Rapid, Automated Analysis of Microplastics Using Laser Direct Infrared Imaging and Spectroscopy

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August

"Big Questions" in microplastics (MPs)

- From where did these plastics originate?
 - Have they penetrated the food chain?
- Are they harmful?
 - Have they absorbed harmful compounds?
- How can the impact be mitigated?



Fundamental questions in the routine analysis of microplastics

- Does my sample (drinking/waste water, food, environmental sample) contain microplastics?
- How many?
- What size are they?
- What are their chemical identities?

Best strategy for answering these questions =

active area of research

- From where did these plastics originate?
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Strategies – what do you do once you've filtered the sample?

- Manual, by-eye approach
 - Inaccurate (unaided by analytical technique)
 - Time-consuming
 - Unlikely to determine chemical identity
- Wet chemistry & related methods
 - Physically intensive and time-consuming
 - Difficult for small or individual particles
 - Destructive particle is gone, no opportunity for further analysis
- Optical micro-spectroscopy methods
 - Chemically-specific (get particle identities)
 - Non-destructive (leave the door open to further analysis)
 - Can be highly automated







What is optical micro-spectroscopy?



It's a microscope:

- Visible images of particles
- Particle count
- Particle size



spectrophotometer:

 Determine chemical identity of a chemical compound by measuring interaction with infrared light

Single combined platform

- MP counting, sizing, identification in one technology
- Non-destructive: can perform subsequent analysis
- Potential for automation or MP-specific workflows

Agilent 8700 LDIR Chemical Imaging System

Routine, robust, automated microplastics analysis by non-experts



- Fully-automated infrared microscopy platform
- Utilizes a new laser light source
 - Fast, crisp infrared images
 - Laser-focused spectroscopy ideal for small particles
- Removes many of the 'chores' associated with traditional infrared microscopy
 - No liquid nitrogen, autofocus, auto collection of background data, etc.

IR Micro-Spectroscopy – what does it actually do?

A material's *response* (absorbance) to a wide range of infrared wavelengths produces a spectrum (signature) unique to each compound.

Thus, IR spectroscopy can be used for chemical identification!



IR Micro-Spectroscopy – what does it actually do?

A traditional IR micro-spectroscopy instrument spreads light over an area.

The good:

• Look at multiple particles simultaneously

The bad:

- Weak signals, slow analysis
- Measures 'empty space' between MPs
- Significant data processing
- >30 seconds per spectrum typical



Only a tiny fraction of the light can be absorbed.

Solution: use a laser!

Laser Direct Infrared (LDIR) Spectroscopy

New instrument architecture

- Bright, focused infrared laser source
- Full automation provides a tailored particle-specific workflow
- Intuitive software and operation reduces user influence and enhances consistency

Other advantages

- One second per spectrum
- Targeted analysis near-zero time measuring empty space
- Real-time data processing and feedback



All light reaches the particle





Workflow

- Insert the sample
- Select an on-board microplastic library
- Draw a box over an area of interest
- Software acquires an image, automatically identifies particle locations
- User may specify a size range (ex. exclude very large or small particles)
- Software automatically takes a spectrum of each particle, performs a library match, continually updates statistics of all particles!

LDIR imaging provides excellent contrast for identifying particle count, locations, sizes



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Training set samples – polycarbonate microplastics

- 41 identified particles over 25mm²
- Size range: 220µm 25µm
- 90% matched as polycarbonate
- Of the others:
 - One was truly polyamide (dust)
 - One was polyester fiber (possibly from clothing)



Training set samples – polypropylene microplastics

- 112 identified particles over 283mm²
- Size range: 495µm 25µm
- 90% matched as polypropylene
- Of the others:
 - Several were truly polyamide (dust)
 - Other contaminants



Testing set samples – mixed microplastics

- Blend of 7 known components
- 72 identified particles over 126mm²
- Size range: 435µm 25µm
- 94% match with known components in the blend
- Some particles were organic contaminants (cellulose-based)







Real-world environmental sample Effluent from wastewater treatment plant

Sewage is used as fertilizer - route for microplastic contamination

Sample preparation protocol¹

- Filtration and enzymatic digestion (protease, cellulase, chitinase, H₂O₂) to remove organic debris
- Density separation to remove inorganics like sand

Retained particles are suspended and aliquoted onto a slide for analysis.

¹ Löder *et al.*, Enzymatic Purification of Microplastics in Environmental Samples. Environmental Science & Technology 2017 51 (24), 14283-14292



Sample courtesy of Dr. Jeff Prevatt, Deputy Director, Treatment Division, Pima County Regional Wastewater Reclamation Dept., Tucson, AZ

LDIR Analysis of Wastewater Sample



5 mm × 5 mm analyzed 1028 particles found

2.5 hours total time

- 8 seconds per particle
- Actual spectrum time 1 second
- All automation, data processing overhead included in this figure
- No operator required for bulk of analysis

Wastewater particle identification (1028 particles)



Polyamides were most prevalent

- Wool, silk, nylon from textiles
- Undigested protein-rich debris

Followed by cellulosic particles, PTFE (Teflon), polystyrene



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Recently announced; shipping later this year.

Meanwhile...

- Improving speed
- Pushing to even smaller particles
- Enhancing statistical analysis and reporting
- Validating on real-world samples from diverse sites in external labs

Workflow still in R&D—we need your feedback! <u>chris moon@agilent.com</u>

Thank you!



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