  
- 7 ± 0 °F



**take care of different storage conditions for different parts of the kit**

# Tips and Tricks

---



## Take of reagent temperature prior to use

- Bring all reagents to room temperature before use
- Shake salt solutions well before use



# Tips and Tricks



## Reactivate bacteria



**Shake well before use to reactivate and homogenize**



# Tips and Tricks

---

## Look at reagent color!

Activity correlates with color

- Red-brown: fresh bacteria
- Dark-brown - grey: getting older
  - Still working well
- Black: inactive / dead bacteria
  - Not usable anymore
  - Take care of storage

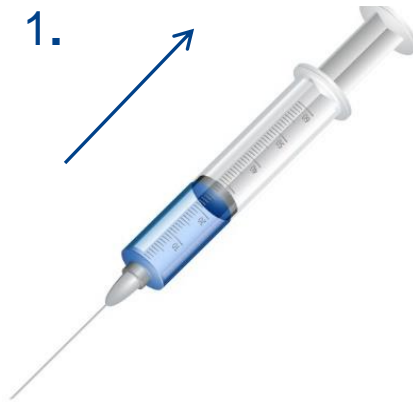


# Tips and Tricks



## Bacteria activity a probelm?

1.



2.



**NO – as long as sample and control are done identical**

# Tips and Tricks

---



## Remove air bubbles

- Take care when assembling
- Take care to carefully wet the adaptor





# Summary

---



# Summary



## Comparison ISO 9509 – BioFix®

|                 | DIN EN ISO 9509                                | BioFix®              |   |
|-----------------|--|----------------------|---|
| Preparation     | Active sludge clean-up                         | Lyophilized bacteria | ✓ |
| Incubation time | 4 h  | 12 min               | ✓ |
| Solutions       | Prepare  | Ready to use         | ✓ |
| Detection       | $\text{NH}_4^+$ / $\text{NO}_3^-$ (any method) | Oxygen probe         | ✓ |



**BioFix® A-Tox/N-Tox – Sold in Europe for more than 15 years**

# Thank you for your attention!

---

Dr. Christian Prokisch | [cprokisch@mn-net.com](mailto:cprokisch@mn-net.com) | +49-2421-969-166

## Image credits

---

© darknightsky (1), SG-design (10), Kaesler Media (11,13), jolopes(14), Vitalinka (26), rcfotostock (27, 28, 35), gertrudda (29), You can more (30), mocka(30), mocka (31) / Fotolia